

REVIEW ARTICLE

Iran J Allergy Asthma Immunol

December 2021; 20(6):647-671.

Doi: 10.18502/ijaai.v20i6.8016

Effectiveness of Coronavirus Vaccines against Syndrome Coronavirus 2 (SARS-CoV-2) and Its New Variants

Afshin Abdi Ghavidel¹, Mahbubeh Rojhannezhad², Bahram Kazemi³, and Mojgan Bandehpour⁴

¹ Student Research Committee, School of Advanced Technologies in Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

² Department of Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran

³ Cellular and Molecular Biology Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁴ Department of Medical Biotechnology, School of Advanced Technologies in Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Received: 12 January 2021; Received in revised form: 26 June 2021; Accepted: 6 July 2021

ABSTRACT

The widespread outbreak of coronavirus disease 2019 in late 2019 caused many people worldwide to die or suffer from certain clinical complications even after the recovery. The virus has many social and economic adverse effects. Studies on severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) have specified that spike, surface glycoprotein antigen, is considered as a major target to stimulate the immune system. This glycoprotein binds to the angiotensin-converting enzyme 2 on the surface of human cells especially lung epithelial cells and facilitates the virus entry. Therefore, the immune response stimulated by vaccination targeting this antigen may cause immunity against the whole virus. Currently, many companies are working on SARS-CoV-2 vaccines. They include 'traditional' vaccines like attenuated or inactivated virus platforms as well as the brand-new generations of vaccines such as viral vector-based, subunit, nucleic acid-based, and virus-like particle vaccines. Certainly, each vaccine platform presents several advantages and disadvantages affecting its efficacy and safety which is the main topic of this paper.

Keywords: COVID-19; COVID-19 vaccines; SARS-CoV-2; SARS-CoV-2 spike protein; SARS-CoV-2 variants

INTRODUCTION

Alpha (α), beta (β), gamma (γ), and delta (δ) coronaviruses constitute the subgroups of the coronavirus family. Until the outbreak of SARS-CoV

in Guangdong, China, these viruses were thought to infect only animals.¹ To date, seven of these viruses have been identified in humans. HCoV-229E and HCoV-NL63 belong to α -coronaviruses, while HCoV-OC43, HCoV-HKU1, MERS-CoV (Middle East respiratory syndrome coronavirus), SARS-CoV (severe acute respiratory syndrome coronavirus), and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) belong to β -coronaviruses. However, SARS-CoV-2, unlike other beta coronaviruses, has exhibited

Corresponding Author: Mojgan Bandehpour, PhD;
Department of Medical Biotechnology, School of Advanced Technologies in Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Tel: (+98 21) 2387 2552, Fax: (+98 21) 2243 9956, E-mail: Bandehpour@gmail.com m.bandehpour@sbm.ac.ir

dramatically higher numbers of person-to-person spread.

Almost two decades later, after the first detection of beta coronavirus infection in China, another pathogenic coronavirus outbreak has been identified in Wuhan, China. In December 2019, a kind of pneumonia was diagnosed, which was caused by an unknown pathogen.² Following the whole genome sequencing of that virus and the similarity of its genome to the SARS-CoV, it was named SARS-CoV-2.³ Also, the disease caused by SARS-CoV-2 was called coronavirus disease 2019 (COVID-19). COVID-19 is known as a “thousand faces disease”, because of the different symptoms of patients affected by COVID-19.⁴ Ghazanfari et al in 2021 reported different biomarkers such as age, sex, underlying diseases, blood oxygen pressure, complete blood count along with C-reactive protein, lactic dehydrogenase, procalcitonin, D-dimer, and interleukin-6 evaluation, which are associate with severity and mortality of COVID-19.⁵

Response to the SARS-CoV-2 takes a different amount of time depending on the type of response. Immunoglobulin (Ig) M and IgA are the short-lived antibody responses against SARS-CoV, while IgG is a response against SARS-CoV that lasts a bit longer (less than six months and approximately twelve months, respectively).⁶ Memory T cells against SARS-CoV can cause the long-term protective response to the SARS-CoV.⁷ Although there is a similarity between the spike protein of SARS-CoV and SARS-CoV-2, neutralizing antibodies against the SARS-CoV spike protein cannot bind to the SARS-CoV-2 spike protein,⁸ which means immunity against SARS-CoV cannot produce protective immunity against SARS-CoV-2.

There are two types of vaccines against SARS-CoV, including DNA vaccines (Spike gene) and inactivated vaccines which are in phase I clinical trials, and four vaccines against MERS-CoV, including DNA vaccines (Spike gene), and viral vector-based vaccines, which are in phase I clinical trial⁹ and also there are some more vaccines in preclinical phases. However, those vaccines have not elicited a specific, long-term, and adequate immune response and protectivity against those viruses. Based on the similarity between these viruses and SARS-CoV-2, the results of those vaccines have been used for developing SARS-CoV-2 vaccines. As we discuss later in this article, based on the published information, spike protein has been used as an antigen in developing SARS-CoV-2 vaccines.

The newer generation of vaccines, such as the viral vector-based vaccines, nucleic acid-based vaccines, and virus-like particles (VLPs) vaccines, have a chance to prove themselves in SARS-CoV-2 outbreak, as most of them have not been licensed for human use.⁷ To pass the tough quarantine conditions and return to previous patterns of working, schooling, and socializing, the traditional vaccines along with the new generation vaccines should effectively control the COVID-19 pandemic.

SARS-CoV-2 Genome

SARS-CoV-2 has a single-stranded positive-sense RNA genome with 29891 nucleotides (GenBank no. MN908947) in length which codes 9860 amino acids. Polyprotein 1a/1ab (pp1a/pp1ab) is directly transcribed from the genomic RNA between ORF1a and ORF1b (open reading frames) and encodes nonstructural proteins (nsps). Open reading frames (ORFs) on one-third of the genome near the 3' terminus are translated to a single long polyprotein. This polyprotein consists of at least four main structural proteins: the spike (S), envelope I, membrane (M), and nucleocapsid (N) proteins, and accessory proteins (involved in immune evasion).¹⁰ The SARS-CoV-2 sequence has a homology of 77.5% and 50% with SARS-CoV, and MERS-CoV, respectively.¹¹ The error rate of the SARS-CoV-2 replication was estimated to be between 10^{-6} to 10^{-7} errors per nucleotide,¹² and several nucleotide deletions and variations at different positions throughout the entire genome of SARS-CoV-2 were detected.¹³

Spike Protein

Spike consists of a glycosylated protein expressed in large numbers on the surface of SARS-CoV-2, thus giving it the characteristic ‘crown’ appearance. This protein has a role in binding the virus to the host cell receptor named angiotensin-converting enzyme 2 (ACE2). Attaching spike to ACE2 initiates the entrance of the virus to the host cells,¹⁴ then TM protease serine 2 (TMPRSS2) activates S protein and promotes the entrance of the virus into the host cell where the viral RNA replicate, and also translates to polyproteins.¹⁵ Although there is considerable similarity, the spike protein of SARS-CoV-2 binds to ACE2 with a higher affinity than the spike protein of SARS-CoV.¹⁶

Spike protein has three main domains consisting of an extracellular N-terminus domain (NTD), a

transmembrane I domain, and a short intracellular C-terminal domain (C-domain) with 180–200 kDa overall size and 1273 amino acids.¹⁷ The first 13 amino acids of the S protein are signal peptides located at the N-terminus. The S1 (14–685 residues) and S2 subunits (686–1273 residues) follow the signal sequence. These two subunits are involved in virus binding attachment and entry into the host cells.¹⁸ The receptor-binding domain (RBD) is placed in the S1 subunit and binds to the ACE2.¹⁹ Fusion peptide (FP) region and two heptads repeat regions, HR1 and HR2, are placed in the S2 subunit.²⁰ Moreover, there is a unique furin-like cleavage site (FCS) in the S protein of SARS-CoV-2 (682 – 685, with RRAR sequence) located between S1 and S2 subunits, which is absent in the other lineage β CoVs.²¹ Serine proteases of target cells are activated immediately after binding of RBD to ACE2, then cleavage of S protein into S1 and S2 subunits happens.²² This cleavage mediates S protein activation, membrane fusion, and releasing the viral package into the host cytoplasm.

Theoretically, based on the central role of spike protein to enter into host cells, it seems that it has to be conserved, and natural mutations may not change it. Therefore, the companies that produce vaccines apply the whole S protein or partial S protein as an antigen to trigger an immune response against the SARS-CoV-2. Analyzing the recovered COVID-19 patients' blood showed neutralizing antibodies (nAb) against the SARS-CoV-2 target RBD.²³ Along with nAb, T cells, especially CD4⁺ and CD8⁺, also respond to the S protein. Recently, scientists reported that there are isolated antibodies from human blood that targets non-RBD epitopes²⁴ such as NTD²⁵ and S2.²⁶ Except for RBD, the whole surface of the S protein is covered by glycans, which reduce the immunogenicity of S protein. RBD is the only subunit of S protein that has no glycan cover; therefore, the immune system can respond to that epitope appropriately²⁷ which means humoral²⁸ and cellular²⁹ immune systems respond to the RBD effectively.

Antibodies against NTD cannot neutralize the virus completely, inhibit viral fusion, and virus infection.²⁶ Due to this fact, NTD-based vaccines have weaker protective efficacy than RBD-based vaccines.³⁰ Antibodies that target the S2 subunit can neutralize the virus and prevent infection.³¹ However because of glycans coverage on subunits,³² the accessibility of these epitopes for the immune system is reduced which

means S2-based vaccines produce a lower immune response than RBD.

The N protein and the M protein can generate the total T cell response.³³ Due to small ectodomains for immune system recognition and small molecular sizes of M and E proteins, they are less immunogenic than S proteins and do not produce a protective immune response against the virus.³⁴ Unlike M and E proteins, N protein is highly immunogenic,³⁵ but N protein-based vaccines do not produce adequate immune response against SARS-CoV³⁶ nor SARS-CoV-2.³⁴

S1 has several desired properties including metastability of the recombinant S1 protein, amenability to transform from pre-fusion to a post-fusion conformation, accessibility for immune recognition system, and last but not least neutralizing properties by targeting it, which make it the immunodominant antigen for the immune system.³⁷ Although, this subunit has many advantages that make it an excellent target for the immune system, its properties need to be improved. Two proline substitutions (2P), substitution at the cleavage sites, and change RRAR to SRAG or QQAQ are examples of those improvements.³⁷ S-2P and SRAG or QQAQ are used in the Janssen vaccine³⁸ and the Novavax vaccine,³⁹ respectively, to stabilize the S protein in its pre-fusion conformation.

SARS-CoV-2 Immune Response

The first line of response to the SARS-CoV-2, following the detection of SARS-CoV-2 components in the body, is innate immune systems. These responses recruit macrophages and monocytes, which start the nonspecific defense mechanisms. These cells release cytokines and trigger adaptive immune cells. Subsequently, the adaptive immune system, *i.e.*, B and T cells, initiate a specific response to virus antigens.

Innate Immunity

The innate immune response to SARS-CoV-2 infection starts with recognizing RNA or other molecular structures of the virus by immune cells. Viral RNA is detected by pathogen-associated molecular patterns (PAMPs), Toll-like receptor 8 (*TLR8*), *TLR7*, and *retinoid-inducible gene (RIG)/ melanoma differentiation-associated 5 genes (MDA5)*. Moreover, the damage-associated molecular patterns (DAMPs) are in charge of detecting the virus ATP, DNA, and ASC oligomers.²¹ Subsequently, several signaling

pathways and, ultimately, transcription factors will be activated.

