

ORIGINAL ARTICLE

Iran J Allergy Asthma Immunol
December 2013; 12(4):391-396.

IL-13 Gene Polymorphisms and Their Association with Atopic Asthma and Rhinitis in Pakistani Patients

Micheal Shazia^{1,2}, Minhas Kanza¹, Ishaque Mehwish¹, Shabbir Irum¹, Ahmed Farida³, and Ahmed Asifa¹

¹ Department of Biosciences, COMSATS Institute of Information Technology, Islamabad-44000, Pakistan,

² Department of Ophthalmology, Radboud University Nijmegen Medical Centre; Nijmegen, the Netherlands

³ National Institute of Health, Islamabad, Pakistan

Received: 10 October 2012; Received in revised form: 5 December 2012; Accepted: 23 December 2012

ABSTRACT

Interleukin-13 (IL-13) is known to be a key regulator in immunoglobulin E (IgE) synthesis, mucus hypersecretion and airway hyperresponsiveness. Single nucleotide polymorphisms in IL-13 are associated with allergic phenotypes in several ethnically diverse populations.

This study was performed in 214 atopic patients (asthma n=108, allergic rhinitis n=106) and sex-matched healthy controls (n=120). Genotyping of IL-13 gene polymorphisms was performed using polymerase chain reaction-based restriction fragment length polymorphism method.

A statistically significant association of the A-1512C polymorphism in IL13 gene was observed with atopy ($p < 0.001$; $\chi^2 = 19.0$). Upon stratification of the data into asthma and AR association was revealed with both asthma ($p = 0.01$; $\chi^2 = 8.80$) and AR ($p < 0.001$; $\chi^2 = 24.3$) in Pakistani patients. Higher odds ratio (OR 95% CI) was observed for AR 3.42 (2.04-5.76) relative to asthma 2.40 (1.41-4.09) for the C allele compared to controls.

In conclusion the study provides the evidence that A-1512C polymorphisms in IL-13 is a risk factor for asthma and AR.

Keywords: Allergic rhinitis(AR); Atopic asthma; Interleukin-13 (IL-13); Polymorphism

INTRODUCTION

IL-13 is a critical cytokine located on chromosome 5q, a region frequently linked to asthma, allergic rhinitis (AR), airway responsiveness, and other

related phenotypes. Higher expression of IL-13 at both mRNA and protein level was observed after exposure to allergen, in the sputum and bronchoalveolar lavage (BAL) fluid of the allergic asthma patients and other related traits such as high eosinophil counts and airway hyper-responsiveness.¹⁻⁴ In AR, IL-13 was observed to be expressed in the nasal mucosa of patients with perennial atopic rhinitis after allergen provocation.^{5,6}

To support the key role of IL-13 mediator in asthma, several groups have studied IL-13 in mouse

Corresponding Author: Micheal Shazia, MD;
Department of Biosciences, COMSATS Institute of Information Technology, Park Road, Chak Shahzad, Islamabad-44000, Pakistan.
Tel: (92 51) 9235 033, Fax: (92 51) 4442 805; E-mail: shaziamicheal@gmail.com

models. IL-13 null mice were unable to produce goblet cells responsible for mucus over-production in asthma and could not recover the basic IgE levels even upon stimulation by IL-4.^{7,8} IL-13 is a major contributor in the development of a late nasal response therefore inhibition of IL-13 may be considered an important therapeutic application in preventing the persistent nasal blockage in AR.⁹ To further support function of IL-13 in allergic responses recently several single nucleotide polymorphisms (SNPs) have been reported for their association with the allergic diseases such as asthma and AR.^{10,11} Graves *et al* have identified seven polymorphisms in various parts of the IL-13 gene. Among these polymorphisms two were in the promoter region C-1055T and A-1512C.¹² C-1055T polymorphism was reported with altered regulation of IL-13 and strong involvement of TT genotype in the Dutch population. In a recent meta-analysis of C-1055T SNP in asthma, a strong association was observed in Caucasians.¹³ In addition to these two SNPs, one was found in the third intron of the IL-13 gene and three were found in the 3' untranslated region (G+2525A, C+2580A and T+2749C).¹²

