Immunogenicity Assessment of *Brucella mellitensis* HSP and TF Proteins by Immunized Rabbit Serum

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Bacteria of the genus Brucella are facultative intracellular pathogens which have developed the capacity to survive and multiply in professional and nonprofessional phagocytes. Due to drawbacks of live attenuated vaccines, much attention has been focused on screening Brucella-protective antigens as subunit vaccine candidates. In order to screen immunogenic candidate antigens for the development of a Brucella subunit vaccine, we cloned, expressed and purified Heat Shock Protein (HSP) and Trigger Factor (TF) from Brucella melitensis. These recombinant antigens were then evaluated by serum from a B. mellitensis-vaccinated rabbit using ELISA and Western blot. Our results showed that the immunized rabbit serum reacted with recombinant HSP and TF in ELISA and Western blot. These results may suggest that B.melitensis rTF and rHSP may serve as candidate subunit vaccine components for protection against the infection.

Brucella spp. are Gram-negative and facultative intracellular bacteria which cause brucellosis, a

worldwide zoonotic disease causing abortion in domestic animals and Malta fever in humans.¹ Brucella melitensis can invade macrophage-monocyte lineage cells and replicate within the phagosomes by inhibiting fusion.² phagosome-lysosome In intracellular environment of macrophages, bacteria are subjected to hard conditions³ thus, the identification of bacterial proteins essential for intracellular survival in this niche is critical in understanding the protective mechanisms and pathogenesis of the disease. Expression of heat shock proteins in this situation, causes bacteria to adapt not only to thermal but also to various other environmental stresses, and the accumulation of heatshock proteins (HSPs) is thought to preserve bacterial cellular functions.⁴ The heat shock protein is a small HSP (sHSP) (Accession No. 1197813). sHSPs are molecular chaperones that suppress protein aggregation and protect against cell stress.⁵ Trigger Factor (TF) protein (Accession No. 1196780) is an ATP independent chaperone⁶ and has also been reported to act as a protective antigen against B.melitensis infection.⁷ In view of the immunological importance of HSP and TF, we decided to clone, express in E. coli and purify the HSP and TF proteins from B. melitensis and study the antibody response to this protein in sera from B. melitensis- vaccinated rabbit by ELISA and

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Western blot. The Gateway cloning system (Invitrogen, NY, USA) was used for cloning of genes. Primers used for synthesis of the two genes were designed (Table 1). Cloning procedure was described previously.⁸ rHSP and rTF were successfully expressed in the insoluble and soluble fractions of *E. coli* cells. Purification of rHSP and rTF were done as described previously.⁷

Purity was assessed by SDS-PAGE and Coomassie blue staining. Sera were obtained from a Newzealand White Rabbit before and after immunization with *B.melitensis Rev.1*. To study the recognition of recombinant proteins by immunized rabbit serum, Western blot and ELISA were used.



Figure 1. Analysis of *B. mellitensis* recombinant proteins and lysate reactivity with immunized rabbit serum A.ELISA analysis of immunized rabbit serum with *B. melitensis* lysate, rTF and rHSP by ELISA B.Western blot analysis of immune reactivity of immunized rabbit serum with rTF (Lane 1) and rHSP (lane 2)

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| Gene | Forward | Reverse |
|------|------------------------------------|--------------------------------|
| hsp | 5- CACC ATGCGTCACG TAGATTTTTCC | 5- ATGCGCGGTCTTCGCCTCAATCG |
| tf | 5- CACCATGACAAGAA G TGAAGG TTTGAAC | 5- AAAAGCCTCTTCGGACTTGCCTTCTTC |

B. melitensis Rev.1, an attenuated smooth strain used to control B. melitensis infection, induces heterologous protection against other Brucella spp. and is currently considered as the best vaccine for the prophylaxis of caprine brucellosis.9 However, due to different problems caused by administration of live attenuated vaccine, a subunit vaccine that is protective against *B. melitensis* is desirable.¹⁰ The immunized rabbit serum reacted with both recombinant proteins in ELISA and Western blot that suggests the immunogenic nature of these subunits. These data are also confirmed by Yang et al. that immunization with rTF protein could protect mice against infection with B. *melitensis.*⁷ To our knowledge, HSP from *B. melitensis* has not been reported as an immunogenic subunit of this organism. Our data may also suggest this protein as a vaccine candidate subunit. Much work is needed to be performed to establish this notion which is the theme of our future research.

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REFERENCES

- Rajashekara G, Glasner JD, Glover DA, Splitter GA. Comparative whole-genome hybridization reveals genomic islands in Brucella species. Journal of bacteriology 2004; 186(15):5040-51.
- Sanakkayala N, Sokolovska A, Gulani J, Hogenesch H, Sriranganathan N, Boyle SM, et al. Induction of antigenspecific Th1-type immune responses by gamma-irradiated

recombinant Brucella abortus RB51. Clin Diagn Lab Immunol 2005; 12(12):1429-36.

- Contreras-Rodriguez A, Ramirez-Zavala B, Contreras A, Schurig GG, Sriranganathan N, Lopez-Merino A. Purification and characterization of an immunogenic aminopeptidase of Brucella melitensis. Infect Immun 2003; 71(9):5238-44.
- Delpino MV, Estein SM, Fossati CA, Baldi PC, Cassataro J. Vaccination with Brucella recombinant DnaK and SurA proteins induces protection against Brucella abortus infection in BALB/c mice. Vaccine 2007; 25(37-38):6721-9.
- Haslbeck M, Buchner J. Chaperone function of sHsps. Prog Mol Subcell Biol 2002; 28:37-59.
- Ferbitz L, Maier T, Patzelt H, Bukau B, Deuerling E, Ban N. Trigger factor in complex with the ribosome forms a molecular cradle for nascent proteins. Nature 2004; 431(7008):590-6.
- Yang X, Hudson M, Walters N, Bargatze RF, Pascual DW. Selection of protective epitopes for Brucella melitensis by DNA vaccination. Infect Immun 2005; 73(11):7297-303.
- Ding XZ, Paulsen IT, Bhattacharjee AK, Nikolich MP, Myers G, Hoover DL. A high efficiency cloning and expression system for proteomic analysis. Proteomics 2006; 6(14):4038-46.
- Marin CM, Barberan M, Jimenez de Bagues MP, Blasco JM. Comparison of subcutaneous and conjunctival routes of Rev 1 vaccination for the prophylaxis of Brucella ovis infection in rams. Res Vet Sci 1990; 48(2):209-15.
- Yang Y, Yin J, Guo D, Lang X, Wang X. Immunization of mice with recombinant S-adenosyl-L-homocysteine hydrolase protein confers protection against Brucella melitensis infection. FEMS Immunol Med Microbiol 2011; 61(2):159-67.