During SARS-CoV-2 infection, secretion of pro-inflammatory cytokines and chemokines, including IL-1 β , IL-6, IFN γ , MCP1, and IP-10, into the blood are elevated.⁴⁰ Then, immune cells absorb the site of cytokines and chemokines secretion.⁴¹ SARS-CoV-2 needs to induce aberrant inflammatory responses to protect itself. Multiple viral structural and nonstructural proteins of SARS-CoV-2 prevent the onset of interferon responses by blocking the interferon signaling pathway, which subverts the body's innate antiviral cytokine responses.⁴²

Adaptive Immunity

Both T and B cells can be detected in the COVID-19 patient's blood after seven days of the onset of symptoms. Two types of T cells are essential in adaptive immune responses to viral infections. CD8⁺ T cells destroy the viruses inside the host cells, and CD4⁺ T cells produce cytokines to induce CD8⁺ T cells and B cells to respond.⁴¹

In blood samples of COVID-19 patients, the number of CD8⁺ T cells was more remarkable than CD4⁺ T cells.²¹ Due to the critical roles of T cells in preventing the reinfection with SARS-CoV-2, a sufficient number of memory T-cells is essential.⁴³ Zhang, F et al have demonstrated that specific memory T-cell responses can be induced and reactivated by several peptides derived from SARS-CoV-2, including S and M proteins, in most COVID-19 patients. Therefore, these peptides are among the potential targets for vaccine development.⁴⁴ However, activating the IFN-mediated pathways of T cells can be an alternative way to deal with the virus.

IgM, IgA, and IgG responses were also determined in most patients with COVID-19, suggesting that antibodies act in the generation production of protective immune responses against SARS-CoV-2.⁴⁵ Neutralizing antibodies are the primary humoral immune responses against SARS-CoV-2.⁴⁶ Since the high concentration of neutralizing antibodies correlate with mild COVID-19 symptoms, triggering the humoral immune responses might be a good strategy for providing future protection against SARS-CoV-2 reinfection.⁴⁷ However, some studies demonstrated a correlation between COVID-19 severity and SARS-CoV-2-specific T cells, which raises an important question regarding vaccine protection in humans.^{48,49}

Immune Respond to SARS-CoV-2

SARS-CoV-2 can trigger the immune system in two ways. One of the ways is virus's phagocytosis by antigen-presenting cells (APCs) and T-cell activation via presenting virus antigens on major histocompatibility complex (MHC) class II. The other way is through the synthesis of virus proteins in infected cells. In this case, these proteins might be presented to CD8⁺ cells using MHC class I or released from the cells and phagocytosed by APCs and presented on MHC class II. Dead vaccines and inactivated vaccines trigger the immune system similar to the native virus. However, viral vector-based vaccines and nucleic acid-based vaccines cannot directly stimulate the immune system. They must enter the cells. Then, specific SARS-CoV-2 related peptides are released and presented on the cell surface by MHC class I. Moreover, APCs phagocyte peptides which be released from the cell and present it on MHC class II. Subunit vaccines and VLPs vaccines can stimulate the immune system as well, through APCs. Presented antigens on MHC class I would activate CD8⁺ T cells and CD4⁺ T cells. CD8⁺ T cells can destroy or damage infected cells directly. CD4⁺ T cells respond to SARS-CoV-2 indirectly by activating both B and CD8⁺ T cells. T and B memory cells play crucial roles in immune system response, and as long as they exist, resulting immune system can remain (Figure 1).

SARS-CoV-2 Suggested Vaccines

Utilizing an effective vaccine would be a good idea to achieve herd immunity against SARS-CoV-2 in the population.⁵⁰ The specific roles of antibodies and T cells in protection against SARS-CoV-2 infection in humans are yet unknown. Howbeit, neutralizing antibodies have shown a protective role in rhesus macaques.⁵¹ Various candidate vaccines, including nucleic acid vaccines, inactivated virus vaccines, live attenuated vaccines, protein or peptide subunit vaccines, and viral-vectored vaccines, are being developed. Vaccines must have high safety and efficacy, which means that the side effects of vaccines should not outweigh the severity of natural diseases, and they should effectively stimulate the immune responses, respectively. Moreover, given the number of detected mutations in the SARS-CoV-2 virus,⁵² vaccines should be designed in such a way that provides immunity not only against SARS-CoV-2 wild-type virus but also SARS-CoV-2 new variants. Table 1

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summarizes different vaccines and demonstrates examples of approved vaccines for human use in each vaccine type.

Candidate vaccines are being tested in preclinical and clinical trials to determine their safety and efficacy. The preclinical trial has two stages, and in each one, the toxicity and teratogenicity of vaccines are tested. In the first stage, vaccines are tested on non-primate animals; if the result is acceptable, vaccines enter the second stage and are tested on non-human primates. Subsequently, the next step is studying the drugs in clinical trials. Clinical trials consist of four phases

and in each phase, different aspects of vaccines are tested on volunteers. In the phase I trial, vaccines are tested on a limited number of healthy human volunteers. In this phase, the appropriate dose and the possible side effects of vaccines are determined. In the phase II trial, the vaccines' efficacy is tested on a small number of human volunteers who are naturally exposed to the pathogen or are challenged with the pathogen deliberately. Finally, in the phase III trial, vaccines are tested on many healthy volunteers who live in an endemic area or are challenged intentionally with pathogens.

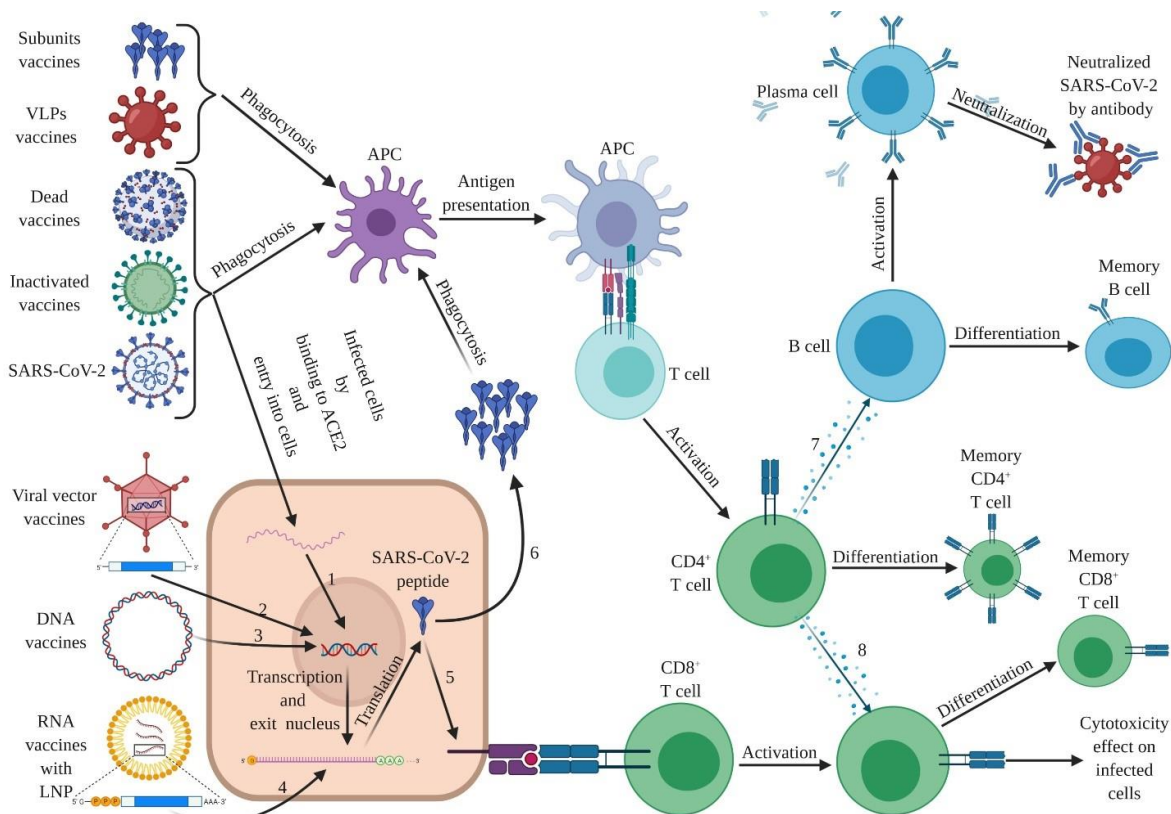


Figure 1. Downstream pathways as a result of immune system stimulation through each type of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccines and also result of native SARS-CoV-2 infections. (1. SARS-CoV-2 genome will reverse transcribed to DNA following cell infection and entry to the nucleus. 2. Viral vector-based vaccines deliver DNA encoding specific SARS-CoV-2 peptides to vaccines cells. 3. DNA vaccines not only must cross the cell membrane but also, they should enter the nucleus. 4. mRNA vaccines that encode the specific SARS-CoV-2 peptides cross the cell membrane, and subsequently, the peptide translates into the cytoplasm. 5. SARS-CoV-2 peptide is processed in the infected cell and subsequently present on the MHC class I. (In nucleic acid base vaccines and viral vector-based vaccines, vaccinees cells present only one type of SARS-CoV-2 peptide, but in a native infection or whole vaccine, all the virus peptides present on the cell surface.) 6. CD4⁺ T cells release cytokines to activate CD8⁺ cells. 7. CD4⁺ T cell release cytokines to activate B cells.). Antigen presentation on MHC class II results in CD4⁺ T cell activation, which stimulates CD8⁺ T and B cells. Three different types of memory cells will be produced, triggering the immune system in reinfection or natural infections. Figure created with Bio Render (<https://biorender.com>).

Table 1. Some examples of different types of approved vaccines before the COVID-19 outbreak.

Types of Vaccines	Description	Some examples of previously approved vaccines
Live attenuated vaccines	Decrease pathogenicity but still immunogenic; may still replicate.	Viral: measles, mumps, rubella, vaccinia, varicella, zoster, yellow fever, rotavirus, intranasal influenza (LAIV), oral polio Bacterial: BCG, oral typhoid
Inactivated (dead) vaccines	Block replication by killing or inactivation of the pathogen but still immunogenic.	Viral: polio, hepatitis A, rabies, Bacterial: pertussis, typhoid, cholera, plague
Nucleic acid-based vaccines	Naked DNA or mRNA containing special sequences of the pathogen can stimulate an immunogenic response to the whole pathogen.	N/A
Viral vector-based vaccines	A virus-based vector containing recombinant DNA of pathogen antigen which infected vaccinees' cells and replicates transcribed and translated continually in there.	rVSV-ZEBOV Dengvaxia
Subunit vaccines	Synthesized or purified pathogen protein, peptides or polysaccharides.	hepatitis B, influenza, acellular pertussis, human papillomavirus, anthrax
Viral-like particles vaccines	Purified from natural sources or synthesized using recombinant DNA methods.	HBV, Gardasil, Cervarix

COVID-19: coronavirus disease 2019; BCG: Bacillus Calmette–Guérin (BCG); HBV: Hepatitis B vaccine

In each trial phase, if the results indicate unsafe or unacceptable efficacy of the vaccines, the trial will be stopped, and the vaccines will be discarded. If the vaccine successfully passes all these three phases of the trial, it will be licensed and distributed for human use. However, in phase IV, the safety and efficacy of vaccines are determined in the real world for rare adverse events. Therefore, if consequences of vaccines occur in real-life licenses of them can be dismissed.⁵³

By May 2021, multiple phases of 3 clinical trials for SARS-CoV-2 vaccines are being performed in different parts of the world. According to the food and drug administration (FDA), at least 50% of vaccinated people must be immune against SARS-CoV-2 to consider SARS-CoV-2 vaccine as efficacious.⁵⁴ According to the world health organization (WHO), twenty-three SARS-CoV-2 candidate vaccines will be in phase 3 clinical trial or would pass this stage successfully, up to June 24th 2021.⁵⁵ Each approach has its own advantages and disadvantages, which will be discussed here.

SARS-CoV-2 Vaccine Targets

Vaccines must stimulate immune responses that generate long-lasting B and T memory cells against the related pathogen. Vaccines contain immunogenic but

not pathogenic epitopes of pathogens. Therefore, they can induce immune responses without infecting the hosts.⁵⁶ The higher affinity of SARS-CoV-2' S protein to ACE2 compared to other coronaviruses is the reason for increased the infection rate of SARS-CoV-2. Several vaccines were developed by using the surface antigen that can stimulate immune response sufficiently.^{57,58} Human-neutralizing antibodies against S protein have a crucial role in protecting against SARS-CoV.⁵⁹ S2 subunit is structurally conserved and has 88% sequence homology with the S2 domain of SARS-CoV.⁶⁰ The same result was detected following the analysis of neutralized antibodies against SARS-CoV-2 S protein.⁶¹ Therefore, using the whole S glycoprotein, or a part of it, has increasingly gained attention for the vaccination purpose.

