Are There Any Epigenetic Similarities Between Treatment Unresponsive Sarcoidosis, COPD and Severe Asthma?

Esmaeil Mortaz^{1,2,3}, Alireza Eslaminejad³, Mohammad Varahram⁴, Jalal Heshmatnia³, Atefeh Abedini³, Arda Kiani³, Aliakbar Velayati⁴, Johan Garssen², and Ian M. Adcock⁵

¹ Department of Immunology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran ² Division of Pharmacology and Pathophysiology Utrecht Institute for Pharmaceutical Sciences, Faculty of Sciences, Utrecht University, Utrecht, The Netherlands

³ Chronic Respiratory Diseases Research Center and National Research Institute of Tuberculosis and Lung Diseases

(NRITLD), Masih Daneshvari Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁴ Department of Infectious Diseases, Mycobacteriology Research Center, National Research Institute

of Tuberculosis and Lung Diseases (NRITLD), Masih Daneshvari Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁵ Airways Disease Section, National Heart and Lung Institute, Imperial College London, London, UK

Received: 12 January 2015; Received in revised form: 5 March 2015; Accepted: 5 April 2015

Sarcoidosis is characterized by non-caseating granulomas and several immunological abnormalities in many tissues including the lungs (pulmonary) and others such as skin, bone, heart (extra pulmonary)¹. The aetiology of the disease is unknown although probably relates to an inflammatory/immune response to an unknown infectious agent.1 This leads to tissue damage, remodeling of airways, airway hyperactivity and a resultant loss of lung function. Corticosteroids remain the mainstay of first line treatment in sarcoidosis although they are not effective in all patients.^{2,3}.Recent evidence suggests that epigenetic mechanisms are involved in the control of inflammation and immune cell function in cancer¹ and in the molecular pathways implicated in other pulmonary disorders such as chronic obstructive lung disease (COPD), severe asthma and interstitial lung disease (IPF).⁴ These diseases are all associated with epithelial and mesenchymal cell remodelling within the airways and alveoli associated with altered patterns

Corresponding Author: Ian M. Adcock, PhD;

Airways Disease Section, National heart & Lung Institute, Imperial College London, Dovehouse Street, London SW3 6LY, UK. Tel: (+44 0207) 594 7840, E-mail: ian.adcock@imperial.ac.uk

of growth factor activity and expression; apoptosis; increased oxidative and endoplasmic reticulum stress; altered cellular senescence along with impaired mucociliary clearance and host defense processes in response to environmental agents such as pollution, cigarette smoke or allergens in the case of asthma.⁵⁻⁷ It is also evident that corticosteroid functions are under the regulation of epigenetic processes.⁸

A range of epigenetic processes such as histone modifications, non-coding RNAs and DNA methylation are associated with the control of gene expression.^{9,10} The development and differentiation of most cell types including T cells ¹¹ are reliant upon these mechanisms for efficient tissue- and cell-specific expression of genes. These epigenetic mechanisms do not act independently of each other but act in a coordinate manner to regulate the induction and sustained expression of the myriad of epigenetic tags or marks that control gene expression.

The deposition of acetylated histone marks by histone acetyltransferases (HATs) is associated with enhanced expression of immune and inflammatory genes.^{7,12,13} Recent evidence indicates that deposition of acetyl tags by HATs is not highly selective whereas removal of these tags by histone deacetylases

Copyright© Autumn 2015, Iran J Allergy Asthma Immunol. All rights reserved.

