REVIEW ARTICLE Iran J Allergy Asthma Immunol September 2012; 11(3): 203-216

The Immunopathogenic Role of Reactive Oxygen Species in Alzheimer Disease

Monireh Mohsenzadegan and Abbas Mirshafiey

Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

Received: 11 November 2011; Received in revised form: 11 March 2012; Accepted: 22 March 2012

ABSTRACT

Reactive oxygen species (ROS) are produced in many normal and abnormal processes in humans, including atheroma, asthma, joint diseases, cancer, and aging. Basal levels of ROS production in cells could be related to several physiological functions including cell proliferation, apoptosis and homeostasis.

However, excessive ROS production above basal levels would impair and oxidize DNA, lipids, sugars and proteins and consequently result in dysfunction of these molecules within cells and finally cell death. A leading theory of the cause of aging indicates that free radical damage and oxidative stress play a major role in the pathogenesis of Alzheimer disease (AD). Because the brain utilizes 20% more oxygen than other tissues that also undergo mitochondrial respiration, the potential for ROS exposure increases.

In fact, AD has been demonstrated to be highly associated with cellular oxidative stress, including augmentation of protein oxidation, protein nitration, glycoloxidation and lipid peroxidation as well as accumulation of Amyloid β (Aβ). The treatment with anti-oxidant compounds can provide protection against oxidative stress and Aβ toxicity.

In this review, our aim was to clarify the role of ROS in pathogenesis of AD and will discuss therapeutic efficacy of some antioxidants studies in recent years in this disease.

Keywords: Alzheimer disease; Reactive oxygen species

INTRODUCTION

A number of studies have been performed for seeking a correlation between Alzheimer disease (AD), inflammation, and oxidative stress and or nitrosative stress.¹ During physiological aging, the emergence of some neurodegenerative related aging diseases like AD with damaged mitochondria of brain cells are unable to maintain the energy required for the cells.²

Thus, the reduced energy for metabolism in AD may be due to dysfunction of some key metabolic enzymes of mitochondria.³ Moreover, neurons are particularly sensitive to oxidative stress as compared to other organs or tissues; therefore the brain is more vulnerable to reactive oxygen species (ROS)-induced damage due to its high rate of oxygen consumption, high polyunsaturated lipid content, and relative weariness of classic antioxidant enzymes.

In fact, oxidative stress and Aβ production are positively correlated to each other. When the brain attempts to repair itself from oxidative damage, which is characterized by over expression of Amyloid β (Aβ)

Corresponding Author: Abbas Mirshafiey, PhD; Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran-14155, Box: 6446, Iran. Fax: (+98 21) 6646 2267, E-mail: mirshafiey@tums.ac.ir

Copyright© 2012, IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY. All rights reserved. 203 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

precursor protein (AβPP), it initiates the formation and accumulation of Aβ that appears to be its end product.4,5 This process can lead to an increased production of free radicals mainly superoxide anion by mitochondria which induces the interruption of oxidative phosphorylation and results in decreased levels of ATP molecules. ROS are potentially toxic for neurons and oligodendrocytes and may induce toxicity by damaging lipids, proteins and nucleic acids of cells and mitochondria.⁶

Therefore, mitochondrial dysfunction results from molecular defects in oxidative phosphorylation (OXPHOS), which most likely plays a key role as a component for the development and maturation of AD.⁷ Moreover, the increased ROS level triggers the opening of mitochondrial permeability transition pore and inner membrane anion channel and a transient increase in ROS generation by the electron transport chain. The involvement of mitochondrial permeability transition pore (mPTP) is implicated in Aβ-induced mitochondrial dysfunction, such as perturbation of intracellular calcium regulation, ROS generation, release of pro-apoptotic factors and impairing mitochondrial morphology.

The ROS burst in the cytosol can activate ROSinduced ROS release in neighboring mitochondria, leading to potentially significant mitochondrial and cellular injuries.⁸ Furthermore, studies indicate that $A\beta$ oligomers can impair mitochondrial function via ROS production and further increase ROS levels. Oxidative stress in brain could also stimulate additional damage via the overexpression of inducible nitric oxide synthase (iNOS) and the action of constitutive neuronal NOS (nNOS) which increases the production of nitric oxide (NO) and its derivative (reactive nitrogen species). $9,10$

The oxidative stress for a long time has been speculated to play a major role as a cause and consequence of AD. Thus, attenuation and/or suppression of oxidative stress have been evaluated as an alternative therapeutical choice for AD treatment.

ROS

ROS are formed continuously in cells as a consequence of oxidative biochemical reactions along with internal and external factors. In general, there are more than six primary sources of free radicals formed endogenously within living organisms: 1. the respiratory generation of ATP using $oxygen;^{11}$ 2. peroxisomal oxidation of fatty acids, which generates H_2O_2 ¹² 3. cytochrome P450 enzymes;¹¹ 4. chronic inflammatory cells, which use a mixture of oxidants to overcome infection by phagocytosis; 12,13 5.other enzymes which are capable to generate oxidants under normal or pathological conditions.¹⁴

Oxygen-derived free radicals are highly reactive chemical species involved in a variety of diseases, including neurodegenerative disorders. Superoxide anion (O2), hydroxyl radical (OH) and hydrogen peroxide (H_2O_2) , known as reactive oxygen species are produced by the reduction of molecular oxygen to water in mitochondria by some amino oxidasecatalyzed reactions and during the activation of phagocytic NADPH oxidase.^{15,16} ROS including superoxide, hydrogen peroxide, and hydroxyl radicals, and their reactive products which are classically described as harmful products of aerobic metabolism are capable of DNA mutations, lipid peroxidation, and protein oxidation, microglial proteosome malfunction, astrocyte activation, inflammation and cell death.¹⁷

The main role of ROS function in immune system is to form an integral part of the organism's defense against invading microbial agents. The formed ROS by NADPH oxidase in phagocytic cells are mainly used to kill invading pathogens and therefore, the lack of functional NADPH oxidase complex results in low resistance to bacterial and fungal infections in humans and chronic granulomatous disease.^{15,18} In addition to its role in killing process and host defense, a large amount of evidence has shown the important roles of ROS in cell proliferation, apoptosis, homeostasis, intracellular signaling, angiogenesis, endocrine-related functions, and oxidative modification of the extracellular matrix.¹⁵ For example, autophagy which is a lysosome-dependent catabolic process mediating turnover of cellular components plays an important role in regulating cellular homeostasis in the nervous system. Because the accumulation of misfolded proteins is a common feature in multiple human neurodegenerative diseases, thus the activation of autophagy can be proposed as a strategy for combating neurodegeneration.^{19,20} The increase in the levels of ROS is a frequent consequence of the accumulation of misfolded proteins and expired organelles such as old mitochondria. Moreover, ROS may serve as an important intracellular signal for the homeostatic activation of autophagy under basal physiological conditions, as well as in neurodegenerative diseases

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

²⁰⁴/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

including AD. The studies indicate that autophagy is specifically up-regulated in AD, due to ROS-dependent activation of the type III PI3 kinase. Indeed it is presented as an acute and long-term attempt by the affected neuronal cells to rid themselves of the harmful effects of Aβ exposure, such as accumulation of defective mitochondrial and protein aggregates. 21 However, if ROS are produced in excess in certain abnormal conditions, such as inflammation and ischemia or in the presence of catalytic iron ions, they could be harmful to cells. Since neurons have an agerelated decrease in the capacity to compensate for redox imbalance, even minor cellular stress has also the ability to lead to irreversible injury and, it can contribute to the pathogenesis of neurodegenerative diseases.²²