Another important SNP, Gln144Arg also known as +2044 G/A (Arg130Gln) studied in different populations, is present in the coding region of exon 4 and was found to be associated with asthma in British and Japanese populations¹⁰ along with increased total serum IgE (TsIgE) levels in the American and German cohorts.¹² Individuals with the GA +2044 genotype had a 40% increased risk of asthma compared to the Homozygous GG and Asians were considered at high risk compared to Caucasians. Vladich *et al* investigated the functionality of the IL-13 R130Q variants in peripheral blood cells of normal individuals and showed that the IL-13 130Q variant was more active than the wild type IL-13 130R variant, and was less neutralized by soluble IL-13 receptor-alpha.¹⁴ Similarly, in a meta-analysis of IL-13 SNPs in AR, a strong association of +2044 A allele was observed.¹⁵

In most of the previous studies the constant genetic associations of IL-13 polymorphisms have been observed with asthma and related traits across diverse ethnic populations. They help to strong contention that IL-13 acts as a major candidate for its association with asthma and atopy. Therefore, the aim of the present case control study was to evaluate the role of IL-13 SNPs in the Pakistani atopic asthma and AR patients.

MATERIALS AND METHODS

Subjects

The case control study included 120 controls and 214 atopic patients (108 atopic asthma patients and 106 AR patients). Selection criteria for the patients and controls were described previously.¹⁶ To be qualified as a control for the study, healthy individual had no history of allergy, and negative skin prick test to different aeroallergens. TsIgE levels were also lower than 100 IU/ml in controls. Patients were diagnosed for atopic asthma according to the following criteria: Skin prick test positive reactions to at least one of the aeroallergens (pollens) being tested as allergens. Allergic asthma patients had a history of attacks of short breathlessness along with wheezing due to chest tightness. Allergic rhinitis patients had symptoms of sneezing, runny nose, nasal obstruction, itchy nose and rhinorrhea. All the patients were recruited from the Allergy Centre of National Institute of Health Islamabad Pakistan.

Genomic DNA and genotyping

This study conforms to the tenets of the Helsinki declaration and has been approved by the Departmental Review and Ethics Committee. All subjects were briefed about the study in their local language and written informed consent was obtained before obtaining their blood samples. Genomic DNA was obtained using a standard phenol chloroform method.¹⁷

Genotyping of SNPs

Performed for the detection of three polymorphisms in IL-13 gene by using the PCR-RFLP procedure.

Genomic DNA was genotyped for three non-synonymous SNPs located in IL-13: rs20541 (c.431G>A; p.Gln144Arg) in exon 4, rs1881457 (c.-1470A>C) and rs847 (c.*695T>C) in 5' and 3' UTR, respectively. The primers given in table 1 were used to amplify products, which spanned polymorphic sites in IL-13 gene. The polymerase chain reaction (PCR) mixture contained 2.5U *Taq* polymerase (Fermentas, Burlington, Ontario), 1x PCR buffer (10mM Tris-HCl, pH 9.0, 50mM KCl, 0.1% Triton X-100, 0.01% gelatin) (Fermentas, Burlington, Ontario), 1.5mM MgCl₂, 10pmol each primer, and 50ng DNA. PCR amplification was performed with an initial denaturation at 95 °C for 5 min, followed by 35 cycles of denaturation at 95 °C for 45 s, primer annealing given