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

(HDACs), of which there are 18, is more selective.¹⁴ Addition of an acetyl group alters the structure of the local chromatin generally allowing enhanced gene expression⁷ which is reversed by the action of HDACs.¹²

An imbalance in HAT/HDAC activity is reported in the airways of patients with severe asthma ¹⁵ and in COPD¹⁶ and linked to the reduction in corticosteroid responsiveness in these two diseases¹⁷. In particular a selective loss of HDAC2 has been implicated in preventing corticosteroid suppression of a number of key inflammatory and immune genes.¹⁸⁻²¹ This action is due to both a reduced ability of the activated glucocorticoid receptor (GR) to remove lysing tags from histones at inflammatory gene promoters and due to changes in GR acetylation status preventing interaction with then nuclear factor $\kappa appa B$ (NF- κB) p65.¹⁸ In addition to GR, many other transcription factors such as NF-KB p65 and p53, transcription coregulators including PGC1, RB and c-Myc, inflammatory signaling pathways such as mitogen activated protein kinases (MAPKs), DNA repair proteins such as Ku 70 and the structural protein βactin can be acetylated. This acetylation can markedly affect protein and cellular function.^{22,23} Complex interactions between acetylated non-histone proteins and HDACs occur, for example, activation of the acetylated transcription factor hypoxia-inducible factoroccurs 1α (HIF-1α) which in the lung microenvironment of patients with COPD, decreases HDAC2 expression, resulting in augmented inflammation and steroid resistance.24

However, overall little data exists from primary cells/tissues regarding altered histone modifications in this disease. The effects of cigarette smoke on primary human airway epithelial cells which causes corticosteroid insensitivity ⁵ indicates some changes such as Histone H4K16 and H3K27 acetylation and H3K27 and DNA methylation could be examined preferentially in these patients and linked to gene and protein expression profiles.²⁵

There are few published links between the above epigenetic modifications or the expression of the various enzymes involved in depositing or removing these marks with corticosteroid function. This remains an area of intense interest. In contrast, in fibroblasts from patients with idiopathic pulmonary fibrosis (IPF), for example, there have been several studies linking changes in histone methylation and acetylation with alterations in their regulatory enzymes and the control of key genes including cyclooxygenase-2 (COX-2). In addition, the function of transforming growth factor beta (TGF- β) has been associated with several microRNAs (miRNAs) such as miR-218, miR-21, miR-155, miR-20 and let-7d which are differentially expressed in fibroblasts from IPF subjects ²⁵. Environmental stresses induce alterations in DNA methylation in COPD patients ²² and these can be mimicked in cells exposed to cigarette smoke which suggests that similar changes should be observed in sarcoidosis.²⁵

Indeed, the accelerated telomere shortening seen in sarcoidosis has been linked to earlier evidence of subtelomerichypomethylation.²⁶ In addition, sarcoid patients also demonstrate higher levels of histone H4 in bronchial alveolar lavage (BAL) and histone H2B in plasma compared to healthy controls.²⁷ Furthermore, there were suggestions that BAL histone H4 proteins were post-translationally modified although this needs to be confirmed. More studies are required in this direction to explore the epigenetic mechanisms underlying sarcoidosis epigenetics in individual response to corticosteroids.

Inflammation in severe asthma, COPD and IPF has also been associated with altered expression of microRNAs and with alterations in DNA and histone methylation⁷. Similar to control of acetylation, methylation at specific residues on histone H3 for example is carefully controlled by the relative activities of histone methyltransferases (HMT)/histone demethylases (HDM) ^{7,28,29} and it is now clear that DNA methylation is also highly regulated.³⁰

Again, there is an interaction between these processes since altered DNA methylation of the miR-17~92 cluster promoter results in the over expression of genes linked to fibroblast proliferation.³¹

Several studies have reported alterations in microRNA expression profiles in severe asthma and COPD³² including changes in miR-19, -21, -27, -29a, -126 and -146a and some of these have been associated with corticosteroid function including miR-145.³³

Limited data exists regarding epigenetic processes in sarcoidosisal though miR-92b and miR-206 expression is elevated in both the lung and lymph nodes of sarcoidosis patients. In contrast, miR-20a and miR-302c expression was elevated in lymph nodes but decreased in lung.³⁴

In conclusion, there is increasing evidence for

^{473/} Iran J Allergy Asthma Immunol, Autumn 2015

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

similarities, as well as differences, in epigenetic marks associated with sarcoidosis and with the pathogenesis of other chronic inflammatory airway diseases. Further analysis of epigenetic changes associated with corticosteroid function should be addressed in future. Ideally, blood-based analysis should be performed and methods to allow mathematical deconvolution of the data to enable links to single cell types have been developed.³⁵ Identification of common epigenetic marks between these diseases or with a lack of corticosteroid responsiveness in association with gene expression data will allow determination of key regulatory modules and delineation of new therapeutic targets^{10,36} which is critical for the welfare of these patients.