ROS have been also identified as important second messenger molecules that carry out a part of signaling steps transduced by pro-inflammatory cytokines. The vascular endothelium, which regulates the passage of macromolecules and circulating cells from blood to tissue, is a major target of oxidant stress in increased vascular endothelial permeability as well as promoted leukocyte adhesions that are involved in alterations in endothelial signal transduction and redox-regulated transcription factors. Therefore, oxidative stress plays a critical role in the pathophysiology of vascular diseases.5,23 ROS signals could be the important factors during lymphocyte transendothelial migration. Interestingly, ROS from NADPH oxidase have been shown to mediate Aβ- induced cerebrovascular dysfunction. 24 The evidence supports this fact that NADPH oxidase may be a common pathway moreover for microglia-mediated neuronal damage. It was shown that ROS production is stimulated by lymphocyte binding to the adhesion molecule, vascular cell adhesion molecule-1 (VCAM-1). The VCAM-1 stimulates endothelial cell NADPH oxidase for the production of low levels of ROS $(1 \mu M H_2O_2)$ and this is required for VCAM-1-dependent lymphocyte migration.^{25,26}

Moreover, there is profound evidence that the pathogenesis of several neurodegenerative diseases, including Parkinson's disease, Friedreich's ataxia, multiple sclerosis, amyotrophic lateral sclerosis and AD may due to the generation of reactive nitrogen species (RNS) associated with mitochondrial dysfunction.²⁷ In MS, macrophages produce a variety of inflammatory mediators like nitric oxide (NO), and proinflammatory cytokines, which all contribute to neuroinflammation and disease progression.²⁸ Furthermore, Peroxynitrites (ONOO⁻)) ,another component of oxidative stress, formed from NO and O_2 is a highly reactive oxidizing and nitrating agent, leading to oxidize cellular components, including proteins, lipids, carbohydrates, and DNA and increased aggregated Aβ , and stimulate inflammatory response, so that ONOO⁻ scavenging and ROS inhibitory effects could be considered as potential anti-AD candidates.^{1,29}

ROS in AD

In the central nervous system (CNS), the high metabolic demand for oxygen can lead to a higher level of oxidative stress via the production of free radicals. Since the extent of ROS formation is associated with oxygen consumption, so that the higher level of ROS is produced by neurons with higher metabolic activity or neuronal segments enriched in mitochondria, such as synapses.³⁰ Loss of mitochondrial membrane can increase the release of cytochrome c, a pathway leading to neuronal apoptosis that is mediated by the Bcl-2 family of proteins. 31 Under pathological conditions such as AD, oxidative stress can enhance the progression of the disease. Partial deficiency of the mitochondrial manganese superoxide dismutase (MnSOD) increased amyloid plaques in Tg19959 mice and tau phosphorylation in Tg2576 mice. $32,33$ In addition, the over expression of MnSOD reduced amyloid plaques, improved memory function and protected synapses. A major link between oxidative stress and mitochondrial dysfunction is the α ketoglutarate dehydrogenase complex (α-KGDHC). In human AD brains, the mitochondrial α -ketoglutarate dehydrogenase activity is markedly reduced in either damaged or relatively undamaged areas^{34,35} α -KGDHC is a crucial mitochondrial enzyme complex that mediates oxidative metabolism. Its activity is reduced in AD patient brain.³²

Furthermore, depending on whether AβPP is cleaved via the amyloidogenic or the nonamyloidogenic process, AβPP fragments themselves are known to induce oxidative stress or act as neuroprotective. 36 In fact, alterations in the lipid composition of cellular membranes and/or in membrane fluidity induced by a permanently increased oxidative microenvironment pave the way to change many different metabolic processes taking place in distinct membrane compartments. Moreover, the higher

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /205 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

plasma membrane fluidity due to methyl-β-cyclodextrininduced cholesterol depletion has been shown to promote α-secretase cleavage of AβPP and, following, the nonamyloidogenic form.³⁷ The increased lanosterol levels (cholesterol precursor) correlate to enhanced αsecretase activity in oxidative-stress-resistant cells. In contrast, cholesterol accumulation in the plasma membrane has been reported to increase the rigidity of the plasma membrane and to decrease α-secretase processing of APP.³⁸

The Role of ROS in Signaling Pathway in AD

 Epidemiological studies have supported the role of inflammation in AD, where results have shown a decreased incidence and severity of AD in patient populations treated with nonsteroidal anti-inflammatory drugs (NSAIDs). The studies are shown that patients taking anti-inflammatory medicine for rheumatoid arthritis are six times less likely to develop AD.³⁹ Moreover, NADPH oxidase initiates an intracellular ROS signaling pathway that can activate microglia and amplify the production of multiple pro-inflammatory cytokines, such as TNF α or PGE2.^{40,41} In fact, several triggers of NADPH oxidase activation in microglia amplify proinflammatory signaling. The microglial activation occurs early in AD development, before neuropil damage, supporting a contributing role of microglia in disease pathology.⁴² It is thought that interaction of microglia with Aβ peptide gives rise to ROS and several other chemokines and cytokines, which work as inflammatory mediators and finally may cause neuronal damage, so that, TNF-α, nitric oxide, and superoxide are produced by microglia in response to $AP₃₉$ Furthermore, activation of complement cascade forms membrane attack complexes, which not only cause substantial damage to the neurons but also can lead to phosphorylation of Tau protein leading to formation of neurofibrillary tangles.⁴³ A β has also been shown to recruit and activate microglia, suggesting a critical role in AD progression.⁴⁴ It was also found that the level of nuclear factor kappa B (NFkB) in the brain is significantly increased in the presence of APOE e4 when compared with its activiation in the presence of APOE e3. Several molecules are capable of activating NFkB including TNFα, Aβ, and secreted AβPP. It should be noted that the activation of NFkB increases transcription of AβPP and BACE-1, which finally leads to increase in Aβ production. $42,45,46$

Moreover, Aβ-induced toxicity appears to involve one or more of the three major mitogen-activated protein kinase (MAPK) pathways, c-jun N-terminal kinase (JNK), p38, and extracellular signal regulated kinase (ERK), which are known to mediate oxidative stressinduced neuronal death.⁴⁷ Moreover, many oxidative stress factors have been shown to trigger apoptosis by stimulating stress-activated protein kinases (SAPKs) such as JNK and $p38MAPK$.¹⁰ In AD model cells, it was shown that the amyloid precursor protein swedish (double mutation at amino acids 670 and 671found in a swedish family is located before the Aβ region of AβPP and result in increased Aβ) enhanced oxidative stress, finally leading to apoptotic cell death through the activation of the c-Jun N-terminal kinase, caspases 3 and 9 and a shift in the BclxL/Bax ratio toward Bax.^{48,49} Protein phosphatase 5 (PP5) is a ubiquitously expressed serine/threonine phosphatase related to PP1, PP2A, and PP2B.⁴⁷ The PP5 is suggested to inhibit MAPK pathways through dephosphorylation of Raf-1, a MAPK kinase initiating the ERK MAPK pathway, and apoptosis signal regulating kinase 1 (ASK1), which activates the JNK and p38 MAPK pathways.^{50,51} The results suggest that understanding the function and regulation of PP5 in brain can provide insight into neuronal responses to Aβ, as well as potential therapeutic strategies for the prevention of Aβ-induced neurodegeneration.

Aβ is the product of amyloid precursor protein cleaved by β and γ secretase. A β generates oxidizing products during its aggregation.⁵² The oxidizing products as well as Aβ affect the functions of sodium-potassium ATPase and calcium ATPase, 53 which in turn cause dysregulation of L type voltage sensitive calcium channel $(LVSCC)$ ⁵⁴, , α-amino-3-hydroxy-5 methylisoxazole-4-propionic acid receptor (AMPAR),⁵⁵ and N-methyl D-aspartate receptor (NMDAR)⁵⁶ and inositol 1,4,5-trisphosphate receptors^{52,57} that then mediate significantly increased calcium flux into the cytosol. Aβ-mediated abnormalities in mitochondrial function are rescued by adding the mitochondrial permeability transition pore (mPTP) inhibitor. Indeed, cyclosporine D (CypD) plays a key role in stabilizing mitochondrial permeability transition (mPT) and serves to open the mPTP, thereby allowing the diffusion of calcium and cytochrome c out of mitochondria matrix into the cytoplasm where they may induce cell death via necrosis and/or apoptosis. There is strong protective effect of CypD deficiency on Aβ-mediated mitochondrial and neuronal toxicity. Thus, CypD

206/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

inhibition could be a feasible and possibly significant therapeutic approach. The inhibitor of CypD, Cyclosporine A, broadly was used in a clinical application, which might be a potential therapeutic option for the treatment of AD.⁵²