Different Type of Vaccines

Currently, there are several methods to develop a vaccine. Previously, design of vaccines was based on using live, attenuated, inactivated organisms or pathogenic products (toxoids). With recent advancements in biotechnology, new types of vaccines, namely subunit vaccines which purified from the natural organism, or synthesized *in vitro*, have been introduced. Lately, DNA and RNA vaccines, together

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with VLPs vaccines, have also been presented. DNA vaccines can be either recombinant vectors or naked DNA vaccines.⁶² Each vaccine type has its advantages and disadvantages (Table 2). Therefore, to get the best immunity against the pathogen, the method used to construct vaccines is of great importance.

The “front runner candidates” are provided by companies or research institutes in China, Russia, Germany, the United Kingdom, and the United States. In the following sections, vaccines that are in phase 3 of clinical trials or have already passed this phase at the time of writing this article will be reviewed (Table 3).

Table 2. Advantages and disadvantages of different types of severe acute respiratory syndrome coronavirus 2 vaccines

Type of vaccine	Advantages	Disadvantages
Live attenuated vaccines	Immunogenicity similar to natural infection (extensive immunity with both humoral and cellular respond) ⁶³ A single dose without a booster often is enough ⁶⁴ Cost-effective ⁶⁵ Present multiple viral proteins ⁶⁶ Epitopes with correct conformation ⁶⁶	The potential transmissibility of the live attenuated vaccine ⁶⁷ Less storage stability and necessity of cold chain transport ⁶⁸ Revert to the virulent forms ⁶⁹ Possibility of contamination during preparations ⁷⁰
Inactivated (dead) vaccines	Possibility of reversion close to zero ⁷¹ Immunogenicity similar to natural infection (extensive immunity with both humoral and cellular respond) ⁷² Stable and no need for cold chain ⁷³ Virus contamination close to zero ⁷¹	Minor immune system stimulation ⁷⁴ Booster and several doses are needed ⁷⁵ Less effective against intracellular pathogens ⁷⁶
Nucleic acid-based vaccines (DNA)	Adjuvant rule of bacterial sequences in plasmids ⁷⁷ Almost stable in the body ⁷⁸ Immunogenicity similar to natural infection (extensive immunity with all humoral, cellular, and innate respond) ⁷⁹ Easy to manipulate and modify ⁸⁰ Ready platform ⁸¹	Lower efficacy of cytotoxic T lymphocytes (CTLs) response ⁸² Lower titer, avidity, and longevity of antibody ⁸³ Possibility of integration and stimulate tumorigenic ⁸⁴ Need nuclear delivery ⁸⁴ Need promoter ⁸⁴ Present just one antigen of pathogens ⁷⁸ Wrong post-translational modifications ⁷⁸
Nucleic acid-based vaccines (mRNA)	Cost-effective ⁸⁵ Fast production ⁸⁵ No infection in the vaccine ⁷⁸ No integration to the genome ⁷⁸ Simple production ⁸⁶ Immunogen with 94–95% efficacy ^{87,88} Ready platform ⁸¹ Room temperature storage ⁸⁹	Present just one antigen of pathogens ⁷⁸ Wrong post-translational modification ⁷⁸ Toxicity ⁹⁰ Autoimmunity response ⁹⁰ Low-temperature storage requirements ⁹¹
Viral vector-based vaccines	High-efficiency gene transduction ⁹² Specific delivery of genes to target cells ⁹² Both humoral and cellular respond ⁹³ Robust immune responses ⁹² Increased cellular immunity ⁹² Do not have the ultracold storage temperature requirements ⁹¹	Pre-existing antiviral vector immunity ⁹⁴ Hepatotoxicity ⁹⁵ Lower efficacy than mRNA vaccines (92-70%) ^{96,97}
Subunit vaccines	No possibility of infections ⁹⁸ Stimulate T and B responses by modifying recombinant peptides ⁹⁹ Stable and easy to transport ¹⁰⁰	Expensive ¹⁰¹ Tough to purify ¹⁰¹ Lower immunogenicity ¹⁰² Delivery system and adjuvant are needed ¹⁰³⁻¹⁰⁵
Viral-like particles vaccines	No possibility of infections ¹⁰⁶ Ease of production ¹⁰⁷ Both humoral and cellular immune responses ¹⁰⁸ More immunogenic than linear peptides ¹⁰⁷ Self-adjuvating properties ¹⁰⁹	Expensive ¹¹⁰

Table 3. Different types of SARS-CoV-2 vaccines which are in phase 3 or 2/3 clinical trial or past these phases

Type of vaccine	Commercial Name	Generic Name	Name of Manufacturer	Address of Manufacturer	Doses	Explanations	Reference
Inactivated vaccine	CoronaVac	Sinovac	Sinovac Life Sciences Co., Ltd.	China	2	Inactivate with aluminum hydroxide	111
	BBIBP-CorV	Sinopharm	Wuhan Institute of biological products and Beijing Institute of biological products	China	2		112
	Covaxin (BBV152)	Bharat	Bharat biotech in collaboration with the Indian council of medical research.	India	2		55
	QazVac	QazCovid	Research institute for Biological safety problems	Kazakhstan	2		55
	-	Inactivated SARS-CoV-2 vaccine	Chinese Academy of Medical Sciences	China	2	Inactivated in Vero cells	55
	-	Minhai	Beijing Minhai Biotechnology Co	China	1,2 or 3	Inactivated in Vero cells	55
	-	Shenzhen Kangtai Biological Products Co	Shenzhen Kangtai biological products Co	China	2	Inactivated in Vero cells	55
-	Valneva	Valneva	France	2		55	
DNA vaccines	-	Zydus Cadila	Zydus Cadila	India	3	Full-length spike (S) protein	113
RNA vaccines	Moderna	Moderna, TAK-919 and mRNA-1273	Moderna, the United States national institute of allergy and infectious diseases (NIAID), and the biomedical advanced research and development authority (BARDA).	United States	2	LNP- mRNA-nano encoded full-length S protein	114
	Comirnaty	Pfizer	BioNTech, Fosun Pharma/Pfizer	Germany	2	LNP-mRNAs-nano encoded full-length S protein	115
	-	CVnCoV	CureVac AG	Germany	2	mRNA-LNP encoding the full-length S protein	55
	ARCoV	Walvax	Academy of Military Science (AMS), Walvax Biotechnology and Suzhou sbogen biosciences	Mexico	2	mRNA-LNP encoding the receptor-binding domain (RBD)	116

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Viral vector vaccines	Vaxzevria, Covishield	nCoV-19, ChAdOx1-S, AstraZeneca	Oxford University and AstraZeneca	England	2	ChAdOx1-S expressing the full-length S protein	117
	-	CanSino	CanSino Biological Inc./ Beijing institute of biotechnology	China	2	Ad-5 Vector expressing the full-length S protein	118
	Sputnik V	Gam-COVID-Vac	Gamaleya research institute	Russian	2	rAd26-S+rAd5-S both expressing the full-length S protein	119
	Janssen	Ad26.COVID.2.S and Johnson & Johnson	Janssen vaccines in Leiden, Netherlands, and its Belgian parent company Janssen Pharmaceuticals, subsidiary of American company Johnson & Johnson	Netherlands, Belgian, United States	2	Ad-26 vector expressing the full-length S protein	120
Subunit vaccines	Covovax, TAK-019	NVX-CoV2373	Novavax and the coalition for epidemic preparedness innovations (CEPI)	United States	2	Full-length S protein vaccine adjuvanted with matrix M	121
	ZIFIVAX	ZF2001	Anhui Zhifei Longcom biopharmaceutical/institute of microbiology, Chinese Academy of Sciences	China	3	The adjuvanted recombinant protein (RBD-dimer) expressed in CHO cells	55
	CIGB-66	ABDALA	Center for genetic engineering and biotechnology (CIGB)	Cuba	3	RBD + aluminum hydroxide	55
	-	EpiVacCorona	Federal budgetary research institution state research center of virology and biotechnology (Vector)	Russian	2	Fragments of S protein	55
	FINLAY-FR-2	SOBERANA 02	Instituto Finlay de Vacunas	Cuba	2	RBD chemically conjugated to tetanus toxoid plus adjuvant	55
	-	Sanofi-GSK	Sanofi Pasteur and GlaxoSmithKline company (GSK)	France and England	2		55
VLPs vaccines	-	CoVLP	Medicago Inc	Canada	2	Plant-derived VLP adjuvanted with AS03	122

Live Attenuated Vaccines

Attenuated vaccines take advantage of using a live infectious agent while it has been attenuated. Live attenuated vaccines mimic natural infection and provoke humoral and cell-mediated immune responses, which provide immunity to prevent infections.⁶³ Highly replicating pathogens in cells (for viruses) or culture media (for bacteria) is one way to prepare attenuated vaccines.¹²³ Through this method, although microorganisms lose their virulence, they can replicate sufficiently. Attenuation is not stable, and the virulent form of the virus may return.⁶⁹

Moreover, to prevent the microorganism from partial or total death, attenuated vaccines must be stored at low temperatures.⁶⁸ Multiple antigens with correct conformational structure can be presented by this type of vaccine to the immune system, which helps the immune system to respond better.⁶⁶ Pathogens in attenuated vaccines can still replicate. Therefore, only one dose⁶⁴ or small amounts of the virus is needed for vaccinations, which reduces vaccination costs.⁶⁵

There is a concern that, live attenuated vaccines can transmit between healthcare workers and/or family members of immunocompromised patients. Therefore, both immunocompromised patients and/or persons receiving immunosuppressive therapy are at risk of being infected by the live attenuated vaccine.⁶⁷ An example was the occurrence risk of vaccine-associated paralytic poliomyelitis (VAPP) in the vaccinees or people in contact with the vaccine back in the 1990s in the United States, while the incidence of polio declined.¹²⁴ Moreover, as the pathogens are alive in this type of vaccine and no critical practice has been done during the vaccine preparation, contamination in vaccines with organisms is possible.⁷⁰ Live attenuated influenzas and vaccinia vaccines are among the other examples of this type of vaccine.^{125,126} However, live attenuated Varicella, Measles, Mumps, and Rubella vaccines are still being used as no transmission cases have been reported.¹²⁷ Due to the possibility of causing severe conditions, live attenuated vaccines against chronic diseases such as HIV, or HCV, have not been developed.

Attenuated coronavirus vaccines stimulate immune responses similar to natural infection; however, reversion to wild type and causing severe infections are also possible. The live attenuated coronavirus vaccines have the advantage of presenting all the coronavirus

antigens to the host immune system. As none of these vaccines have entered the phase 3 clinical trial study up to writing this article, information about these types of vaccines is limited. Natural SARS-CoV-2 infection does not provide proper and long-lasting immunity in the infected persons. Therefore, using this type of vaccine requires more research to prevail coronavirus infection. If both the transmissibility and the possibility of reversion to the virulent type would be omitted, one single dose should be enough to provide proper immunity. Attenuated vaccines typically have mounted both humoral and cell-mediated responses. Covi-Vac is one of the potential live attenuated vaccines that has entered into the clinical trial phases till the last revise of this article. This vaccine has developed in collaboration with Codagenix and the serum institute of India. Covi-Vac applies in a single-dose format and the virus was attenuated by codon de-optimization, which stimulates the immune system against all SARS-CoV-2 proteins.¹²⁸ BBIBP-CorV and PiCoVacc are two other examples of potential live attenuated SARS-CoV-2 vaccines that have been passaged many times and deactivated by β -propiolactone.^{129,130}

Inactivated (dead) Vaccines

In dead vaccines, the whole microorganism becomes inactivated using a physical, γ -irradiation, or chemical method. Pathogens in inactivated vaccines have lost their ability for replication which reduces the possibility of transmission and reversion to an infectious form. However, inactivated vaccines induce immune responses to a lesser extent compared to attenuated vaccines.¹³¹ Polio (Salk vaccine), Rabies, influenza, and Japanese B encephalitis are examples of inactivated viral vaccines.¹³²

These vaccines are more stable, conservative, and safer than attenuated vaccines. However, the stimulation of the immune system by the dead virus is lower than the attenuated vaccines,⁷⁴ which means not only a large number of dead pathogens is required, but also several doses are needed to trigger the immune responses. These amounts can increase the risk of an allergic response and vaccination costs.⁷⁵ The main limitation of dead vaccines is that they stimulate the immune system less, as the dead pathogens cannot actively penetrate host cells and present their antigens on MHC class I.¹³³ Dead vaccines are stable; therefore, the cold chain transition can be omitted.⁷³ They are also

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safe as the vaccine preparation process dramatically reduces the risk of contamination with other live organisms.⁷¹

Sinovac, Wuhan Institute of Biological Products, Beijing Institute of Biological Products, Bharat Biotech, Research Institute for Biological Safety Problems, Rep of Kazakhstan, Beijing Minhai biotechnology Co, Shenzhen Kangtai Biological Products Co, Valneva, and Chinese Academy of Medical Sciences in collaboration with the Chinese Academy of Medical Sciences, have introduced inactivated vaccine candidates against SARS-CoV-2. Results in mice, rats, non-human primates, and 3 phases of clinical trials have shown the excellent immunogenicity of vaccines, while the number of adverse reactions was lower compared with viral-vectored vaccines, DNA, or RNA vaccines.¹³⁴ The Vero cell lines are continuous cell lines (CCLs) used to produce inactivated vaccines for many years.¹³⁵ This platform is in use by several developers to inactivate SARS-CoV-2 viruses and produce Vaccines too.