IL-13 Gene Polymorphisms and Atopic Asthma and Rhinitis

Table 1. List of primers used for genotyping IL-13 SNPs

Primers	Primers Sequence	°C†	PCR Product
G+2044A_F	CTTCCGTGAGGACTGAATGAGACGGTC	60	235
G+2044A_R	GCAAATAATGAGCTTTCGAAGTTTCAGTGGA		
A-1512C_F	CAACCGCCGCGCCAGCGCCTTCTC	65	245
A-1512C_R	CCGCTACTTGGCCGTGTGACCGC		
C+2749T_F	GGACAGGGACCCACTTCACAC	44	390
C+2749T_R	GCTAACATATTTAATATTTATGTAC		

°C†: annealing temperature

in table 1 for 45 s, extension at 72⁰C for 45 s, followed by a final extension for 7 min at 72⁰C. PCR products were separated on a 2% agarose gel.

For genotyping of three SNPs rs20541, rs1881457 and rs847, 16 µl aliquot of PCR product was subjected to restriction enzyme digestion at 37 °C overnight with 10 U of *NlaIV*, *BstUI*, and *BsrGI* restriction enzymes, respectively according to the manufacturer's instructions (Fermentas, Burlington, Ontario). The resulting digested products were resolved on 3% agarose gels (Table 2).

Statistical Analysis

The associations between the genotype and allele frequencies in patients compared to controls were analyzed by computing the Pearson chi-square (χ^2) and odds ratio (OR 95% CI) using statistical software StatCalc EpiInfo package v.6 (Atlanta, GA).

RESULTS

The distribution of the IL-13 genotype and allele frequencies of SNPs rs20541, rs1881457 and rs847 were analyzed in asthma and AR patients sensitized to allergens and control individuals.

Table 2. Genomic Sequence polymorphisms Analysis of IL-13 Gene by RFLP

cDNA Coordinates of SNPs	rs #	AA change	Position	R. E	RFLP Fragments
Genomic Sequence polymorphisms Analysis of IL-13 Gene					
c.431G>A	rs20541	p.Gln144Arg	Exon 4	<i>NlaIV</i>	G=209,26 A=178,31,26
c.-1470A>C	rs1881457	NA	5'UTR	<i>BstUI</i>	A= 214,20,9,2 C=192,22,20,9,2
c.*695T>C	rs847	NA	3'UTR	<i>BsrGI</i>	T= 223, 142, 25, C= 223,167

Table 3. Genotypic frequencies of IL-13 polymorphisms in controls and atopic patients

Genotypes	Controls (120)	Patients (214)	p-value (χ^2)
Interleukin 13 G+2044A			
GG	47 (39.2)	81 (37.9)	0.24 (2.83)
GA	44 (36.7)	64 (29.9)	
AA	29 (24.1)	69 (32.2)	
Interleukin 13 A-1512C			
AA	97 (80.8)	123 (57.5)	<0.001 (19.06)
AC	18 (15)	64 (29.9)	
CC	5 (4.2)	27 (12.6)	
Interleukin 13 T+2749C			
TT	5 (4.2)	18 (8.4)	0.30 (2.37)
TC	16 (13.3)	31 (14.5)	
CC	99 (82.5)	165 (77.1)	

Table 4. Genotypic frequencies of IL-13 polymorphisms in controls, asthma and AR

Genotypes	Controls (120)	Asthma (108)	p-value (χ^2)	AR (106)	p-value (χ^2)
Interleukin 13 G+2044A					
GG	47 (39.2)	46 (42.6)	0.52 (1.29)	35 (33.0)	0.11 (4.27)
GA	44 (36.7)	32 (29.6)		32 (30.2)	
AA	29 (24.1)	30 (27.8)		39 (36.8)	
Interleukin 13 A -1512C					
AA	97 (80.8)	70 (64.8)	0.01 (8.80)	53 (50)	<0.001 (24.03)
AC	18 (15)	24 (22.2)		40 (37.7)	
CC	5 (4.2)	14 (13)		13 (12.3)	
Interleukin 13 T+2749C					
TT	5 (4.2)	10 (9.25)	0.21 (3.08)	8 (7.6)	0.19 (3.26)
TC	16 (13.3)	10 (9.25)		21 (19.8)	
CC	99 (82.5)	88 (81.5)		77 (72.6)	

The A-1512C SNP showed a marked difference in genotype frequencies of non-atopic controls and atopic patients [$p<0.001$ (19.06)] while no significant difference was found for G+2044A and T+2749C SNPs when the patients were compared to controls (Table 3).