On the other hand, both Aβ- and glutamate-induced toxicities in neurons are largely dependent on cyclindependent kinase $(Cdk5)^8$. However, the role of CdK5 in promoting oxidative stress in AD has not completly been elucidated. Cdk5, a serine-threonine kinase, belongs to the Cdk family. It was shown that Cdk5 directly causes excessive oxidative stress and mitochondrial dysfunction in neurons leading to cell death, the downstream of Aβ and glutamate, both of which play key roles in AD pathology. Two antioxidant enzymes, Prx-I and Prx-II which belong to the Prx family of peroxidases can efficiently scavenge cellular ROS.58,59 Identification of Prx-I and Prx-II as Cdk5 substrates suggested that Cdk5 deregulation may maintain sustained oxidative stress in AD by compromising the cellular anti-oxidant defense system. Furthermore, Cdk5 deregulation increases oxidative stress via inactivation of two antioxidant enzymes, Prx1 and Prx2, which lead to ROS-mediated JNK and c-Jun activation. In neuronal cells, JNK is preferentially activated by oxidative stress and is a key mediator of Aβ, glutamate induced neurotoxicity and Neurofibrillary Tangles (NFT) formation which are critical in AD.60,61 Recent studies have further identified a vital role for JNK in neurotoxic Aβ formation by activating γ-secretase upon oxidative stress. Uncontrolled Cdk5 activates c-Jun via ROSmediated activation of JNK. Thus, Cdk5 inhibition endows superior protection against neurotoxicity, suggesting that Cdk5 could be a preferable therapeutic target for AD .^{61,62}

In addition, recent studies showed that oxidative stress induced by H_2O_2 and 4-hydroxy-nonenal (HNE) treatments activate the feedback between the γ- and βsecretase cleavages of the β-amyloid precursor protein, leading to an increase in Aβ40 and Aβ42 production as well as Aβ42/40 ratios. This kind of feedback requires the activation of the JNK/c-jun pathway. 63 On the other side, the MAPK pathway which play a role in AD pathology, was found that after treatment of N2a/APP695 and N2a/APPswe cells with H_2O_2 , exhibited an enhanced protein expression level of ERK and JNK activation compared to control cells that were not treated with H2O2.10,64

The Role of Metal Ions Associated with ROS in AD

The *in vitro* and *in vivo* studies indicate the important role of metal ions in AD.⁶⁵ Metal-mediated ROS generation is one of the leading causes of oxidative stress and formation of ROS at the metal site as well as the reactivity of the "metallo-ROS" center in CuAβ which may contribute to the oxidative stress in AD ⁶⁶. Interestingly, iron, zinc and aluminum tend to co-localize with Aβ42 peptides in the senile plaque cores that characterize AD brain.⁶⁷ Excessive Cu and Fe ions binding to Aβ were suggested to have a deleterious effect on promoting both the aggregation of Aβ and the generation of ROS. A large body of evidence shows that Cu ion is bound to Aβ in AD and Cu (I/II)– $\mathbf{A}\beta$ complex is involved in ROS (H₂O₂, OH) production. Although *in vitro* studies demonstrated a particular role for Cu (I/II)–Aβ complex in ROS production, the pro-oxidant role of the $Cu(I/II)-A\beta$ systems is controversial.^{65,68,69} The oxidation of A β peptide has two different properties, a protective role, Aβ acting as a sacrificial scavenger of the ROS produced and/ or a toxic role, Aβ being as an initiator of ROS propagation. In the former case, residues involved in the Cu coordination are most affected whereas in the latter case, Tyr10 and/or Met35 in Aβ peptide are affected.^{70,71} It has been shown that \overrightarrow{AB} directly generates ROS in the presence of iron or copper ions via methionine- $35³²$ Two electrons of Met oxidations leading to Met sulfoxide can also be a way of trapping free radicals and thus be neuroprotective, with reduced A β peptide aggregation property.^{72,73} It has been suggested that Met35 in Aβ1–40 serve, as a reducing agent responsible for the initiation of the redox cycling of the CuII center in CuAβ1–40 which can lead to H_2O_2 production.⁶⁶ The oxidation of the thioether moiety of Met to its sulfoxide form in Aβ1– 40 is involved in aggregation, lipid peroxidation and a redox reaction in association with the metal center. Despite the lack of Met and/or any redox-active amino acid, the fragments CuAβ1–16 and CuAβ1–20 exhibit a significant metal-centered oxidative activity which indicates the redox role of Met35 might have been overstretched.74,75

Moreover, cultured human neural cells (HN) of the central nervous system are highly sensitive to nanomolar amounts of aluminum, a known, environmentally abundant neurotoxin that is ubiquitous in biosphere. 67 The genotoxic and neurotoxic effects of aluminum are due to the excessive aluminum mediated

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /207 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

cellular generation of ROS and activation of pathogenic gene expression that redirects brain cells toward genetic dysfunction, neural cell atrophy, apoptosis and cell death. Treatment of HN cells with 100 nM of aluminum-sulfate can emulate many of the gene expression changes observed in the brains of moderateto-late stage AD.76-78

Oxidative Stress and Apolipoprotein E

Apolipoproteins act as antioxidants, however, apolipoprotein E4 allele is less effective.¹² It is reported that polymorphisms of the ApoE gene correlate with onset and risk of developing AD, thus 50% of AD patients have at least one ApoE4 allele that is a major genetic risk factor of the more common late onset form of AD.79,80 APOE gene has three common alleles, epsilon 2 (ε2), epsilon 3 (ε3), and epsilon 4 (ε4). The ε 2 allele is considered as a protective factor but presence of the ε4 could be a risk factor for developing late onset AD, and this allele increases the risk for AD from 20% to 90% and decreases the age of onset from 84 to 68 years depending on gene dose of ε 4 alleles.⁴² ApoE is a 34-kDa glycoprotein that is synthesized and secreted mainly by astrocytes and microglia in the CNS. Moreover, increased oxidative damage is found in specific brain regions of AD patients with the ApoE4 genotype.⁸¹ The detrimental processes of ApoE4 have been shown to influence on AD pathological processes, including lipid homeostasis and NFT formation, suggesting the brain vascular alternations play a key role in the progression of AD. If the delivery of lipophilic antioxidants is impaired due to ApoE4, this could lead to oxidative stress.^{5,82,83}

Furthermore, the relationship between hypercholesterolemia and AD arose in great extent from ApoE4, a major carrier of cholesterol in the CNS. The studies demonstrate that dietary cholesterol increases Aβ accumulation and accelerates AD-related pathology in animals.⁸⁴ The diet-induced hypercholesterolemia in AD mice leads to a significantly elevated levels of formic acid-extractable Aβ peptides in CNS. The total level of Aβ is strongly correlated with the level of cholesterol in both the plasma and CNS.⁵

Antioxidants

The oxidative stress induced by Aβ could be triggered through a number of reactions including increased ROS production, decreased endogenous antioxidant defenses due to glutathione peroxidase (GPx) and superoxide dismutase activities and decrease in the levels of non-enzymatic antioxidants such as reduced glutathione (GSH), vitamin E and ascorbic acid. 85

Moreover, One of other endogenous antioxidants is heat-shock proteins (HSPs) which could be up-regulated in several neurodegenerative diseasess. HSPs can also protect brain cells against free radical injury and oxidative stress. 86 In the CNS, HSP synthesis is induced after hyperthermia, by alterations in intracellular redox environment and by exposure to heavy metals, amino acid analogs and/or cytotoxic drugs. In AD, HSP expression is associated with Aβ deposition and neurofibrillary tangles. Recent findings suggest that HSPs prevent the accumulation of $\text{A}\beta$.^{87,88} HSP 27, among all HSPs, is strongly induced after stresses such as oxidative stress, anticancer drugs, or irradiation.⁸⁶ High level of HSP 27 might be the cause of the occurrence of hyperphosphorylated tau protein (tau proteins are microtubule-associated proteins that are abundant in neurons in the central nervous system) and consequent formation of NFTs. Many investigations showed that HSP 27 directly binds to hyperphosphorylated tau, thereby protecting against cell death.⁸⁹ Thus, the alterations in chaperone protein systems might be a mechanism in pathogenesis and progression of AD, so that a therapeutic approach to induce HSP levels could be a potential strategy to treat or delay the onset of AD.