Nucleic Acid-based Vaccines

Instead of using the whole pathogens or part of them, the nucleic acid sequence of the immunogenic protein can be used in nucleic acid-based vaccines. Following the vaccination, cells uptake the nucleic acid and synthesize immunogens. Synthesized immunogens are then processed through the endogenous pathway, presented on cells, and activate cell-mediated responses.¹³⁶ Both DNA and RNA can be used in nucleic acid-based vaccines. Choosing the desired protein for encoding through nucleic acid vaccines is of high importance. A DNA vaccine is a plasmid that encodes the interested protein.¹³⁷ DNA is more stable and can express immunogens for a more extended period, but there is also a risk of integration into the genome.^{138, 139}

Moreover, the produced antibody response to DNA vaccines has a lower titer, avidity, longevity, and CTL response than natural infections.^{82,83} Special elements, which are needed for DNA vaccines, increase the cost and size of DNA vaccines.⁸⁴ However, only a microgram of this construct can trigger an immune response.¹⁴⁰ Besides, unlike the mRNA vaccines, toxicity and autoimmune responses have not been detected in DNA vaccines.^{90,141,142} Bacterial sequences in plasmids can be manipulated to act as an adjuvant.⁷⁷

Also, the sequences of antigens can be easily modified for Special goals.⁸⁰

RNA vaccines consist of messenger RNA (mRNA) molecules with pathogen-specific antigen sequences producing antigenic peptides or proteins *in situ* after entry into vaccinees cells. Following antigen presentation, the immune system will be triggered.¹⁴³ RNA vaccines only need to enter the cells and do not need a carrier to lead them to the nucleus.⁸⁶ Therefore using RNA vaccines needs fewer co-particles.⁷⁸ Also, the production of mRNA vaccines is more comfortable than other vaccines and more cost-effective.⁸⁵

Moreover, some synthetic RNA sequences like CpG can stimulate an immune response.¹⁴⁴ New lipid nanoparticles (LNP)-encapsulated mRNA vaccines named ARCoV, increase the stability of mRNA, which could be stored at room temperature. This stability reduces the difficulty of transportation and decreases the cost of storage.⁸⁹ RNA vaccines are safer than DNA vaccines since RNA vaccines are generated in cell-free conditions.¹⁴⁵ Despite the reduced risk of integration, according to the high sensitivity of RNA, synthesis and delivery of this type of vaccine is very challenging.⁷⁸ However, in the case of the previous infection of vaccinees by a retrovirus, the required proteins for integrating mRNA into the genome are exist in cells.¹⁴⁶

Nucleic acid vaccines are free of any living organisms, completely excluding the risk of using live organisms. The nucleic acid sequences can be modified and optimized to stimulate immune responses better.¹⁴⁷ Nucleic acid-based vaccines efficiently stimulate both humoral and cell-mediated responses together with innate immune responses.⁷⁹ It is of note that nucleic acid vaccines can only be used for mimicking the “one or a few” “peptide” antigens of the whole pathogen.⁷⁸ Human cells possess different post-translational modifications compared to most of the pathogens, affecting the immune response of encoded protein.⁸¹

Moderna, BioNTech, ARCoV, and CureVac have developed mRNA vaccines against SARS-CoV-2. Moderna and BioNTech groups have used lipid nanoparticles (LNPs) for the non-viral delivery of mRNA and protect mRNA from destruction. The formulation of LNPs is vital in terms of preventing RNA degradation. Also, triggering the immune responses with these particles has been shown in several studies.¹⁴⁸ Both of these vaccines have finished phase 3 clinical trials with high efficacy (approximately

95%) against SARS-CoV-2 infection.^{87,88} CureVac's CvnCoV is an mRNA vaccine candidate that utilizes nucleotides without chemical modifications in the mRNA. The mRNA encodes the full-length spike protein of SARS-CoV-2 and is formulated with lipid nanoparticles. ARCoV is an mRNA-LNP vaccine that encoded RBD and can elicit robust neutralizing antibodies and cellular responses against SARS-CoV-2. One of the more critical advantages of ARCoV is storage at room temperature for at least one week.¹¹⁶ The only type of DNA vaccine in phase 3 against SARS-CoV-2 was developed by Zydus Cadila and named that nCov vaccine.

Viral Vector-based Vaccines

Vector-based vaccines are composed of a carrier virus unrelated to the pathogen of interest, such as *Adeno* or *Vaccinia* viruses. These viruses become attenuated and modified to carry a gene of the pathogen.^{149,150} The encoded gene by this viral vector will express in recipient cells, and consequently, the immune system will respond to the expressed protein which presents on MHC I. Although viral vector vaccines trigger both humoral and cell-mediated immune cell responses efficiently, the CTL activation may cause side effects.^{92,93} Furthermore, these vaccines can be directed to specific issues for stimulating the immune response more sufficiently. Therefore, only a lower dose of vaccines can trigger an appropriate immune response.

Viral vector-based vaccines do not have any of the pathogens' genes except the gene which encodes the antigen. However, the possibility of viral vector reversion is not eliminated. The immune system response to the viral vector is another limitation of viral vector-based vaccines, which can neutralize the vector completely.⁹⁴ Serotypes with lesser exposure in humans can be used as a viral vector to prevent the neutralization of viral-based vectors. In this regard, chimpanzee adenovirus serotype Y25 (ChAdY25), is one of the most suitable serotypes of adenoviruses since it triggers no or a weak immune response in the human body. Besides, other chimpanzee adenovirus serotypes such as ChAd63 and ChAd3 and also human adenovirus serotype-5 (HadV-5), HadV-6, HadV-35 can be good candidates for as application in viral vectors.¹⁵¹ Many biotech companies have explored several vector-based vaccine structures to develop new vaccines.^{152,153} Before the COVID-19 outbreak and up

to 2019, two viral vector-based vaccines (rVSV-ZEBOV and Dengvaxia) got the license for human use.

Among human adenovirus serotypes, Ad serotype 5 (Had5) has been mainly used as a viral vector in human vaccines. This vector has several advantages, such as high transduction efficiency, a high level of transgene expression, and a broad range of viral tropism. However, due to the high number of CARs on the hepatocyte surface, using Had5-based vectors may lead to the entrance of many viruses into the liver, which causes hepatotoxicity.⁹⁵

Beijing Institute of biotechnology, University of Oxford, Gamaleya research institute, and Janssen pharmaceutical companies have introduced viral-based vaccines against SARS-CoV-2 which used ChAdOx1-S, Ad5, rAd26-S+rAd5-S, and Ad26 vector, respectively. Russian vaccine induced strong humoral and cellular immune responses against SARS-CoV-2,⁵⁴ which provide 92% protection.⁹⁷ These results for the Russian vaccine can be due to the use of two different viral vectors in each dose, which is new to the immune system at each injection, and there is no previous response to it in the recipient's body. The Oxford vaccine was reported to provide both humoral and cellular immune responses against SARS-CoV-2 with 70% protection⁹⁶ and no serious adverse events related to ChAdOx1.¹⁵⁵

Subunit Vaccines

Any components of pathogens with the ability to trigger immune system responses can be used as a vaccine. These components can be generated through purification or by applying recombinant DNA technology.¹⁵⁶ These vaccines can consist of whole macromolecules, large fragments of macromolecules, polysaccharide chains, or polypeptide fragments of pathogens.¹⁵⁷

Hepatitis B surface antigen (HbsAg), the first viral subunit vaccine, is an example of a vaccine with the whole macromolecules (of Hepatitis B).¹⁵⁸ In subunit vaccines, the possibility of contamination and reversion is excluded,⁹⁸ and despite the low immunogenicity,¹⁰² modified subunit vaccines can trigger humoral and cellular immune responses.⁹⁹ Due to the stability of these vaccines, no special care is needed during transport.¹⁰⁰ Since even tiny changes in subunit vaccines may stimulate an inappropriate immune response, conformation and post-translational modification of components is extremely important.¹⁵⁹

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The half-life of subunit vaccines in the body is too short to be detected by the immune system; therefore, adjuvants and delivery vehicles seem necessary to protect them from degradation, or more doses are needed, which increases costs.¹⁰³⁻¹⁰⁵ Moreover, it is critical to purify this type of vaccine.¹⁰¹

Currently, Novavax, center for genetic engineering and biotechnology (CIGB), Federal budgetary research institution state research center of virology and biotechnology, institute Finlay de vacunas, Sanofi Pasteur in collaborating with GlaxoSmithKline company (GSK), and Anhui Zhifei Longcom biopharmaceutical have developed the SARS-CoV-2 subunit vaccine, which reached to phase3 clinical trial successfully. Novavax has a full-length recombinant Spike glycoprotein nanoparticle vaccine adjuvanted with matrix M.¹⁶⁰ Matrix-M™ adjuvant in Novavax includes two 40-nm particles with different saponin fractions. Matrix-A and -C particles are produced from the leaves of the tree *Quillaja Brasiliensis* with cholesterol and phospholipid.¹⁶¹ The role of this adjuvant in stimulating antigen-specific humoral and cellular immune responses was proven.¹⁶²

In contrast, Anhui Zhifei Longcom Biopharmaceutical used the RBD, which is produced in the Chinese hamster ovary (CHO) cells. CIGB-66 and EpiVac vaccines have an RBD domain and fragments of a spike protein in an aluminum-containing adjuvant (aluminum hydroxide), respectively. FINLAY-FR-2 vaccine has an RBD domain produced in Chinese hamster ovary (CHO) cells conjugated chemically to tetanus toxoid. It seems that, in case of sufficient induction of humoral and cellular immune response against SARS-CoV-2 this type of vaccine, will provide a safer method without special adverse events and concerns in humans.