ACKNOWLEDGEMENTS

Ian M Adcock is supported by the MRC (G1001367/1) and the Welcome Trust (093080/Z/10/Z). Research by IMA is also supported by the NIHR Respiratory Disease Biomedical Research Unit at the Royal Brompton NHS Foundation Trust and Imperial College London. Esmaeil Mortaz is supported by an ERS Long Term Fellowship (LTRF 2013-2052).

REFERENCES

- Mortaz E, Sereshki HA, Abedini A, Kiani A, Mirsaeidi M, Soroush D, et al. Alternation of serum TNF-α, IL-8 and free light chain with HLA-DR B alleles expression in pulmonary and extra-pulmonary sarcoidosis. J Inflamm (Lond) 2015; 12:21.
- Paramothayan S, Jones PW. Corticosteroid therapy in pulmonary sarcoidosis:a systematic review. JAMA 2002; 287(10):1301–7.
- Grutters JC, van den Bosch JM. Corticosteroid treatment in sarcoidosis. Eur Respir J 2006; 28:627–36.
- Liu Y, Li H, Xiao T, Lu Q. Epigenetics in Immune-Mediated Pulmonary Diseases. Clin Rev Allergy Immunol 2013; 45(3):314–30.
- Barnes PJ. Corticosteroid resistance in patients with asthma and chronic obstructive pulmonary disease. J Allergy Clin Immunol 2013; 131(3):636-45.
- 6. Yang IV, Schwartz DA. Epigenetics of idiopathic pulmonary fibrosis.Transl Res 2015; 165(1):48-60.
- Mortaz E, Masjedi MR, Barnes PJ, Adcock IM. Epigenetics and Chromatin Remodeling Play a Role in Lung Disease. Tanaffos 2011; 10(4):7–16.

- Barnes PJ. Corticosteroid resistance in patients with asthma and chronic obstructive pulmonary disease. J Allergy ClinImmunol 2013; 131(3):636-4.
- Tripathi SK, Lahesmaa R. Transcriptional and epigenetic regulation of T-helper lineage specification. Immunol Rev 2014; 261(1):62–83.
- Moncef Zouali, The Epigenetics of Autoimmune Diseases.472 pages April 2009. WILEY-BLACKWELL press 2009.
- Kitagawa Y, Ohkura N, Sakaguchi S. Molecular determinants of regulatory T celldevelopment: the essential roles of epigenetic changes. Front Immunol 2013; 10(4):106.
- Strahl BD, Allis CD. The language of covalent histone modifications. Nature 2000; 403(6765):41–5.
- Filippakopoulos P, Knapp S. Targeting bromodomains: epigenetic readersoflysine acetylation. Nat Rev Drug Discov 2014; 13(5):337-56.
- Feller C, Forné I, Imhof A, Becker PB. Global and specific responses of thehistoneacetylome to systematic perturbation. Mol Cell 2015; 57(3):559-71.
- Su RC, Becker AB, Kozyrskyj AL, Hayglass KT. Altered epigenetic regulation and increasing severity of bronchial hyperresponsiveness in atopic asthmaticchildren. J Allergy ClinImmunol 2009; 124(5):1116-8.
- Ito K, Ito M, Elliott WM, Cosio B, Caramori G, Kon OM, et al. Decreased histone deacetylase activity in chronic obstructive pulmonary disease. N Engl J Med 2005; 352(19):1967-76.
- Adcock IM. Histone deacetylase inhibitors as novel antiinflammatory agents. Curr Opin Investig Drugs 2006; 7(11):966–73.
- Ito K, Yamamura S, Essilfie-Quaye S, Cosio B, Ito M, Barnes PJ, et al. Histone deacetylase 2-mediated deacetylation of the glucocorticoid receptor enables NFkappaB suppression. J Exp Med 2006; 203(1):7–13.
- Kobayashi Y, Bossley C, Gupta A, Akashi K, Tsartsali L, Mercado N, et al. Passive smoking impairs histone deacetylase-2 in children with severe asthma. Chest 2014; 145(2):305-12.
- To Y, Elliott WM, Ito M, et al. Total histone deacetylase activity decreases with increasing clinical stage of COPD. Am J Respir Crit Care Med 2004; 169:A276.
- Barnes PJ. Reduced histone deacetylase in COPD: clinical implications. Chest 2006; 129(1):151–5.
- 22. Farria A, Li W, Dent SY. KATs in cancer: functions and therapies. Oncogene 2015; doi:10.1038/onc.2014.453.
- 23. Philp A, Rowland T, Perez-Schindler J, Schenk S. Understanding the acetylome: translating targeted proteomics into meaningful physiology. Am J Physiol