In general, substances that can reduce oxidative stress are proposed as potential drug candidates for treatment or preventative therapy of neurodegenerative diseases such as AD. In addition, the studies have shown that high antioxidant in diets may decrease the risk of developing $AD.³⁹$

In this review, we will discuss the therapeutic efficacy of substances or drugs which have antioxidant property and might be effective in treatment of AD.

Alkaloids

As mentioned above, protection and inhibition against oxidative stress such as ONOO plays an important role in the production of anti-AD agents. ONOO⁻, formed from NO and O_2 ⁻, is a highly reactive oxidizing and nitrating agent, leading to oxidize cellular components, increased aggregated Aβ, and stimulated inflammatory response.^{1,90} Studies on both cholinesterases (ChEs) and BACE1 (β secretase) inhibitory effects, as well as antioxidant effects, including ONOO- scavenging and ROS inhibitory

208/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

effects of coptidis rhizoma alkaloids, could be considered as promising anti-AD agents. Protoberberine alkaloids such as groenlandicine and jateorrhizine exhibited potent ONOO scavenging effects compared to a well known ONOO scavenger, penicillamine (in a dose-dependent manner).¹

Moreover, mitochondrial antioxidant therapy is a promising treatment for AD patients. Brain, an organ with high energy metabolism and abundance of oxidizable materials is exceedingly susceptible to oxidative damage. $91,92$ Epidemiological evidence oxidative damage. $91,92$ Epidemiological evidence demonstrated that nicotine has beneficial and protective property in some neurodegenerative diseases including AD. The complex I respiratory chain which generates superoxide anion and nicotine is able to inhibit ROS generation on rat brain mitochondria. Nicotine binds to complex I of the respiratory chain and inhibits the NADH-ubiquinone reductase activity.⁹³ Furthermore, nicotine prevents activation of NF-κB and c-Myc by inhibiting the activation of MAP kinases. In fact, nicotine decreases Aβ by the activation of a7nAChRs through MAPK, NF-κB, and c-myc pathways. Nicotinic cholinergic receptor stimulation also induces neuroprotection against glutamate cytotoxicity by its inhibitory action on NO-formation and consequently, the activity of iNOS and the production of NO are down-regulated.⁹⁴

On the other hand, galantamine is a tertiary alkaloid originating from botanical sources, in addition to its neuroprotective property, galantamine prevents ROS production and lipoperoxidation induced by $A\beta$ 1–40, suggesting its antioxidant action. Recently, it was also reported that galantamine prevented ROS production and mitochondrial dysfunction induced by H_2O_2 in a neuroblastoma cell line.⁹⁵

Therefore, alkaloids may be accounted as exogenous drug agents for AD antioxidant therapy. However, the precise and detailed mechanism of function of these alkaloids remains unknown.

NSAIDs

The epidemiological studies has shown that taking NSAIDs for at least one month is associated with lower probability of AD.³⁹ The aggregated synthetic A β 1-40 peptides can induce COX-2 expression in SH-SY5Y neuroblastoma cells, since Aβ1-40 has been shown to stimulate COX-2 oxygenase and peroxidase activity in a cell free system. Furthermore, the two step oxygenase and peroxidase action of COX leads to the formation of

ROS and prostaglandin H_2 , $96,97$ Targets of ROS include activation of COX-1 and-2, which could be blocked by NSAIDs. Daily doses of NSAIDs can increase circulating levels of antioxidants. Furthermore, COX-2 does play an important role in the oxidative stress in AD that may be controlled using selective COX-2 inhibitors like rofecoxib. It has been shown rofecoxib may reduce the free radical load in the rat brain with chronic administration. Flurbiprofen, another NSAID that possesses analgesic and antipyretic properties may also interact with the anti-oxidant system in the AD model rat brain to disrupt the normal oxy-radical (antioxidant balance in the brain).^{96,98}

Moreover, dextromethorphan (DXM) which has anti-inflammatory effects and is a noncompetitive Nmethyl-d-aspartate (NMDA) receptor antagonist exerts its neuroprotective effects through inhibition of microglial activation and NADPH oxidase activation.⁹⁹ In addition, DXM, has been reported in studies of *in vitro* and *in vivo* models of Parkinson's disease to protect patients against neuron damage through the inhibition of microglial activation in methamphetamine-induced neurotoxicity.^{100,101}

Thus, NSAIDs have protective role in AD and COX-2 inhibitor of NSAIDs is also an important target for reducing AD-related oxidative stress.

Polyphenols

To date, one of the most important aspects of current polyphenol research is the focus on the neuroprotective capacity of this broad family of compounds.¹⁰² Polyphenols are a class of phytoalexins found in the tissues of a widespread range of plants. Resveratrol (3–4′-5-trihydroxy stilbene), curcumin, quercetin, and (−)-epigallocatechin-3- gallate (EGCG) of polyphenolic compounds have antioxidant property. The polyphenols have the capacity to chelate metal ions and to directly quench free radical species that contribute to oxidative damage.¹⁰³

Resveratrol was reported to reduce Aβ production in cell line HEK293 expressing Swedish mutant APP695.¹⁰⁴ Additionally, studies indicate the involvement of resveratrol in proteosome clearance of Aβ and reducing toxicity in AD brains. No reduction in the activity of γ-secretase mediated-cleavages of AβPP in the presence of resveratrol was found.¹⁰⁵ Thus, it excludes the possibility that resveratrol lowers Aβ by promoting the proteosomal degradation of C99 (C terminal fragment of AβPP upon cleavage by BACE).

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /209 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

Several studies have investigated curcumin's antioxidant and anti-inflammatory properties. Curcumin enhances the activity of detoxifying enzymes like glutathione-S-transferase.⁴² Low concentrations of curcumin upregulate endothelial heme oxygenase 1 gene and protein expression Heme oxygenase 1 can indirectly protect endothelial cells from peroxide mediated toxicity by degradation heme to iron and biliverdin which later converts to bilirubin and bilirubin then protects endothelial cells from oxidative damage.¹⁰⁶ Curcumin and one of its stable metabolites tetrahydrocurcumin (THC) can significantly decrease production of both iNOS protein and mRNA in transgenic mice brain. In addition, metal chelation activity of curcumin with binding to copper and iron ions reduces Aβ plaque and subsequent ROS generation.¹⁰⁷

Recent investigations have shown that oral administration of green tea polyphenols to mice induces prostate cancer and decrease the production of NF-κB among other regulatory molecules.¹⁰² Green tea polyphenols such as EGCG were capable of reducing the inflammatory markers, cyclooxygenase-2 and prostaglandin-E2, which were associated with development of tumor skin.¹⁰⁸ Moreover, EGCG may be involved in the downregulation of NO production in 4T1 murine mammary carcinoma cells under *in vitro* conditions.¹⁰⁹

In addition to antioxidant capacity of resveratrol and EGCG, these compounds also can regulate the cytotoxic effects of Aβ oligomers and fibrils via phosphorylation of phosphokinase C (PKC) and activation of α-secretase protein. α-secretase catalyzes the formation of a soluble, non-amyloidogenic (non transmembrane plaque-forming) protein from the AβPP which is specifically located in the membrane of neuronal cells. Via this pathway, soluble AβPP is formed and thus does not allow for the formation of neuritic plaques, a hallmark feature of AD.^{102,110}

Quercetin, another component of polyphenol family, is believed to show pharmacological properties to address diseases and risk-factors associated with aging.¹¹¹ The beneficial effect of quercetin against the development of atherosclerosis is inhibition of cell adhesion molecules such as ICAM-1 and VCAM-1 induced TNF- α , however, the prevalent quercetin conjugates were not able to downregulate the proinflammatory ICAMs and VCAMs produced by human umbilical artery smooth muscle cells exposed to either TNF- α or LPS. This finding indicates the need to encourage the focus of research efforts to the metabolic alterations of polyphenolic compounds against various risk factors of aging. 102