Virus-like Particles (VLPs)

Virus-like particles (VLPs) include self-assembled viral protein(s) in virus-shaped particles. These proteins mainly belong to the envelope; however, they sometimes contain viral core proteins like hepatitis B virus core proteins.¹⁶³

Due to the absence of viral proteins encoding nucleic acids, VLPs cannot replicate independently. Therefore, the risk of replication and infection in the vaccinees body would be omitted.¹⁰⁶ VLPs are more immunogenic than subunit Vaccines and can induce both humoral and cellular arms of the immune

system.¹⁰⁸ Epitope display on VLPs are similar to the native virus that can induce a potent antibody response as the epitope's conformation is correct. Moreover, VLPs size is suitable for uptake by dendritic cells (DCs).¹⁶⁴ VLPs can act as an adjuvant for their carrier epitopes, so unlike the subunit vaccines, there is no need for adjuvants to stimulate the immune system.¹⁰⁹ Besides, the preparation of VLPs is easier than viral vector-based vaccines, and unlike viral vector-based vaccine preparation, there is no need for human cells to produce VLPs vaccines. Preparation of VLPs vaccine is too laborious¹¹⁰ which increases the vaccine price.¹⁶⁵

There is just one VLP vaccine against SARS-CoV-2 currently in phase 2/3 clinical trial. In this regard, Medicago has developed self-assembling VLPs having SARS-CoV-2's spike protein on its surface and AS03 as the adjuvant. The spike protein is transiently expressed in *Nicotiana benthamiana*, a close relative of tobacco. Medicago's VLP has a similar shape and size to SARS-CoV-2. This vaccine can elicit both humoral and cellular responses and establish protective immunity in vaccinees. Medicago's platform has been previously applied to produce Vaccines for avian and seasonal influenza.¹²² AS03 adjuvant is an oil-in-water emulsion, consisting of α -tocopherol, squalene, and polysorbate 80. AS03 has already been investigated in several nonclinical and clinical studies.¹⁶⁶

Adverse Reactions of Some Vaccines

The most common side effects of vaccines are pain at the injection site, fever, myalgia, fatigue, and headache. One of the most significant adverse reactions of vaccination is anaphylactic reactions which can lead to death. Activation of mast cells followed by releasing mediators from them can cause anaphylactic reactions.¹⁶⁷ The possibility of anaphylactic reactions is higher in a person with a history of allergies to any food, drug, or vaccine. For most known vaccines, anaphylactic reactions occur one case per million injections.¹⁶⁸ Based on the published clinical trial data of SAR-CoV-2 vaccines, there were mild to moderate adverse reactions with few severe reactions in vaccinees.¹⁶⁹ Due to no previous licensed viral vector-based, DNA, and mRNA vaccines; some populations may be at higher risk for adverse reactions related to a component of these vaccines.

Several cases of anaphylaxis (some of them had allergies history, but some others had no known allergies) associated with the Pfizer mRNA vaccine

have been reported in the United States and UK.¹⁶⁷ After that, persons who have an allergy history have been excluded from vaccination programs. Several reports indicate Moderna vaccine adverse reactions are more frequently reported compared with the Pfizer vaccine. The clinical trial data of the Sinopharm, which is a whole virus vaccine, indicated that the occurrence of four severe adverse events was not related to the vaccination.¹¹² In ChAdOx1 nCoV-19 (AZD1222) vaccine, hemolytic anemia, transverse myelitis, fever higher than 40°C were reported as adverse events. There was one non-COVID-19 death in the experimental vaccine group, but these were considered unrelated to the vaccine.¹⁷⁰ Sputnik vaccine adverse events were mild, and most of the adverse reactions occurred after the second vaccination.¹⁷¹ Clinical trials data of the Janssen vaccine demonstrated five serious adverse events (SAE), including hypotension, bilateral nephrolithiasis, legionella pneumonia, worsening of multiple sclerosis, and fever leading to hospitalization. Except for the last one, all these SAE were deemed unrelated to the vaccine.¹⁷² There are no severe adverse reactions for the Novavax vaccine¹⁷³ and Covaxin vaccine¹⁷⁴ based on their clinical trials.

The New Variation of SARS-CoV-2

Due to the higher rate of mutation in the RNA virus, new variants of SARS-CoV-2 are expected to occur over time worldwide. By recognition of a new variant of SARS-CoV-2, its ability to spread, illness severity, symptoms, and also vaccine efficacy against it should be examined. SARS-CoV-2 interagency group (SIG) classified variants into three groups and named variants of interest (VOI), variants of concern (VOC), variants of high consequence (VOHC).¹⁷⁵ Based on SIG information, five variants of SARS-CoV-2 circulating in the United States (The B.1.1.7, B.1.351, P.1, B.1.427, and B.1.429 are among VOI variants) are classified as VOI.

B.1.1.7 (the United Kingdom or 501Y.V1) is the name of the variant which initially detected in the UK. It has 23 mutations with 17 amino acid changes.¹⁷⁶ Transmission¹⁷⁷ and case fatality¹⁷⁸ of this variant are more than the first identified SARS-CoV-2. Mutations in B.1.1.7 have no impact on treatment¹⁷⁹ and have little impact on vaccine efficacy,¹⁸⁰ which can be ignored. The variant was initially detected in South Africa in December 2020 named B.1.351 (501Y.V2 or South Africa), with 23 mutations with 17 amino acid

changes.¹⁷⁶ This variant treatment strategy was changed due to decrease in susceptibility to antibodies.¹⁷⁹ P.1 (Brazil, B.1.1.28.1, or 501Y.V3) with approximately 35 mutations and 17 amino acid changes was initially identified in travelers from Brazil, who were tested during routine screening at an airport in Japan in early January. An alternative treatment strategy was needed for this variant¹⁷⁹ and also vaccines efficacy was reduced against this variant.¹⁸⁰

SIG also classified P.2, B.1.525, B.1.526.1, and B.1.526 in the VOI group. B.1.526.1 and B.1.526 were initially detected in the United States. B.1.525 and P.2 were initially detected in United Kingdom/Nigeria and Brazil, respectively. All these variants also have different treatment strategies and vaccine efficacy, but the transmission rate did not change significantly.¹⁸² Several variants are not classified by SIG. B.1.617 (India) with several spike mutations is one of those variants. It was initially reported in India in early 2021 with a higher transmission rate than the first identified SARS-CoV-2.¹⁸³ B.1.427 and B.1.429 were first identified in California in February 2021 and were classified as VOCs in March 2021. These variants also have different transmission rate, treatment strategies, and vaccine efficacy.¹⁸¹ Still, there is no variant in the VOHC group. Several mutations of significant SARS-CoV-2 variants were shown in Table 4.

There are a few published studies that analyzed the efficacy of the vaccine in patients with different SARS-CoV-2 variants. One of those studies confirmed AstraZeneca ChAdOx1 vaccine has 75% protection against B.1.1.7.¹⁹⁵ However, protection of this vaccine against the B.1.351 variant was just 10%.¹⁹⁶ Another study showed that the AstraZeneca ChAdOx1 vaccine has 74% and 22% efficacy against B.1.1.7¹⁹⁶ and the B.1.351¹⁹⁷ variants, respectively. In comparison, Johnson & Johnson's vaccine showed 64% protection against the B.1.351 variant.¹²⁰ The Novavax vaccine demonstrated 50% and 86% protection against B.1.351 and B.1.1.7 variants, respectively¹⁹⁵ which almost was confirmed by Wadman et al.¹⁹⁸ The CoronaVac/Sinovac vaccine efficacy against the P.1 variant was estimated to be about 50%.¹⁹⁹ Moderna vaccine has lower efficacy against B.1.1.7, B.1.351, and P.1 too.¹⁷⁶ Although some vaccines still have enough protection against some of the new variants, formulating new vaccines may be necessary to control some new SARS-CoV-2 variants.

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Table 4. Several mutations of each variant of SARS-CoV-2 with its position and effect.

Variant	Mutations	Position	Effect	Reference
B.1.1.7	L18F	N-terminal domain (NTD)	Reduced sensitivity to neutralizing antibodies	184
	69–70del 144del		Impact on immune escape	185
	E484K	Receptor-binding site (RBS)	Escape from neutralizing antibodies	186
	N501Y	RBD	Improve the interaction between spike protein and angiotensin-converting enzyme 2 (ACE2)	187
	D614G	near the RBD	Increased transmissibility for viruses	
	P681H T716I	near the furin cleavage site		
	S982A	heptad repeat 1 (HR1)		188
	D1118H	near the heptad repeat 2 (HR 2)		
B.1.351	L18F D80A D215G 242–244del	NTD	Impair the efficacy of the current vaccine	189
	R246I K417N			
	E484K N501Y	RBD	Enhancement of the binding of RBD to ACE2 Escape from neutralizing antibodies	190
	A701V	near the furin cleavage site	Improve the interaction between spike protein and ACE2	191
P.1	L18F T20N P26S D138Y R190S	NTD	Reduced sensitivity to neutralizing antibodies	
	K417N			
	E484K	RBD	Enhancement of the binding of RBD to ACE2 Escape from neutralizing antibodies	192
	K417T	RBD	Improve the interaction between spike protein and ACE2	
	N501Y			
	H655Y	near the RBD		
	T1027I	near the HR1		
B.1.617 (183)	G142D E154K	NTD	Improve the interaction between spike protein and ACE2	193
	L452R E484Q			
	D614G	near the RBD	Increased transmissibility for viruses	
	P681R	near the RBD		194
	Q1071H	near the HR2		

DISCUSSION

Based on vaccine efficiency, almost 67% of the population must be vaccinated to reach herd immunity for COVID-19.²⁰⁰ This percent may be variable for different types of vaccines with various efficiencies. Although the vaccination does not mean the end of COVID-19, it can stop the disease's high prevalence. Based on what has been discussed, the safest vaccines with the most negligible long-term side effects are subunit vaccines. These vaccines, along with VLPs have a history of usage in humans, and due to the lack of certain harmful substances in these vaccines, they may not cause long-term side effects. A large number of subunit vaccines are being tested for SARS-CoV-2.⁵⁵ In addition to the lower risk, there is considerable focus on subunit vaccines due to their efficiency against other pathogens. However, it is of note that none of the subunit vaccines have provided a protective immune response against other RNA viruses, such as HIV and influenza. Viral vector-based vaccines and nucleic acid vaccines are placed in the following ranks of vaccine testing for SARS-CoV-2.⁵⁵ Due to the shorter lifespan of mRNA vaccines,⁷⁸ they seem to be more acceptable among the new generation of vaccines. However, for all vaccine types, longer-term follow-ups are needed. Vaccines against SARS-CoV-2 have shown up to 95% efficacy;⁸⁷ however, in order to have protection against new variants of SARS-CoV-2, it seems that several changes and improvements in vaccines may be needed.

Due to the requirement of vaccinating a large population and the unspecificity of the immunization period in vaccinated people, and the possible need for booster doses, the vaccine price is of high importance. Viral vector-based vaccines have been the most cost-effective type of vaccine, and each dose of this vaccine seems to be purchased for almost 3-10 dollars compared to \$19.50 for Pfizer and \$32-37 for Moderna. Another critical issue to note, is that whether the vaccine completely prevents infection or just reduces the symptoms. Since if reducing the symptoms is the case, the virus carriers will increase, contributing to the further spread of it. Another concern is how to distribute the vaccine properly. Delay to reach herd immunization in many countries, showed the importance of a precise and detailed planning to identify susceptible individuals who need to be vaccinated immediately. In case of new pandemic in

the future, it would be a good idea to consider plans of successful countries all around the world.

In conclusion, SARS-CoV-2 vaccines have been developed faster than any other vaccines in history (the previous fastest production for vaccines belongs to the mumps vaccine, with a period of approximately four years). The reason for that may be the increasing number of companies and institutes working on the vaccines and more funds are assigned to this research area. Developing SARS-CoV-2 vaccines may affect the speed of the preparation process for other vaccines such as HIV and influenza.

CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

ACKNOWLEDGEMENTS

This study is related to project NO. 1399/63358 From Student Research Committee, Shahid Beheshti University of Medical Sciences, Tehran, Iran. We also appreciate the "Student Research Committee" and "Research & Technology Chancellor" at Shahid Beheshti University of Medical Sciences for their financial support of this study.