Upon the stratification of the data according to diseases type, A-1512C SNP was significantly associated with both atopic conditions asthma and AR. In controls, 80.8% individuals had the homozygous wild type genotype compared to patients of asthma (64.8%) and AR (50%). The homozygous CC variant was observed in 4.2% controls relative to 13% in asthma [$p=0.01$ (5.76), OR 0.29 (0.09-0.91)] and 12.3% in AR [$p=0.02$ (5.03), OR 0.31 (0.09-0.98)]. No significant association of the T+2749C SNP was observed either in asthma [$p=0.21$ (3.08)] or in AR [$p=0.19$ (4.27)] (Table 4).

No significant difference in allelic frequencies was observed for G+2044A polymorphism in asthma

[$p=0.98$ (0.00), OR 1.00 (0.68-1.48)] while in AR a significant difference in allele frequencies relative to controls was observed [$p=0.04$ (3.98), OR 1.46 (0.99-2.15)] (Table 5).

DISCUSSION

In the current study we evaluated the role of three SNPs rs20541 (G+2044A), rs1881457 (A-1512C) and rs847 (T+2749C) in the IL-13 gene in allergic asthma and AR patients sensitized to pollens. We did not find association with the G+2044A and T+2749 polymorphisms with both asthma and AR. Although association of A allele in case of G+2044A SNP was observed in a meta-analysis of six populations with AR.¹⁵ Instead of this SNP in Pakistani asthma and AR patients, strong association was observed with the A-1512C polymorphism which might be due to diverse genetic heterogeneity among different populations.

Table 5. Allele frequencies of IL-13 polymorphisms

Alleles	Controls	Asthma	p-value (χ^2)	OR (95% CI)	AR	p-value (χ^2)	OR (95% CI)
Interleukin 13 G+2044A							
G	138 (57.5)	124 (57.4)	0.98 (0.00)	1.00 (0.68-1.48)	102 (48.1)	0.04 (3.98)	1.46 (0.99-2.15)
A	102 (42.5)	92 (42.6)			110 (51.9)		
Interleukin 13 A-1512C							
A	212 (88.3)	164 (75.9)	<0.001	2.40 (1.41-4.09)	146 (68.9)	<0.001	3.42 (2.04-5.76)
C	28 (11.7)	52 (24.1)	(12.10)		66 (31.1)	(25.89)	
Interleukin 13 T+2749C							
T	26 (10.8)	30 (13.9)	0.32 (0.99)	0.75 (0.41-1.37)	37 (17.5)	0.04 (4.11)	0.57 (0.32-1.02)
C	214 (89.2)	186 (86.1)			175 (82.5)		

IL-13 Gene Polymorphisms and Atopic Asthma and Rhinitis

For the first time, in the British population, evidence was provided for involvement of A-1512C SNP in IL-13 gene with the total IgE levels but functional effect of this polymorphism was not studied.¹⁸

Our study supported the fact that this SNP alone was enough to change the IgE levels as we found significant association of A-1512C with both asthma and AR.

From the results of the present study, we do believe that the polymorphisms of IL-13 gene are playing an important role in atopic diseases and it might be a good target for the therapeutic intervention. In the previous studies, it was also observed that high levels of IL-13 were expressed in atopic and non-atopic asthma^{19, 20}, atopic dermatitis²¹, and allergic rhinitis.²² IL-13 is produced by Th2 cells and is important in the pathophysiological allergic reactions and its effects were the hallmark features of the asthma such as increased secretion of the mucus, high levels of IgE, AHR, eosinophil recruitment.²³⁻²⁶ Therefore, IL-13 has an important role in inflammation and is responsible for driving the epithelial cells into a hypersecretory phase which is a cause of increased allergic reactions in the airways.²⁷ IL-13 also has an important role in the contraction of the smooth airway muscles and in the development of the allergic airway hyper-reactivity.