Statins

Statins can increase cerebrovascular perfusion by upregulating eNOS that might decrease the inflammation related to $\mathbf{A}\mathbf{\beta}$ deposition.⁴² The main mechanism of action of statins is based on the inhibition of 3- hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase and limitating of the enzyme in the biosynthesis of cholesterol. In addition to, cholesterol-dependent mechanisms of action, statins also directly up-regulate endothelial nitric oxide synthase (eNOS, also termed NOS3 or NOS III) expression, independent of cholesterol levels.¹⁵ Statins could also alter isoprenoid levels. Inhibition of isoprenylation (the process by which isoprenoids are transferred in to target proteins) resulting in the inhibition of small GTPases Ras and Rho activation, both of which are critical for iNOS transcription.¹¹² Furthermore, statins could inhibit endotoxin induced activation of the IκB/NF-κB pathway in a small GTPase-dependent/ independent manner. In the primary astrocytes and macrophages, lovastatin was also found to reduce IFNγ-induced STAT1 phosphorylation and iNOS expression.¹¹³ Thus, the anti-inflammatory effects of statins may have clinical impact in a number of non-vascular conditions including multiple sclerosis, rheumatoid arthritis and decreased inflammations in Aβ depositions.

It has been shown that simvastatin and lovastatin reduce intracellular and extracellular levels of Aβ 42 and Aβ 40 in primary cultures of hippocampal and mixed cortical neurons. 114 In addition, guinea pigs treated with high-dose simvastatin showed a reduction in cerebral A β levels, including the A β 42 isoform.¹¹⁵ Moreover, lovastatin can reduce the production of components of the senile plaques in AD. Lovastatin is able to reduce cellular formation of $A\beta$ in living hippocampal neurons by 70%, and this effect was reversed by the re-addition of cholesterol to previously depleted cells.¹¹⁶

Thus, statins possess beneficial effects in neurological diseases due to their antioxidant and antiinflammatory properties and may be used as a therapeutic drug in neurodegenerative diseases including AD.

²¹⁰/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

Catalpol

Catalpol, an iridoid glucoside derived from the root of rehmannia glutinosa, possesses a broad range of biological and pharmacological activity. The protective effects of catalpol on H_2O_2 , LPS, MPP⁺ and rotenone induced neurotoxicity *in vitro* and *in vivo* has been reported.117,118 Moreover, exposure of cortical neurons to catalpol attenuated (β-Amyloid), Aβ1-42-induced apoptosis mainly via mitochondrial-dependent caspase, as well as decreasing intracellular ROS accumulation, Bax level, cytochrome c release and increasing integrity of mitochondrial membrane potential. These results suggested that catalpol, might be potentially an anti-apoptotic agent against neurodegenerative diseases including AD.¹¹⁹

CONCLUSION

The oxidative damage of cellular molecules plays an important role in neurodegenerative disorders. The brain is a sensitive tissue to oxidative damage, because it contains high concentrations of oxidizable polyunsaturated fatty acids, a high rate of oxygen consumption per unit mass, along with a relatively modest antioxidant defense system. The inhibition of ROS accumulation by different antioxidants is connected to the location of ROS generation. In addition, the mitochondrial dysfunction results in molecular defects in oxidative phosphorylation, which most likely plays a key role in the development and maturation of AD. Thus, targeting mitochondria with antioxidants might be a therapeutic aim, capable of eliminating oxidative damage in the brain, restoring cell integrity and improving cognitive function and spatial memory.

REFERENCES

- 1. Jung HA, Min BS, Yokozawa T, Lee JH, Kim YS, Choi JS. Anti-Alzheimer and antioxidant activities of Coptidis Rhizoma alkaloids. Biol Pharm Bull 2009; 32(8):1433-8.
- 2. Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, et al. Mitochondrial abnormalities in Alzheimer's disease. J Neurosci 2001; 21(9):3017-23.
- 3. Castegna A, Aksenov M, Aksenova M, Thongboonkerd V, Klein JB, Pierce WM, et al. Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part I: creatine kinase BB, glutamine synthase, and

ubiquitin carboxy-terminal hydrolase L-1. Free Radic Biol Med 2002; 33(4):562-71.

- 4. Aliev G, Liu J, Shenk JC, Fischbach K, Pacheco GJ, Chen SG, et al. Neuronal mitochondrial amelioration by feeding acetyl-L-carnitine and lipoic acid to aged rats. J Cell Mol Med 2009; 13(2):320-33.
- 5. Aliev G, Palacios HH, Walrafen B, Lipsitt AE, Obrenovich ME, Morales L. Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. Int J Biochem Cell Biol 2009; 41(10):1989-2004.
- 6. Vladimirova O, Lu FM, Shawver L, Kalman B. The activation of protein kinase C induces higher production of reactive oxygen species by mononuclear cells in patients with multiple sclerosis than in controls. Inflamm Res 1999; 48(7):412-6.
- 7. Manczak M, Park BS, Jung Y, Reddy PH. Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease: implications for early mitochondrial dysfunction and oxidative damage. Neuromolecular Med 2004; 5(2):147-62.
- 8. Sun KH, de Pablo Y, Vincent F, Shah K. Deregulated Cdk5 promotes oxidative stress and mitochondrial dysfunction. J Neurochem 2008; 107(1):265-78.
- 9. Sarti P, Arese M, Giuffrè A. The molecular mechanisms by which nitric oxide controls mitochondrial complex IV. Ital J Biochem 2003; 52(1):37-42.
- 10. Sheng B, Gong K, Niu Y, Liu L, Yan Y, Lu G, et al. Inhibition of gamma-secretase activity reduces Abeta production, reduces oxidative stress, increases mitochondrial activity and leads to reduced vulnerability to apoptosis: Implications for the treatment of Alzheimer's disease. Free Radic Biol Med 2009; 46(10):1362-75.
- 11. Beckman KB, Ames BN. The free radical theory of aging matures. Physiol Rev 1998; 78(2):547–81.
- 12. Aliev G, Obrenovich ME, Reddy VP, Shenk JC, Moreira PI, Nunomura A, et al. Antioxidant therapy in Alzheimer's disease: theory and practice. Mini Rev Med Chem 2008; 8(13):1395-406.
- 13. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants, and the degenerative diseases of aging. Proc Natl Acad Sci U S A 1993; 90(17):7915–22.
- 14. Beal MF. Mitochondria, oxidative damage, and inflammation in Parkinson's disease. Ann N Y Acad Sci 2003; 991:120–31.
- 15. Mirshafiey A, Mohsenzadegan M. Antioxidant therapy in multiple sclerosis. Immunopharmacol Immunotoxicol 2009; 31(1):13-29.

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /211 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