REFERENCES

1. Zhong N, Zheng B, Li Y, Poon L, Xie Z, Chan K, et al. Epidemiology and cause of severe acute respiratory syndrome (SARS) in Guangdong, People's Republic of China, in February, 2003. *Lancet*. 2003;362(9393):1353-8.
2. Nourizadeh M, Rasaei MJ, Moin M. COVID-19 Pandemic: A Big Challenge in Iran and the World. *Iran J Allergy Asthma Immunol*. 2020;19(S1):1-2.
3. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med*. 2020;382(7):727-33.
4. Mostafaei A, Ghojzadeh M, Hajebrahimi S, Abolhasanpour N, Salehi-Pourmehr H. Clinical Presentation of Iranian Patients Affected with COVID-19: A Thousand Faces Disease. *Iran J Allergy Asthma Immunol*. 2021;20(2):140-6.
5. Ghazanfari T, Salehi MR, Namaki S, Arabkheradmand J, Rostamian A, Rajabnia Chenary M, et al. Interpretation of Hematological, Biochemical, and Immunological Findings of COVID-19 Disease: Biomarkers Associated

Pro and Cons of Coronavirus Vaccines

- with Severity and Mortality. *Iran J Allergy Asthma Immunol.* 2021;20(1):46-66.
6. Seow J, Graham C, Merrick B, Acors S, Steel KJ, Hemmings O, et al. Longitudinal evaluation and decline of antibody responses in SARS-CoV-2 infection. *MedRxiv.* 2020;7(9):20148429.
 7. Tregoning JS, Brown ES, Cheeseman HM, Flight KE, Higham SL, Lemm NM, et al. Vaccines for COVID-19. *Clin Exp Immunol.* 2020;202(2):162-92.
 8. Tian X, Li C, Huang A, Xia S, Lu S, Shi Z, et al. Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody. *Emerg Microbes Infect.* 2020;9(1):382-5.
 9. Al-Kassmy J, Pedersen J, Kobinger G. Vaccine Candidates against Coronavirus Infections. Where Does COVID-19 Stand? *Viruses.* 2020;12(8):861-9.
 10. Chan JF-W, Kok K-H, Zhu Z, Chu H, To KK-W, Yuan S, et al. Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan. *Emerg Microbes Infect.* 2020;9(1):221-36.
 11. Kim J-M, Chung Y-S, Jo HJ, Lee N-J, Kim MS, Woo SH, et al. Identification of Coronavirus Isolated from a Patient in Korea with COVID-19. *Osong Public Health Res Perspect.* 2020;11(1):3-7.
 12. Sariol A, Perlman S. Lessons for COVID-19 Immunity from Other Coronavirus Infections. *Immunity.* 2020;53(2):248-63.
 13. Islam MR, Hoque MN, Rahman MS, Alam ASMRU, Akther M, Puspo JA, et al. Genome-wide analysis of SARS-CoV-2 virus strains circulating worldwide implicates heterogeneity. *Sci Rep.* 2020;10(1):14004.
 14. Musavi H, Abazari O, Safaee MS, Variji A, Koohshekan B, Kalaki-Jouybari F, et al. Mechanisms of COVID-19 Entry into the Cell: Potential Therapeutic Approaches Based on Virus Entry Inhibition in COVID-19 Patients with Underlying Diseases. *Iran J Allergy Asthma Immunol.* 2021;20(1):11-23.
 15. Fehr AR, Perlman S. Coronaviruses: an overview of their replication and pathogenesis. *Methods Mol Biol.* 2015;1282:1-23.
 16. Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science.* 2020;367(6483):1260-3.
 17. Bosch BJ, van der Zee R, de Haan CA, Rottier PJ. The coronavirus spike protein is a class I virus fusion protein: structural and functional characterization of the fusion core complex. *J Virol.* 2003;77(16):8801-11.
 18. Xia S, Zhu Y, Liu M, Lan Q, Xu W, Wu Y, et al. Fusion mechanism of 2019-nCoV and fusion inhibitors targeting HR1 domain in spike protein. *Cell Mol Immunol.* 2020;17(7):765-7.
 19. Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature.* 2020;579(7798):270-3.
 20. Liu S, Xiao G, Chen Y, He Y, Niu J, Escalante CR, et al. Interaction between heptad repeat 1 and 2 regions in spike protein of SARS-associated coronavirus: implications for virus fusogenic mechanism and identification of fusion inhibitors. *Lancet.* 2004;363(9413):938-47.
 21. Li G, Fan Y, Lai Y, Han T, Li Z, Zhou P, et al. Coronavirus infections and immune responses. *J Med Virol.* 2020;92(4):424-32.
 22. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell.* 2020;181(2):271-80.
 23. Wang C, Li W, Drabek D, Okba NMA, van Haperen R, Osterhaus A, et al. A human monoclonal antibody blocking SARS-CoV-2 infection. *Nat Commun.* 2020;11(1):2251-5.
 24. Seydoux E, Homad LJ, MacCamy AJ, Parks KR, Hurlburt NK, Jennewein MF, et al. Analysis of a SARS-CoV-2-Infected Individual Reveals Development of Potent Neutralizing Antibodies with Limited Somatic Mutation. *Immunity.* 2020;53(1):98-105.
 25. Chi X, Yan R, Zhang J, Zhang G, Zhang Y, Hao M, et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. *Science.* 2020;369(6504):650-5.
 26. Chi X, Yan R, Zhang J, Zhang G, Zhang Y, Hao M, et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. *Science.* 2020;369(6504):650-5.
 27. Grant OC, Montgomery D, Ito K, Woods RJ. Analysis of the SARS-CoV-2 spike protein glycan shield reveals implications for immune recognition. *Sci Rep.* 2020;10(1):1-11.
 28. Su S, Du L, Jiang S. Learning from the past: development of safe and effective COVID-19 vaccines. *Nat Rev Microbiol.* 2021;19(3):211-9.
 29. Yang J, Wang W, Chen Z, Lu S, Yang F, Bi Z, et al. A vaccine targeting the RBD of the S protein of SARS-

- CoV-2 induces protective immunity. *Nature*. 2020;586(7830):572-7.
30. Jiaming L, Yanfeng Y, Yao D, Yawei H, Linlin B, Baoying H, et al. The recombinant N-terminal domain of spike proteins is a potential vaccine against Middle East respiratory syndrome coronavirus (MERS-CoV) infection. *Vaccine*. 2017;35(1):10-8.
 31. Ng KW, Faulkner N, Cornish GH, Rosa A, Harvey R, Hussain S, et al. Preexisting and de novo humoral immunity to SARS-CoV-2 in humans. *Science*. 2020;370(6522):1339-43.
 32. Watanabe Y, Allen JD, Wrapp D, McLellan JS, Crispin M. Site-specific glycan analysis of the SARS-CoV-2 spike. *Science*. 2020;369(6501):330-3.
 33. Grifoni A, Weiskopf D, Ramirez SI, Mateus J, Dan JM, Moderbacher CR, et al. Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals. *Cell*. 2020;181(7):1489-501.e15.
 34. Sun J, Zhuang Z, Zheng J, Li K, Wong RL-Y, Liu D, et al. Generation of a broadly useful model for COVID-19 pathogenesis, vaccination, and treatment. *Cell*. 2020;182(3):734-43. e5.
 35. Long Q-X, Liu B-Z, Deng H-J, Wu G-C, Deng K, Chen Y-K, et al. Antibody responses to SARS-CoV-2 in patients with COVID-19. *Nat Med*. 2020;26(6):845-8.
 36. Yasui F, Kai C, Kitabatake M, Inoue S, Yoneda M, Yokochi S, et al. Prior immunization with severe acute respiratory syndrome (SARS)-associated coronavirus (SARS-CoV) nucleocapsid protein causes severe pneumonia in mice infected with SARS-CoV. *J Immunol*. 2008;181(9):6337-48.
 37. Dai L, Gao GF. Viral targets for vaccines against COVID-19. *Nat Rev Immunol*. 2021;21(2):73-82.
 38. Bos R, Rutten L, van der Lubbe JEM, Bakkers MJG, Hardenberg G, Wegmann F, et al. Ad26 vector-based COVID-19 vaccine encoding a prefusion-stabilized SARS-CoV-2 Spike immunogen induces potent humoral and cellular immune responses. *BioRxiv*. 2020;7(30):227470.
 39. Bangaru S, Ozorowski G, Turner HL, Antanasijevic A, Huang D, Wang X, et al. Structural analysis of full-length SARS-CoV-2 spike protein from an advanced vaccine candidate. *Science*. 2020;370(6520):1089-94.
 40. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395(10223):497-506.
 41. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med*. 2020;8(4):420-2.
 42. Tay MZ, Poh CM, Rénia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. *Nat Rev Immunol*. 2020;20(6):363-74.
 43. Channappanavar R, Fett C, Zhao J, Meyerholz DK, Perlman S. Virus-specific memory CD8 T cells provide substantial protection from lethal severe acute respiratory syndrome coronavirus infection. *J Virol*. 2014;88(19):11034-44.
 44. Zhang F, Gan R, Zhen Z, Hu X, Li X, Zhou F, et al. Adaptive immune responses to SARS-CoV-2 infection in severe versus mild individuals. *Signal Transduct Target Ther*. 2020;5(1):156.
 45. Ni L, Ye F, Cheng M-L, Feng Y, Deng Y-Q, Zhao H, et al. Detection of SARS-CoV-2-specific humoral and cellular immunity in COVID-19 convalescent individuals. *Immunity*. 2020;5286(14):971-7.
 46. Poland GA, Ovsyannikova IG, Kennedy RB. SARS-CoV-2 immunity: review and applications to phase 3 vaccine candidates. *Lancet*. 2020;396(10262):1095-606.
 47. Moderbacher CR, Ramirez SI, Dan JM, Grifoni A, Hastie KM, Weiskopf D, et al. Antigen-specific adaptive immunity to SARS-CoV-2 in acute COVID-19 and associations with age and disease severity. *Cell*. 2020;183(4):996-1012.
 48. Grifoni A, Weiskopf D, Ramirez SI, Mateus J, Dan JM, Moderbacher CR, et al. Targets of T cell responses to SARS-CoV-2 coronavirus in humans with COVID-19 disease and unexposed individuals. *Cell*. 2020;181(7):1489-501.
 49. Le Bert N, Tan AT, Kunasegaran K, Tham CY, Hafezi M, Chia A, et al. SARS-CoV-2-specific T cell immunity in cases of COVID-19 and SARS, and uninfected controls. *Nature*. 2020;584(7821):457-62.
 50. Fontanet A, Cauchemez S. COVID-19 herd immunity: where are we? *Nat Rev Immunol*. 2020;20(10):583-4.
 51. Deng W, Bao L, Liu J, Xiao C, Liu J, Xue J, et al. Primary exposure to SARS-CoV-2 protects against reinfection in rhesus macaques. *Science*. 2020;369(6505):818-23.
 52. Long SW, Olsen RJ, Christensen PA, Bernard DW, Davis JJ, Shukla M, et al. Molecular architecture of early dissemination and massive second wave of the SARS-CoV-2 virus in a major metropolitan area. *mBio*. 2020;11(6):e02707-20.