In conclusion we found a strong association of A-1512C polymorphism with both asthma and AR in Pakistani patients. Further studies conducted on the functional level are required to contribute to the existing knowledge on these respiratory diseases.

REFERENCES

1. Prieto J, Lensmar C, Roquet A, van der Ploeg I, Gigliotti D, Eklund A, et al. Increased interleukin-13 mRNA expression in bronchoalveolar lavage cells of atopic patients with mild asthma after repeated low dose allergen provocations. *Respir Med* 2000; 94(8):806-14.
2. Bodey KJ, Semper AE, Redington AE, Madden J, Teran LM, Holgate ST, et al. Cytokine profiles of BALT cells and T-cell clones obtained from human asthmatic airways after local allergen challenge. *Allergy* 1999; 54(10):1083-93.
3. Truyen E, Coteur L, Dilissen E, Overbergh L, Dupont LJ, Ceuppens JL, et al. Evaluation of airway inflammation by quantitative Th1/Th2 cytokine mRNA measurement in sputum of asthma patients. *Thorax* 2006; 61(3):202-8.
4. Park SW, Jangm HK, An MH, Min JW, Jang AS, Lee JH, et al. Interleukin-13 and interleukin-5 in induced sputum of eosinophilic bronchitis: comparison with asthma. *Chest* 2005; 128(4):1921-7.
5. Pawankar RU, Okuda M, Hasegawa S, Suzuki K, Yssel H, Okubo K, et al. Interleukin-13 expression in the nasal mucosa of perennial allergic rhinitis. *Am J Respir Crit Care Med* 1995; 152(6 Pt 1):2059-67.
6. Ghaffar O, Laberge S, Jacobson MR, Lowhagen O, Rak S, Durham SR, et al. IL-13 mRNA and immunoreactivity in allergen-induced rhinitis: comparison with IL-4 expression and modulation by topical glucocorticoid therapy. *Am J Respir Cell Mol Biol* 1997; 17(1):17-24.
7. McKenzie GJ, Bancroft A, Grecis RK, McKenzie AN. A distinct role for interleukin-13 in Th2-cell-mediated immune responses. *Curr Biol* 1998; 8(6):339-42.
8. Grunig G, Warnock M, Wakil AE, Venkayya R, Brombacher F, Rennick DM, et al. Requirement for IL-13 independently of IL-4 in experimental asthma. *Science* 1998; 282(5397):2261-3.
9. Miyahara S, Miyahara N, Matsubara S, Takeda K, Koya T, Gelfand EW. IL-13 is essential to the late-phase response in allergic rhinitis. *J Allergy Clin Immunol* 2006; 118(5):1110-6.
10. Heinzmann A, Mao XQ, Akaiwa M, Kreomer RT, Gao PS, Ohshima K, et al. Genetic variants of IL-13 signalling and human asthma and atopy. *Hum Mol Genet* 2000; 9(4):549-59.
11. Bottema RWB, Nolte IM, Howard TD, Koppelman GH, Dubois AEJ, de Meer G, et al. Interleukin 13 and Interleukin 4 Receptor- α Polymorphisms in Rhinitis and Asthma. *Int Arch Allergy Immunol* 2010; 153(3):259-67.
12. Graves PE, Kabesch M, Halonen M, Holberg CJ, Baldini M, Fritsch C, et al. A cluster of seven tightly linked polymorphisms in the IL-13 gene is associated with total serum IgE levels in three populations of white children. *J Allergy Clin Immunol* 2000; 105(3):506-13.
13. Yang H, Dong H, Dai Y, Zheng Y. Association of interleukin-13 C-1112T and G+2044A polymorphisms with asthma: ameta-analysis. *Respirology* 2011; 16(7):1127-35.
14. Vladich FD, Brazille SM, Stern D, Peck ML, Ghittoni R, Vercelli D. IL-13 R130Q, a common variant associated with allergy and asthma, enhances effector mechanisms essential for human allergic inflammation. *J Clin Invest* 2005; 115(3):747-54.
15. Bunyavanich S, Shargorodsky J, Celedón JC. A meta-analysis of Th2 pathway genetic variants and risk for allergic rhinitis. *Pediatr Allergy Immunol* 2011;