- 16. Mishafiey A, Mohsenzadegan M. immunotoxicological effect of reative oxygen species in multiple sclerosis.J of Chinese clinical medicine 2008; 3:7.
- 17. Zhu X, Su B, Wang X, Smith MA,Perry G. Causes of oxidative stress in Alzheimer disease. Cell. Mol. Life Sci 2007; 64(17):2202–10.
- 18. Lambeth JD. NOX enzymes and the biology of reactive oxygen. Nat Rev Immunol 2004; 4(3):181–9.
- 19. Ravikumar B, Vacher C, Berger Z, Davies JE, Luo S, Oroz LG, et al. Inhibition of mTOR induces autophagy and reduces toxicity of polyglutamine expansions in fly and mouse models of Huntington disease. Nat Genet 2004; 36(6):585-95.
- 20. Levine B, Klionsky DJ. Development by self-digestion: molecular mechanisms and biological functions of autophagy. Dev Cell 2004; 6(4):463-77.
- 21. Lipinski MM, Zheng B, Lu T, Yan Z, Py BF, Ng A, et al. Genome-wide analysis reveals mechanisms modulating autophagy in normal brain aging and in Alzheimer's disease. Proc Natl Acad Sci U S A 2010; 107(32):14164-9.
- 22. Ilhan A, Akyol O, Gurel A, Armutcu F, Iraz M, Oztas E. Protective effects of caffeic acid phenethyl ester against experimental allergic encephalomyelitis induced oxidative stress in rats. Free Radic Biol Med 2004; 37(3):386–94.
- 23. Lum H, Roebuck KA. Oxidant stress and endothelial cell dysfunction. Am J Physiol. Cell Physiol 2001; 280(4):C719-41.
- 24. Park L, Anrather J, Zhou P, Frys K, Pitstick R, Younkin S, et al. NADPH-oxidase-derived reactive oxygen species mediate the cerebrovascular dysfunction induced by the amyloid beta peptide. J Neurosci 2005; 25(7):1769-77.
- 25. Abdala-Valencia H, Cook-Mills JM. VCAM-1 signals activate endothelial cell protein kinase cα via oxidation. J Immunol 2006; 177(9):6379–87.
- 26. Mirshafiey A, Mohsenzadegan M. The role of reactive oxygen species in immunopathogenesis of rheumatoid arthritis. Iran J Allergy Asthma Immunol 2008 ; 7(4):195- 202.
- 27. Calabrese V, Lodi R, Tonon C, D'Agata V, Sapienza M, Scapagnini G, et al. Oxidative stress, mitochondrial dysfunction and cellular stress response in Friedreich's ataxia. J Neurol Sci 2005; 233 (1-2):145–62.
- 28. Ruuls SR, Bauer J, Sontrop K, Huitinga I, 't Hart BA, Dijkstra CD. Reactive oxygen species are involved in the pathogenesis of experimental allergic encephalomyelitis in Lewis rats, J. Neuroimmunol. 1995; 56(2):207–17.
- 29. Torreilles F, Salman-Tabcheh S, Guérin M, Torreilles J. Brain Res 1999; 30(2):153-63.
- 30. Mirshafiey A, Matsuo H, Nakane S, Rehm BH, Koh CS, Miyashi S. Novel immunosuppressive therapy by M2000 in experimental multiple sclerosis. Immunopharmacol Immunotoxicol 2005; 27(2):255–65.
- 31. Vrabec JP, Lieven CJ, Levin LA. Cell-type-specific opening of the retinal ganglion cell mitochondrial permeability transition pore. Invest. Ophthalmol. Vis.Sci 2003; 44(6):2774–82.
- 32. Dumont M, Ho DJ, Calingasan NY, Xu H, Gibson G, Beal MF. Mitochondrial dihydrolipoyl succinyltransferase deficiency accelerates amyloid pathology and memory deficit in a transgenic mouse model of amyloid deposition. Free Radic Biol Med 2009; 47(7):1019-27.
- 33. Melov S, Adlard PA, Morten K, Johnson F, Golden TR, Hinerfeld D, et al. Mitochondrial oxidative stress causes hyperphosphorylation of tau. PLoS One 2007; 2(6):e536.
- 34. Dumont M, Wille E, Stack C, Calingasan NY, Beal MF, Lin MT. Reduction of oxidative stress, amyloid deposition, and memory deficit by manganese superoxide dismutase overexpression in a transgenic mouse model of Alzheimer's disease. FASEB J 2009; 23(8):2459-66.
- 35. Gibson GE, Blass JP, Beal MF, Bunik V. The alphaketoglutarate-dehydrogenase complex: a mediator between mitochondria and oxidative stress in neurodegeneration. Mol Neurobiol 2005; 31(1-3):43–63.
- 36. Clement AB, Gimpl G, Behl C. Oxidative stress resistance in hippocampal cells is associated with altered membrane fluidity and enhanced nonamyloidogenic cleavage of endogenous amyloid precursor protein. Free Radic Biol Med. 2010; 48(9):1236-41.
- 37. Kojro E, Gimpl G, Lammich S, Marz W, Fahrenholz F. Low cholesterol stimulates the nonamyloidogenic pathway by its effect on the alpha-secretase ADAM 10. Proc Natl Acad Sci USA 2001; 98(10):5815–20.
- 38. Bodovitz S,Klein WL. Cholesterol modulates alphasecretase cleavage of amyloid precursor protein. J Biol Chem 1996; 271(8):4436–40.
- 39. Block ML. NADPH oxidase as a therapeutic target in Alzheimer's disease. BMC Neurosci 2008; 9 Suppl 2:S8.
- 40. Qin L, Liu Y, Wang T, Wei SJ, Block ML, Wilson B, et al. NADPH oxidase mediates lipopolysaccharideinduced neurotoxicity and proinflammatory gene expression in activated microglia. J Biol Chem 2004; 279(2):1415-21.
- 41. Wang T, Qin L, Liu B, Liu Y, Wilson B, Eling TE, et al. Role of reactive oxygen species in LPS-induced

²¹²/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

production of prostaglandin E2 in microglia. J Neurochem 2004; 88(4):939-47.

- 42. Ray B, Lahiri DK. Neuroinflammation in Alzheimer's disease: different molecular targets and potential therapeutic agents including curcumin. Curr Opin Pharmacol 2009; 9(4):434-44.
- 43. Arnaud L, Robakis NK, Figueiredo-Pereira ME.It may take inflammation, phosphorylation and ubiquitination to 'tangle' in Alzheimer's disease. Neurodegener Dis 2006; 3(6):313-19.
- 44. Davis JB, McMurray HF, Schubert D.The amyloid betaprotein of Alzheimer's disease is chemotactic for mononuclear phagocytes. Biochem Biophys Res Commun 1992; 189(2):1096-100.
- 45. Ophir G, Amariglio N, Jacob-Hirsch J, Elkon R, Rechavi G, Michaelson DM. Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. Neurobiol Dis 2005; 20(3):709-18.
- 46. Barger SW, Mattson MP. Induction of neuroprotective kappa Bdependent transcription by secreted forms of the Alzheimer's beta-amyloid precursor. Brain Res Mol Brain Res 1996; 40(1):116-26.
- 47. Sanchez-Ortiz E, Hahm BK, Armstrong DL, Rossie S. Protein phosphatase 5 protects neurons against amyloidbeta toxicity. J Neurochem 2009; 111(2):391-402.
- 48. Marques CA,Keil U,Bonert A,Steiner B,Haass C,Muller WE,Eckert A. Neurotoxic mechanisms caused by the Alzheimer's disease-linked Swedish amyloid precursor protein mutation: oxidative stress, caspases, and the JNK pathway. J Biol Chem 2003; 278(30):28294–302.
- 49. Keil U, Bonert A, Marques CA, Scherping I, Weyermann J, Strosznajder JB, et al. Amyloid β-induced Changes in Nitric Oxide Production and Mitochondrial Activity Lead to Apoptosis. J Biol Chem 2004; 279(48):50310–20.
- 50. Zhou G, Golden T, Aragon IV, Honkanen RE. Ser/Thr protein phosphatase 5 inactivates hypoxia-induced activation of an apoptosis signal-regulating kinase 1/MKK-4/JNK signaling cascade. J Biol Chem 2004; 279(45):46595–605.
- 51. Shinoda S, Skradski SL, Araki T, Schindler CK, Meller R, Lan JQ, et al. Formation of a tumour necrosis factor receptor 1 molecular scaffolding complex and activation of apoptosis signal-regulating kinase 1 during seizureinduced neuronal death. Eur J Neurosci 2003; 17(10):2065–76.
- 52. Du H, Yan SS. Mitochondrial permeability transition pore in Alzheimer's disease: cyclophilin D and amyloid beta. Biochim Biophys Acta 2010; 1802(1):198-204.
- 53. Mark RJ, Hensley K, Butterfield DA, Mattson MP. Amyloid beta-peptide impairs ion-motive ATPase activities: evidence for a role in loss of neuronal Ca2+ homeostasis and cell death. J Neurosci 1995; 15(9):6239-49.
- 54. Ekinci FJ, Malik KU, Shea TB. Activation of the L voltage-sensitive calcium channel by mitogen-activated protein (MAP) kinase following exposure of neuronal cells to beta-amyloid. MAP kinase mediates betaamyloid-induced neurodegeneration. J Biol Chem 1999; 274(42):30322-7.
- 55. Cowburn RF, Wiehager B, Trief E, Li-Li M, Sundström E. Effects of beta-amyloid-(25-35) peptides on radioligand binding to excitatory amino acid receptors and voltage-dependent calcium channels: evidence for a selective affinity for the glutamate and glycine recognition sites of the NMDA receptor. Neurochem Res 1997; 22(12):1437-42.
- 56. Hiruma H, Katakura T, Takahashi S, Ichikawa T, Kawakami T. Glutamate and amyloid beta-protein rapidly inhibit fast axonal transport in cultured rat hippocampal neurons by different mechanisms. J Neurosci 2003; 23(26):8967-77.
- 57. Kim JH, Rah JC, Fraser SP, Chang KA, Djamgoz MB, Suh YH. Carboxyl-terminal peptide of beta-amyloid precursor protein blocks inositol 1,4,5-trisphosphatesensitive Ca2+ release in Xenopus laevis oocytes. J Biol Chem 2002; 277(23):20256-63.
- 58. Veal EA, Day AM,Morgan BA. Hydrogen peroxide sensing and signaling. Mol Cell 2007; 26(1):1–14.
- 59. Chang TS, Jeong W, Choi SY, Yu S, Kang SW, Rhee SG. Regulation of peroxiredoxin I activity by Cdc2 mediated phosphorylation. J Biol Chem 2002; 277(28):25370–76.
- 60. Borsello T, Forloni G. JNK signalling: a possible target to prevent neurodegeneration. Curr Pharmaceut Design 2007; 13(18):1875–86.
- 61. Sun KH, Lee HG, Smith MA, Shah K. Direct and indirect roles of cyclin-dependent kinase 5 as an upstream regulator in the c-Jun NH2-terminal kinase cascade: relevance to neurotoxic insults in Alzheimer's disease. Mol Biol Cell 2009; 20(21):4611-9.
- 62. Shen C, ChenY, Liu H, Zhang K, Zhang T, Lin A, et al. Hydrogen peroxide promotes Abeta production through JNK-dependent activation of gamma-secretase. J Biol Chem 2008; 283(25):17721–30.
- 63. Tamagno E, Guglielmotto M, Aragno M, Borghi R, Autelli R, Giliberto L, et al. Oxidative stress activates a positive feedback between the gammaand beta-secretase