Pro and Cons of Coronavirus Vaccines

53. Mak TW, Saunders ME. The immune response. Basic and clinical principles: Elsevier; 1st. Academic Press. 2006;469-74.
54. Food, Administration D. Coronavirus (COVID-19) Update: FDA Takes Action to Help Facilitate Timely Development of Safe, Effective COVID-19 Vaccines. WHO. 2020. Episode#26.
55. <https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines>.
56. Oliveira SC, Rosinha GMS, de-Brito CFA, Fonseca CT, Afonso RR, Costa MCMS, et al. Immunological properties of gene vaccines delivered by different routes. *Brazilian Journal of Medical and Biological Research*. 1999;32(2): 207-14.
57. Bandehpour M, Khodabandeh M, Kazemi B. Cloning and Expression of Hepatitis B Surface Antigen. *Hepat Mon*. 8(1):17-21.
58. Mehdinejadani K, Bandehpour M, Hashemi A, Ranjbar MM, Taheri S, Jalali SA, et al. In Silico Design and Evaluation of *Acinetobacter baumannii* Outer Membrane Protein a Antigenic Peptides As Vaccine Candidate in Immunized Mice. *Iran J Allergy Asthma Immunol*. 2019;18(6):655-63.
59. Traggiai E, Becker S, Subbarao K, Kolesnikova L, Uematsu Y, Gismondo MR, et al. An efficient method to make human monoclonal antibodies from memory B cells: potent neutralization of SARS coronavirus. *Nat Med*. 2004;10(8):871-5.
60. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270-3.
61. Walls AC, Park Y-J, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell*. 2020;183(6):1735.
62. Sautto GA, Kirchenbaum GA, Diotti RA, Criscuolo E, Ferrara F. Next Generation Vaccines for Infectious Diseases. *J Immunol Res*. 2019;2019:5890962.
63. Côté-Gravel J, Brouillette E, Malouin F. Vaccination with a live-attenuated small-colony variant improves the humoral and cell-mediated responses against *Staphylococcus aureus*. *PLoS One*. 2019;14(12):e0227109.
64. Xie X, Kum DB, Xia H, Luo H, Shan C, Zou J, et al. A single-dose live-attenuated Zika virus vaccine with controlled infection rounds that protects against vertical transmission. *Cell Host Microbe*. 2018;24(4):487-99. e5.
65. de Boer PT, van Lier A, de Melker H, van Wijck AJ, Wilschut JC, van Hoek AJ, et al. Cost-effectiveness of vaccination of immunocompetent older adults against herpes zoster in the Netherlands: a comparison between the adjuvanted subunit and live-attenuated vaccines. *BMC Med*. 2018;16(1):228-31.
66. Roper RL, Rehm KE. SARS vaccines: where are we? *Expert Rev Vaccines*. 2009;8(7):887-98.
67. Kamboj M, Sepkowitz KA. Risk of Transmission Associated With Live Attenuated Vaccines Given to Healthy Persons Caring for or Residing With an Immunocompromised Patient. *Infect Control Hosp Epidemiol*. 2007;28(6):702-7.
68. Harper SA, Fukuda K, Cox NJ, Bridges CB. Using live, attenuated influenza vaccine for prevention and control of influenza. *MMWR Recomm Rep*. 2003;52(3):1-8.
69. Selvapandiyan A, Dey R, Gannavaram S, Solanki S, Salotra P, Nakhasi HL. Generation of growth arrested *Leishmania amastigotes*: a tool to develop live attenuated vaccine candidates against visceral leishmaniasis. *Vaccine*. 2014;32(31):3895-901.
70. Boumart Z, Daouam S, Belkourati I, Rafi L, Tuppurainen E, Tadlaoui KO, et al. Comparative innocuity and efficacy of live and inactivated sheepox vaccines. *BMC Vet Res*. 2016;12(1):1-6.
71. Vetter V, Denizer G, Friedland LR, Krishnan J, Shapiro M. Understanding modern-day vaccines: what you need to know. *Ann Med*. 2018;50(2):110-20.
72. Toman M, Celer V, Kavanová L, Levá L, Frolichova J, Ondráčková P, et al. Dynamics and Differences in Systemic and Local Immune Responses After Vaccination With Inactivated and Live Commercial Vaccines and Subsequent Subclinical Infection With PRRS Virus. *Front Immunol*. 2019;10(9):1689-91.
73. Van TTH, Lin Y-C, Color PJ, Smooker PM. From animals to humans: can vaccines make the transition. Nova Science Publishers: Hauppauge, NY, USA; 2012; 1-63.
74. Sanders B, Koldijk M, Schuitemaker H. Inactivated Viral Vaccines. In: Nunnally B., Turula V., Sitrin R. (eds) *Vaccine Analysis: Strategies, Principles, and Control*. Springer, Berlin, Heidelberg. 2015; 45-80.
75. Strugnell R, Zepp F, Cunningham A, Tantawichien T. Vaccine antigens. *Perspect Vaccinol*. 2011;1(1):61-88.
76. Kumar A, McElhaney JE, Walrond L, Cyr TD, Merani S, Kollmann TR, et al. Cellular immune responses of older adults to four influenza vaccines: results of a randomized, controlled comparison. *Hum Vaccin Immunother*. 2017;13(9):2048-57.
77. Arrington J, Braun RP, Dong L, Fuller DH, Macklin MD, Umlauf SW, et al. Plasmid vectors encoding cholera toxin

- or the heat-labile enterotoxin from *Escherichia coli* are strong adjuvants for DNA vaccines. *J Virol.* 2002;76(9):4536-46.
78. Liu MA. A comparison of plasmid DNA and mrna as vaccine technologies. *Vaccines.* 2019;7(2):37.
 79. Azizi H, Kazemi B, Bandehpour M, Mohebbali M, Khamesipour A, Aryaeipour M, et al. Modulation of the Immune Response to DNA Vaccine Encoding Gene of 8-kDa Subunit of *Echinococcus granulosus* Antigen B Using Murine Interleukin-12 Plasmid in BALB/c Mice. *Iran J Parasitol.* 2016;11(4):480-9.
 80. Johansson P, Lindgren T, Lundström M, Holmström A, Elgh F, Bucht G. PCR-generated linear DNA fragments utilized as a hantavirus DNA vaccine. *Vaccine.* 2002;20(27-28):3379-88.
 81. Schmeer M, Buchholz T, Schleef M. Plasmid DNA manufacturing for indirect and direct clinical applications. *Human Gene Ther.* 2017;28(10):856-61.
 82. Wu M, Zhao H, Li M, Yue Y, Xiong S, Xu W. Intranasal vaccination with mannosylated chitosan formulated DNA vaccine enables robust IgA and cellular response induction in the lungs of mice and improves protection against pulmonary mycobacterial challenge. *Front Cell Infect Microbiol.* 2017;7(2):445-8.
 83. Modjarrad K, Roberts CC, Mills KT, Castellano AR, Paolino K, Muthumani K, et al. Safety and immunogenicity of an anti-Middle East respiratory syndrome coronavirus DNA vaccine: a phase 1, open-label, single-arm, dose-escalation trial. *Lancet Infect Dis.* 2019;19(9):1013-22.
 84. Bloom K, van den Berg F, Arbutnot P. Self-amplifying RNA vaccines for infectious diseases. *Gene Ther.* 2021;28(3-4):117-29.
 85. Jackson NA, Kester KE, Casimiro D, Gurunathan S, DeRosa F. The promise of mRNA vaccines: A biotech and industrial perspective. *NPJ Vaccines.* 2020;5(1):1-6.
 86. Sharifnia Z, Bandehpour M, Kazemi B, Zarghami N. Design and Development of Modified mRNA Encoding Core Antigen of Hepatitis C Virus: a Possible Application in Vaccine Production. *Iran Biomed J.* 2019;23(1):57-67.
 87. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *N Engl J Med.* 2021;384(5):403-16.
 88. Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. *N Engl J Med.* 2020;383(27):2603-15.
 89. Zhang N-N, Li X-F, Deng Y-Q, Zhao H, Huang Y-J, Yang G, et al. A thermostable mRNA vaccine against COVID-19. *Cell.* 2020;182(5):1271-83.
 90. Alberer M, Gnad-Vogt U, Hong HS, Mehr KT, Backert L, Finak G, et al. Safety and immunogenicity of a mRNA rabies vaccine in healthy adults: an open-label, non-randomised, prospective, first-in-human phase 1 clinical trial. *Lancet.* 2017;390(10101):1511-20.
 91. Fontanet A, Autran B, Lina B, Kieny MP, Karim SSA, Sridhar D. SARS-CoV-2 variants and ending the COVID-19 pandemic. *Lancet.* 2021;397(10278):952-4.
 92. Ura T, Okuda K, Shimada M. Developments in viral vector-based vaccines. *Vaccines.* 2014;2(3):624-41.
 93. Van Riel D, de Wit E. Next-generation vaccine platforms for COVID-19. *Nat Mater.* 2020;19(8):810-2.
 94. Sumida SM, Truitt DM, Lemckert AA, Vogels R, Custers JH, Addo MM, et al. Neutralizing antibodies to adenovirus serotype 5 vaccine vectors are directed primarily against the adenovirus hexon protein. *J Immunol.* 2005;174(11):7179-85.
 95. Yang Y, Nunes FA, Berencsi K, Furth EE, Gönczöl E, Wilson JM. Cellular immunity to viral antigens limits E1-deleted adenoviruses for gene therapy. *Proc Natl Acad Sci U S A.* 1994;91(10):4407-11.
 96. Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet.* 2021;397(10269):99-111.
 97. Logunov DY, Dolzhikova IV, Shcheblyakov DV, Tukhvatulin AI, Zubkova OV, Dzharullaeva AS, et al. Safety and efficacy of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine: an interim analysis of a randomised controlled phase 3 trial in Russia. *Lancet.* 2021;397(10275):671-81.
 98. Hu J, Ni Y, Dryman BA, Meng X, Zhang C. Immunogenicity study of plant-made oral subunit vaccine against porcine reproductive and respiratory syndrome virus (PRRSV). *Vaccine.* 2012;30(12):2068-74.
 99. Cassataro J, Pasquevich KA, Estein SM, Laplagne DA, Velikovskiy CA, de la Barrera S, et al. A recombinant subunit vaccine based on the insertion of 27 amino acids from Omp31 to the N-terminus of BLS induced a similar degree of protection against *B. ovis* than Rev.1 vaccination. *Vaccine.* 2007;25(22):4437-46.
 100. Rajan V. An Oral Vaccine for TGEV Immunization of Pigs. *Commercial Plant-Produced Recombinant Protein Products: Springer.* 2014; 135-52.

Pro and Cons of Coronavirus Vaccines

101. Daniell H. Production of biopharmaceuticals and vaccines in plants via the chloroplast genome. *Biotechnol J Healthcare Nutr Technol*. 2006;1(10):1071-9.
102. Cappel R. Comparison of the humoral and cellular immune responses after immunization with live, UV inactivated herpes simplex virus and a subunit vaccine and efficacy of these immunizations. *Arch Virol*. 1976;52(1-2):29-35.
103. Aucouturier J, Dupuis L, Ganne V. Adjuvants designed for veterinary and human vaccines. *Vaccine*. 2001;19(17-19):2666-72.
104. Nevagi RJ, Skwarczynski M, Toth I. Polymers for subunit vaccine delivery. *European Polymer Journal*. 2019;114(17):397-410.
105. Foged C, Rades T, Perrie Y, Hook S. Subunit vaccine delivery: Springer. 2015: XIV, 431.
106. Pankrac J, Klein K, McKay PF, King DF, Bain K, Knapp J, et al. A heterogeneous human immunodeficiency virus-like particle (VLP) formulation produced by a novel vector system. *NPJ vaccines*. 2018;3(1):1-10.
107. Wang JW, Roden RB. Virus-like particles for the prevention of human papillomavirus-associated malignancies. *Expert Rev Vaccines*. 2013;12(2):129-41.
108. Dai S, Wang H, Deng F. Advances and challenges in enveloped virus-like particle (VLP)-based vaccines. *J Immunol Sci*. 2018;2(2):36-41.
109. Mohsen MO, Zha L, Cabral-Miranda G, Bachmann MF, editors. Major findings and recent advances in virus-like particle (VLP)-based vaccines. *Semin Immunol*. 2017;34(8):123-32.
110. Madrid-Marina V, Torres-Poveda K, López-Toledo G, García-Carrancá A. Advantages and Disadvantages of Current Prophylactic Vaccines Against HPV. *Arch Med Res*. 2009;40(6):471-7.
111. Zhang Y, Zeng G, Pan H, Li C, Kan B, Hu Y, et al. Immunogenicity and safety of a SARS-CoV-2 inactivated vaccine in healthy adults aged 18-59 years: report of the randomized, double-blind, and placebo-controlled phase 2 clinical trial. *Med Rxiv*. 2020;(31):20161216.
112. Xia S, Duan K, Zhang Y, Zhao D, Zhang H, Xie Z, et al. Effect of an Inactivated Vaccine Against SARS-CoV-2 on Safety and Immunogenicity Outcomes: Interim Analysis of 2 Randomized Clinical Trials. *JAMA*. 2020;324(10):951-60.
113. Dey A, Chozhavel Rajanathan TM, Chandra H, Pericherla HPR, Kumar S, Choonia HS, et al. Immunogenic Potential of DNA Vaccine candidate, ZyCoV-D against SARS-CoV-2 in Animal Models. *bioRxiv*. 2021;1(26):428240-8.
114. Jackson LA, Anderson EJ, Roupael NG, Roberts PC, Makhene M, Coler RN, et al. An mRNA Vaccine against SARS-CoV-2—preliminary report. *N Eng J Med*. 2020;383(25):1920-31.
115. Mulligan MJ, Lyke KE, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Phase I/II study of COVID-19 RNA vaccine BNT162b1 in adults. *Nature*. 2020;586(7830):589-93.
116. Zhang N-N, Li X-F, Deng Y-Q, Zhao H, Huang Y-J, Yang G, et al. A Thermostable mRNA Vaccine against COVID-19. *Cell*. 2020;182(5):1271-83.e16.
117. Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S, et al. Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial. *Lancet*. 2020;396(10249):467-78.
118. Van Doremalen N, Lambe T, Spencer A, Belij-Rammerstorfer S, Purushotham JN, Port JR, et al. ChAdOx1 nCoV-19 vaccine prevents SARS-CoV-2 pneumonia in rhesus macaques. *Nature*. 2020;586(7830):578-82.
119. Logunov DY, Dolzikhova IV, Zubkova OV, Tukhvatullin AI, Shcheblyakov DV, Dzharullaeva AS, et al. Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia. *Lancet*. 2020;396(10255):887-97.
120. Sadoff J, Gray G, Vandebosch A, Cárdenas V, Shukarev G, Grinsztejn B, et al. Safety and Efficacy of Single-Dose Ad26.COV2.S Vaccine against Covid-19. *New Eng J Med*. 2021;384(23):2187-201.
121. Mahase E. Covid-19: Novavax vaccine efficacy is 86% against UK variant and 60% against South African variant. *BMJ*. 2021;372:n296.
122. Ward BJ, Gobeil P, Séguin A, Atkins J, Boulay I, Charbonneau P-Y, et al. Phase 1 trial of a Candidate Recombinant Virus-Like Particle Vaccine for Covid-19 Disease Produced in Plants. *Med Rxiv*. 2020;11(4):20226282.
123. Wareing M, Tannock G. Live attenuated vaccines against influenza; an historical review. *Vaccine*. 2001;19(25-26):3320-30.
124. Ni L, Seward JF, Santibanez TA, Pallansch MA, Kew OM, Prevots DR, et al. Vaccine policy changes and epidemiology of poliomyelitis in the United States. *JAMA*. 2004;292(14):1696-701.