- 22(4):378-87.
16. Micheal S, Minhas K, Ishaque M, Ahmed F, Ahmed A. Promoter polymorphisms of the CD14 gene are associated with atopy in Pakistani adults. *J Investig Allergol Clin Immunol* 2011; 21(5):394-7.
 17. Sambrook J, Russell DW, Sambrook J. The condensed protocols from Molecular cloning: a laboratory manual. Cold Spring Harbor (NY): Cold Spring Harbor Laboratory Press; 2006.
 18. Maier LM, Howson JM, Walker N, Spickett GP, Jones RW, Ring SM, et al. Association of IL13 with total IgE: evidence against an inverse association of atopy and diabetes. *J Allergy Clin Immunol* 2006; 117(6):1306-13.
 19. Humbert M, Durham SR, Kimmitt P, Powell N, Assoufi B, Pfister R, et al. Elevated expression of messenger ribonucleic acid encoding IL-13 in the bronchial mucosa of atopic and nonatopic subjects with asthma. *J Allergy Clin Immunol* 1997; 99(5):657-65.
 20. Huang SK, Xiao HQ, Kleine-Tebbe J, Paciotti G, Marsh DG, Lichtenstein LM, et al. IL-13 expression at the sites of allergen challenge in patients with asthma. *J Immunol* 1995; 155(5):2688-94.
 21. Akdis M, Akdis CA, Weigl L, Disch R, Blaser K. Skin-homing, CLA₊ memory T cells are activated in atopic dermatitis and regulate IgE by an IL-13-dominated cytokine pattern: IgG4 counter-regulation by CLA₊ memory T cells. *J Immunol* 1997; 159(9):4611-9.
 22. Katagiri K, Itami S, Hatano Y, Takayasu S. Increased levels of IL-13 mRNA, but not IL-4 mRNA, are found in vivo in peripheral blood mononuclear cells (PBMC) of patients with atopic dermatitis (AD). *Clin Exp Immunol* 1997; 108(2):289-94.
 23. Emson CL, Bell SE, Jones A, Wisden W, McKenzie AN. Interleukin (IL)-4-independent induction of immunoglobulin (Ig)E, and perturbation of T cell development in transgenic mice expressing IL-13. *J Exp Med* 1998; 188(2):399-404.
 24. Zurawski G, de Vries JE. Interleukin-13, an interleukin-4 like cytokine that acts on monocytes and B cells, but not on T cells. *Immunol Today* 1994; 15(1):19-26.
 25. Horie S, Okubo Y, Hossain M, Sato E, Nomura H, Koyama S, et al. Interleukin-13 but not interleukin-4 prolongs eosinophil survival and induces eosinophil chemotaxis. *Intern Med* 1997; 36(3):179-85.
 26. Wills-Karp M, Luyimbazi J, Xu X, Schofield B, Neben TY, Karp CL, et al. Interleukin-13: central mediator of allergic asthma. *Science* 1998; 282(5397):2258-61.
 27. Danahay H, Atherton H, Jones G, Bridges RJ, Poll CT. Interleukin-13 induces a hypersecretory ion transport phenotype in human bronchial epithelial cells. *Am J Physiol Lung Cell Mol Physiol* 2002; 282(2):L226-36.