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /213 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

cleavages of the beta-amyloid precursor protein. J Neurochem 2008; 104(3):683–95.

- 64. Marques CA, Keil U, Bonert A, Steiner B, Haass C, Muller WE, et al. Neurotoxic mechanisms caused by the Alzheimer's disease-linked Swedish amyloid precursor protein mutation: oxidative stress, caspases, and the JNK pathway. J Biol Chem 2003; 278(30):28294–302.
- 65. Hureau C, Faller P. Abeta-mediated ROS production by Cu ions: structural insights, mechanisms and relevance to Alzheimer's disease. Biochimie 2009; 91(10):1212-7.
- 66. da Silva GF, Lykourinou V, Angerhofer A, Ming LJ. Methionine does not reduce Cu(II)-beta-amyloid!- rectification of the roles of methionine-35 and reducing agents in metal-centered oxidation chemistry of Cu(II) beta-amyloid. Biochim Biophys Acta 2009; 1792(1):49- 55.
- 67. Pogue AI, Li YY, Cui JG, Zhao Y, Kruck TP, Percy ME, et al. Characterization of an NF-kappaB-regulated, miRNA-146a-mediated down-regulation of complement factor H (CFH) in metal-sulfate-stressed human brain cells. J Inorg Biochem 2009; 103(11):1591-5.
- 68. Guilloreau L, Combalbert S, Sournia-Saquet A, Marzaguil H, Faller P. Redox chemistry of copperamyloid-beta: the generation of hydroxyl radical in the presence of ascorbate is linked to redox-potentials and aggregation state. Chem Biol Chem 2007; 8(11):1317–25.
- 69. Baruch-Suchodolsky R, Fischer B. Soluble amyloid beta1-28-copper(I)/copper(II)/Iron(II) complexes are potent antioxidants in cell-free systems. Biochemistry 2008; 47(30):7796–806.
- 70. Kowalik-Jankowska T, Ruta M, Wisniewska K, Lankiewicz L, Dyba M. Products of Cu(II)-catalyzed oxidation in the presence of hydrogen peroxide of the 1– 10, 1–16 fragments of human and mouse b-amyloid peptide. J Inorg Biochem 2004; 98(6):940–50.
- 71. Murakami K, Hara H, Masuda Y, Ohigashi H, Irie K. Distance measurement between Tyr10 and Met35 in amyloid beta by site-directed spin-labeling ESR spectroscopy: implications for the stronger neurotoxicity of Abeta42 than Abeta40. Chem Bio Chem.2007; 8: 2308–14.
- 72. Hou L, Kang I, Marchant RE, Zagorski MG. Methionine 35 oxidation reduces fibril assembly of the amyloid abeta- (1–42) peptide of Alzheimer's disease. J Biol Chem 2002; 277(43):40173–76.
- 73. Johansson AS, Bergquist J, Volbracht C, Päiviö A, Leist M, Lannfelt L, et al. Attenuated amyloid-beta aggregation and neurotoxicity owing to methionine oxidation. Neuroreport 2007; 18(6):559-63.
- 74. Clementi ME, Martorana GE, Pezzotti M, Giardina B, Misiti F. Methionine 35 oxidation reduces toxic effects of the amyloid β-protein fragment(31–35) on human red blood cell. Int J Biochem Cell Biol 2004; 36(10):2066– 76.
- 75. da Silva GF, Ming LJ. Alzheimer's disease related copper (II)-beta-amyloid peptide exhibits phenol monooxygenase and catechol oxidase activities. Angew Chem Int Ed Engl 2005; 44(34):5501-4.
- 76. Campbell A, Yang EY, Tsai-Turton M, Bondy SC. Proinflammatory effects of aluminum in human glioblastoma cells. Brain Res 2002; 933(1):60-5.
- 77. Kawahara M. Effects of aluminum on the nervous system and its possible link with neurodegenerative diseases. J Alzheimers Dis 2005; 8(2):171-82.
- 78. Lukiw WJ, Percy ME, Kruck TP. Nanomolar aluminum induces pro-inflammatory and pro-apoptotic gene expression in human brain cells in primary culture. J Inorg Biochem 2005; 99(9):1895-8.
- 79. Saunders AM, Schmader K, Breitner JC, Benson MD, Brown WT, Goldfarb L, et al. Apolipoprotein E epsilon 4 allele distributions in late-onset Alzheimer's disease and in other amyloid-forming diseases. Lancet 1993; 342(8873):710–1.
- 80. Ashford JW. APOE genotype effects on Alzheimer's disease onset and epidemiology. J Mol Neurosci 2004; 23(3):157–65.
- 81. Ramassamy C, Averill D, Beffert U, Theroux L, Lussier-Cacan S, Cohn JS, et al. Oxidative insults are associated with apolipoprotein E genotype in Alzheimer's disease brain. Neurobiol Dis 2000; 7(1):23–37.
- 82. Cedazo-Minguez A. Apolipoprotein E and Alzheimer's disease: molecular mechanisms and therapeutic opportunities. J Cell Mol Med 2007; 11(6):1227–38.
- 83. Reid PC, Urano Y, Kodama T, Hamakubo T. Alzheimer's disease: cholesterol, membrane rafts, isoprenoids and statins. J Cell Mol Med 2007; 11(3):383-92.
- 84. Refolo LM, Malester B, LaFrancois J, Bryant-Thomas T, Wang R, Tint GS, et al. Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. Neurobiol Dis 2000; 7(4):321- 31.
- 85. Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, et al. Mitochondrial abnormalities in Alzheimer's disease. J Neurosci 2001; 21(9):3017-23.
- 86. Di Domenico F, Sultana R, Tiu GF, Scheff NN, Perluigi M, Cini C, et al. Protein levels of heat shock proteins 27, 32, 60, 70, 90 and thioredoxin-1 in amnestic mild cognitive impairment: an investigation on the role of

²¹⁴/ IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY Vol. 11, No. 3, September 2012

Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

cellular stress response in the progression of Alzheimer disease. Brain Res 2010; 1333:72-81.