Iran J Allergy Asthma Immunol, Autumn 2015/474

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

Cell Physiol 2014; 307(9):C763-73.

- Charron CE, Chou PC, Coutts DJ, Kumar V, To M, Akashi K, et al. Hypoxia-inducible factor lalpha induces corticosteroid-insensitive inflammation via reduction of histone deacetylase-2 transcription. J Biol Chem 2009; 284(52):36047–54.
- 25. Yang IV, Schwartz DA. Epigenetics of idiopathic pulmonary fibrosis. Transl Res 2015; 165(1):48-60.
- Maeda T, Guan JZ, Higuchi Y, Oyama J, Makino N. Agingrelated alterations of subtelomeric methylation in sarcoidosis patients. J Gerontol A BiolSci Med Sci 2009; 64(7):752–60.
- 27. Teirilä L, Karvala K, Ahonen N, Riska H, Pietinalho A, Tuominen P, et al. Proteomic Changes of Alveolar Lining Fluid in Illnesses Associated with Exposure to Inhaled Non-Infectious Microbial Particles. PLoS One 2014; 9(7):e102624.
- Kouzarides T. Histone methylation in transcriptional control. CurrOpin Genet Dev 2002; 12:198–209.
- Rea S, Eisenhaber F, O'Carroll D, Strahl BD, Sun ZW, Schmid M, et al. Regulation of chromatin structure by site-specific histone H3 methyltransferases. Nature 2000; 406(6796):593–9.
- Dawson MA, Kouzarides T. Cancer epigenetics: from mechanism to therapy. Cell 2012; 150(1):12-27.

- 31. Dakhlallah D, Batte K, Wang Y, Cantemir-Stone CZ, Yan P, Nuovo G, et al. Epigenetic regulation of miR-17~92 contributes to the pathogenesis of pulmonary fibrosis. Am J RespirCrit Care Med 2013; 187(4):397-405.
- 32. Booton R, Lindsay MA. Emerging role of MicroRNAs and long noncoding RNAs in respiratory disease. Chest 2014; 146(1):193-204.
- 33. Collison A, Mattes J, Plank M, Foster PS. Inhibition of house dustmite-induced allergic airways disease by antagonism of microRNA-145 is comparableto glucocorticoid treatment. J Allergy Clin Immunol 2011; 128(1):160-7.
- 34. Crouser ED¹, Julian MW, Crawford M, Shao G, Yu L, Planck SR, Rosenbaum JT, Patrick Nana-Sinkam S. Differential expression of microRNA and predicted targets in pulmonary sarcoidosis. Biochem Biophys Res Commun 2012; 417(2):886-91.
- 35. Liang L, Willis-Owen SA, Laprise C, Wong KC, Davies GA, Hudson TJ, et al. An epigenome-wide association study of total serum immunoglobulin E concentration. Nature 2015; 520(7549):670-4.
- 36. Gustafsson M, Nestor CE, Zhang H, Barabási AL, Baranzini S, Brunak S, et al. Modules, networks and systems medicine for understanding disease and aiding diagnosis. Genome Med 2014; 6(10):82.