- 87. Mosser DD, Morimoto RI. Molecular chaperones and the stress of oncogenesis. Oncogene 2004; 23(16):2907–18.
- 88. Evans CG, Wisen S, Gestwicki JE. Heat shock proteins 70 and 90 inhibit early stages of amyloid beta-(1-42) aggregation in vitro. J Biol Chem 2006; 281(44):33182– 91.
- 89. Bjorkdahl C, Sjogren MJ, Zhou X, Concha H, Avila J, Winblad B, et al. Small heat shock proteins Hsp27 or alphaB-crystallin and the protein components of neurofibrillary tangles: tau and neurofilaments. J Neurosci Res 2008; 86(6):1343–52.
- 90. Pérez-De La Cruz V, Elinos-Calderón D, Robledo-Arratia Y, Medina-Campos ON, Pedraza-Chaverrí J, Ali SF, et al. Targeting oxidative/nitrergic stress ameliorates motor impairment, and attenuates synaptic mitochondrial dysfunction and lipid peroxidation in two models of Huntington's disease. Behav Brain Res 2009; 199(2):210- 7.
- 91. Gomez-Pinilla F. Brain foods: the effects of nutrients on brain function. Nat Rev Neurosci 2008; 9(7):568–78.
- 92. Shenk JC, Liu J, Fischbach K, Xu K, Puchowicz M, Obrenovich ME, et al. The effect of acetyl-L-carnitine and R-alpha-lipoic acid treatment in ApoE4 mouse as a model of human Alzheimer's disease. J Neurol Sci 2009; 283(1-2):199-206.
- 93. Cormier A, Morin C, Zini R, Tillement JP, Lagrue G. In vitro effects of nicotine on mitochondrial respiration and superoxide anion generation. Brain Res 2001; 900(1):72- 9.
- 94. Shimohama S, Akaike A, Kimura J. Nicotine-induced protection against glutamate cytotoxicity. Nicotinic cholinergic receptor-mediated inhibition of nitric oxide formation. Ann N Y Acad Sci 1996; 777:356-61.
- 95. Ezoulin MJ, Li J, Wu G, Dong CZ, Ombetta JE, Chen HZ, et al. Differential effect of PMS777, a new type of acetylcholinesterase inhibitor, and galanthamine on oxidative injury induced in human neuroblastoma SK-N-SH cells. Neurosci Lett 2005; 389(2):61-5.
- 96. Ferrera P. Differential effects of COX inhibitors against beta-amyloid-induced neurotoxicity in human neuroblastoma cells. Arias C. Neurochem Int 2005; 47(8):589-96.
- 97. Lee M, Sparatore A, Del Soldato P, McGeer E, McGeer PL. Hydrogen sulfide-releasing NSAIDs attenuate neuroinflammation induced by microglial and astrocytic activation. Glia 2010; 58(1):103-13.
- 98. Nivsarkar M, Banerjee A, Padh H. Cyclooxygenase inhibitors: a novel direction for Alzheimer's management. Pharmacol Rep 2008; 60(5):692-8.
- 99. Liu Y, Qin L, Li G, Zhang W, An L, Liu B, et al. Dextromethorphan protects dopamanergic neurons against inflammationmediated degeneration through inhibition of microglial activation. J Pharmacol Exp Ther 2003; $305(1):1-7.$
- 100. Li G, Cui G, Tzeng NS, Wei SJ, Wang T, Block ML, et al. Femtomolar concentrations of dextromethorphan protect mesencephalic dopaminergic neurons from inflammatory damage. Faseb J 2005; 19(6):489-96.
- 101. Zhang W, Wang T, Qin L, Gao HM, Wilson B, Ali SF, et al. Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase. Faseb J 2004; 18(3):589-91.
- 102. Queen BL, Tollefsbol TO. Polyphenols and aging. Curr Aging Sci 2010; 3(1):34-42.
- 103. Perron NR, Brumaghim JL. A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. Cell Biochem Biophys 2009; 53(2):75–100.
- 104. Vingtdeux V, Chandakkar P, Zhao H, d'Abramo C, Davies P, Marambaud P. Novel synthetic small-molecule activators of AMPK as enhancers of autophagy and amyloid-β peptide degradation. FASEB J 2011; 25(1):219-31.
- 105. Wang J, Ho L, Zhao Z, Seror I, Humala N, Dickstein DL, et al. Moderate consumption of Cabernet Sauvignon attenuates Abeta neuropathology in a mouse model of Alzheimer's disease. FASEB J 2006; 20(13):2313-20.
- 106. Motterlini R, Foresti R, Bassi R, Green CJ. Curcumin, an antioxidant and anti-inflammatory agent, induces heme oxygenase-1 and protects endothelial cells against oxidative stress. Free Radic Biol Med 2000; 28(8):1303- 12.
- 107. Baum L, Ng A. Curcumin interaction with copper and iron suggests one possible mechanism of action in Alzheimer's disease animal models. J Alzheimers Dis 2004; 6(4):367-77.
- 108. Meeran SM, Akhtar S, Katiyar SK. Inhibition of UVBinduced skin tumor development by drinking green tea polyphenols is mediated through DNA repair and subsequent inhibition of inflammation. J Invest Dermatol 2009; 129(5):1258–70.
- 109. Punathil T, Tollefsbol TO, Katiyar SK. EGCG inhibits mammary cancer cell migration through inhibition of nitric oxide synthase and guanylate cyclase. Biochem Biophys Res Commun 2008; 375(1):162–7.

Vol. 11, No. 3, September 2012 IRANIAN JOURNAL OF ALLERGY, ASTHMA AND IMMUNOLOGY /215 Published by Tehran University of Medical Sciences (http://ijaai.tums.ac.ir)

- 110. Bastianetto S, Brouillette J, Quirion R. Neuroprotective effects of natural products: interaction with intracellular kinases, amyloid peptides and a possible role for transthyretin. Neurochem Res 2007; 32(10):1720–5.
- 111. Boots AW, Haenen GR, Bast A. Health effects of quercetin: from antioxidant to nutraceutical. Eur J Pharmacol 2008; 585(2-3):325–37.
- 112. Guy J, Ellis EA, Hope GM, Rao NA. Antioxidant enzyme suppression of demyelination in experimental optic neuritis. Curr Eye Res 1989; 8(5):467–77.
- 113. Pahan K, Namboodiri AM, Sheikh FG, Smith BT, Singh I. Increasing Camp attenuates induction of inducible nitric-oxide synthase in rat primary astrocytes. J Biol Chem 1997; 272(12):7786–91.
- 114. Fassbender K, Simons M, Bergmann C, Stroick M, Lutjohann D, Keller P, et al. Simvastatin strongly reduces levels of Alzheimer's disease beta -amyloid peptides Abeta 42 and Abeta 40 in vitro and in vivo. Proc Natl Acad Sci U S A 2001; 98(10):5856-61.
- 115. Fraunberger P, Gröne E, Gröne HJ, Walli AK. Simvastatin reduces endotoxin-induced nuclear factor kappaB activation and mortality in guinea pigs despite

lowering circulating low-density lipoprotein cholesterol. Shock 2009; 32(2):159-63.

- 116. Won JS, Im YB, Khan M, Contreras M, Singh AK, Singh I. Lovastatin inhibits amyloid precursor protein (APP) beta-cleavage through reduction of APP distribution in Lubrol WX extractable low density lipid rafts. J Neurochem 2008; 105(4):1536-49.
- 117. Bi J, Jiang B, Hao S, Zhang A, Dong Y, Jiang T, et al. Catalpol attenuates nitric oxide increase via ERK signaling pathways induced by rotenone in mesencephalic neurons. Neurochem Int 2009; 54(3- 4):264–70.
- 118. Jiang B, Zhang H, Bing J, Zhang XL. Neuroprotective activities of catalpol on MPP+/MPTP-induced neurotoxicity. Neurol Res 2008; 30(6):639-44.
- 119. Liang JH, Du J, Xu LD, Jiang T, Hao S, Bi J, et al. Catalpol protects primary cultured cortical neurons induced by Abeta(1-42) through a mitochondrialdependent caspase pathway. Neurochem Int 2009; 55(8):741-6.