125. Smith NM, Bresee JS, Shay DK, Uyeki TM, Cox NJ, Strikas RA, et al. Prevention and control of influenza: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep.* 2006;55(RR-10):1-42.
126. Wharton M, Strikas R, Harpaz R, Rotz L, Schwartz B, Casey C. Advisory Committee on Immunization Practices; Healthcare Infection Control Practices Advisory Committee: Recommendations for using smallpox vaccine in a pre-event vaccination program. Supplemental recommendations of the Advisory Committee on Immunization Practices (ACIP) and the Healthcare Infection Control Practices. Advisory Committee (HICPAC). *MMWR Recomm Rep.* 2003;52(RR-7):1-16.
127. Watson J, Hadler S, Dykewicz C, Reef S, Phillips L. Vaccine use and strategies for elimination of measles, rubella, and congenital rubella syndrome and control of mumps: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep.* 1998;47(8):1-57.
128. Draft Landscape of COVID-19 Candidate Vaccines. <https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines>.
129. Wang H, Zhang Y, Huang B, Deng W, Quan Y, Wang W, et al. Development of an inactivated vaccine candidate, BBIBP-CorV, with potent protection against SARS-CoV-2. *Cell.* 2020;182(3):713-21.
130. Gao Q, Bao L, Mao H, Wang L, Xu K, Yang M, et al. Development of an inactivated vaccine candidate for SARS-CoV-2. *Science.* 2020;369(6499):77-81.
131. Stauffer F, El-Bacha T, Da Poian AT. Advances in the development of inactivated virus vaccines. *Recent Pat Antiinfect Drug Discov.* 2006;1(3):291-6.
132. Barrett PN, Mundt W, Kistner O, Howard MK. Vero cell platform in vaccine production: moving towards cell culture-based viral vaccines. *Expert Rev Vaccines.* 2009;8(5):607-18.
133. Huckriede A, Bungener L, Stegmann T, Daemen T, Medema J, Palache AM, et al. The virosome concept for influenza vaccines. *Vaccine.* 2005;23:S26-S38.
134. Zhang Y, Zeng G, Pan H, Li C, Hu Y, Chu K, et al. Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial. *Lancet Infect Dis.* 2021;21(2):181-92.
135. Barrett PN, Mundt W, Kistner O, Howard MK. Vero cell platform in vaccine production: moving towards cell culture-based viral vaccines. *Expert Rev Vaccines.* 2009;8(5):607-18.
136. Smith TR, Schultheis K, Broderick KE. Nucleic acid-based vaccines targeting respiratory syncytial virus: Delivering the goods. *Hum Vaccin Immunother.* 2017;13(11):2626-9.
137. Cui Z. DNA vaccine. *Adv Genet.* 2005;54:257-89.
138. Authority EFS, Houston R, Moxon S, Nogu e F, Papadopoulou N, Ramon M, et al. Assessment of the potential integration of the DNA plasmid vaccine CLYNAV into the salmon genome. *EFSA J.* 2017;15(1):e04689.
139. Nichols WW, Ledwith BJ, Manam SV, Troilo PJ. Potential DNA vaccine integration into host cell genome. *Ann N Y Acad Sci.* 1995;772:30-9.
140. Abdulhaqq SA, Weiner DB. DNA vaccines: developing new strategies to enhance immune responses. *Immunol Res.* 2008;42(1-3):219-32.
141. Gottlieb P, Utz PJ, Robinson W, Steinman L. Clinical optimization of antigen specific modulation of type 1 diabetes with the plasmid DNA platform. *Clinic Immunol.* 2013;149(3):297-306.
142. Edwards DK, Jasny E, Yoon H, Horscroft N, Schanen B, Geter T, et al. Adjuvant effects of a sequence-engineered mRNA vaccine: translational profiling demonstrates similar human and murine innate response. *J Transl Med.* 2017;15(1):1-18.
143. Fuller DH, Berglund P. Amplifying RNA vaccine development. *N Engl J Med.* 2020;382(25):2469-71.
144. Zhang L, Bai J, Liu J, Wang X, Li Y, Jiang P. Toll-like receptor ligands enhance the protective effects of vaccination against porcine reproductive and respiratory syndrome virus in swine. *Vet Microbiol.* 2013;164(3-4):253-60.
145. Reautschnig P, Vogel P, Stafforst T. The notorious RNA in the spotlight-drug or target for the treatment of disease. *RNA biology.* 2017;14(5):651-68.
146. Douville RN, Nath A. Human endogenous retroviruses and the nervous system. *Handb Clin Neurol.* 2014;123(12):465-85.
147. Campbell JD. Development of the CpG adjuvant 1018: a case study. *Methods Mol Biol.* 2017;1494(26):15-27.
148. Blakney AK, McKay PF, Yus BI, Aldon Y, Shattock RJ. Inside out: optimization of lipid nanoparticle formulations for exterior complexation and in vivo delivery of saRNA. *Gene Ther.* 2019;26(9):363-72.
149. Sekaly R-P. The failed HIV Merck vaccine study: a step back or a launching point for future vaccine development? *J Exp Med.* 2008;205(1):7-12.

Pro and Cons of Coronavirus Vaccines

150. Rerks-Ngarm S, Pitisuttithum P, Nitayaphan S, Kaewkungwal J, Chiu J, Paris R, et al. Vaccination with ALVAC and AIDSVAX to prevent HIV-1 infection in Thailand. *N Engl J Med*. 2009;361(23):2209-20.
151. Dicks MDJ, Spencer AJ, Edwards NJ, Wadell G, Bojang K, Gilbert SC, et al. A Novel Chimpanzee Adenovirus Vector with Low Human Seroprevalence: Improved Systems for Vector Derivation and Comparative Immunogenicity. *PLOS ONE*. 2012;7(7):e40385.
152. Watkins DI, Burton DR, Kallas EG, Moore JP, Koff WC. Nonhuman primate models and the failure of the Merck HIV-1 vaccine in humans. *Nat Med*. 2008;14(6):617-21.
153. Priddy FH, Brown D, Kublin J, Monahan K, Wright DP, Lalezari J, et al. Safety and immunogenicity of a replication-incompetent adenovirus type 5 HIV-1 clade B gag/pol/nef vaccine in healthy adults. *Clin Infect Dis*. 2008;46(11):1769-81.
154. Logunov DY, Dolzhikova IV, Zubkova OV, Tukhvatullin AI, Shcheblyakov DV, Dzharullaeva AS, et al. Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia. *Lancet*. 2020;396(10255):887-97.
155. Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S, et al. Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial. *Lancet*. 2020;396(10249):467-78.
156. Moyle PM, Toth I. Modern subunit vaccines: development, components, and research opportunities. *ChemMedChem*. 2013;8(3):360-76.
157. Karch CP, Burkhard P. Vaccine technologies: From whole organisms to rationally designed protein assemblies. *Biochem Pharmacol*. 2016;120(9):1-14.
158. Eyigün CP, Yilmaz S, Gül C, Sengül A, Hacibektasoglu A, Van Thiel DH. A comparative trial of two surface subunit recombinant hepatitis B vaccines vs a surface and PreS subunit vaccine for immunization of healthy adults. *J Viral Hepat*. 1998;5(4):265-9.
159. Feenstra F, Van Rijn PA. Current and next-generation bluetongue vaccines: Requirements, strategies, and prospects for different field situations. *Crit Rev Microbiol*. 2017;43(2):142-55.
160. Keech C, Albert G, Reed P, Neal S, Plested JS, Zhu M, et al. First-in-Human Trial of a SARS CoV 2 Recombinant Spike Protein Nanoparticle Vaccine. *MedRxiv*. 2020;8(5):20168435.
161. Lövgren K, Morein B. The requirement of lipids for the formation of immunostimulating complexes (iscoms). *Biotechnol Appl Biochem*. 1988;10(2):161-72.
162. Magnusson SE, Altenburg AF, Bengtsson KL, Bosman F, de Vries RD, Rimmelzwaan GF, et al. Matrix-M™ adjuvant enhances immunogenicity of both protein- and modified vaccinia virus Ankara-based influenza vaccines in mice. *Immunol Res*. 2018;66(2):224-33.
163. Bachmann MF, Jennings GT. Vaccine delivery: a matter of size, geometry, kinetics and molecular patterns. *Nat Rev Immunol*. 2010;10(11):787-96.
164. Grgacic EVL, Anderson DA. Virus-like particles: Passport to immune recognition. *Methods*. 2006;40(1):60-5.
165. Andrus JK, Sherris J, Fitzsimmons JW, Kane MA, Aguado MT. Introduction of human papillomavirus vaccines into developing countries - international strategies for funding and procurement. *Vaccine*. 2008;26(Suppl 10):K87-92.
166. Garçon N, Vaughn DW, Didierlaurent AM. Development and evaluation of AS03, an Adjuvant System containing α -tocopherol and squalene in an oil-in-water emulsion. *Expert Rev Vaccines*. 2012;11(3):349-66.
167. Castells MC, Phillips EJ. Maintaining Safety with SARS-CoV-2 Vaccines. *N Engl J Med*. 2020;384(7):643-9.
168. Stone Jr CA, Rukasin CR, Beachkofsky TM, Phillips EJ. Immune-mediated adverse reactions to vaccines. *Br J Clin Pharmacol*. 2019;85(12):2694-706.
169. Kaur RJ, Dutta S, Bhardwaj P, Charan J, Dhingra S, Mitra P, et al. Adverse Events Reported From COVID-19 Vaccine Trials: A Systematic Review. *Indian J Clin Biochem*. 2021;36(4):1-13.
170. Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet*. 2021;397(10269):99-111.
171. Logunov DY, Dolzhikova IV, Zubkova OV, Tukhvatullin AI, Shcheblyakov DV, Dzharullaeva AS, et al. Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia. *Lancet*. 2020;396(10255):887-97.
172. Sadoff J, Gars ML, Shukarev G, Heerwegh D, Truyers C, de Groot AM, et al. Safety and immunogenicity of the Ad26.COV2.S COVID-19 vaccine candidate: interim results of a phase 1/2a, double-blind, randomized, placebo-controlled trial. *MedRxiv*. 2020;9(23):20199604.

173. Keech C, Albert G, Cho I, Robertson A, Reed P, Neal S, et al. Phase 1-2 Trial of a SARS-CoV-2 Recombinant Spike Protein Nanoparticle Vaccine. *N Engl J Med*. 2020;383(24):2320-32.
174. Ella R, Vadrevu KM, Jogdand H, Prasad S, Reddy S, Sarangi V, et al. A Phase 1: Safety and Immunogenicity Trial of an Inactivated SARS-CoV-2 Vaccine-BBV152. *MedRxiv*. 2020;12(11):20210419.
175. <https://www.cdc.gov/coronavirus/2019-ncov/cases-updates/variant-surveillance/variant-info.html>.
176. Abdool Karim SS, de Oliveira T. New SARS-CoV-2 Variants — Clinical, Public Health, and Vaccine Implications. *N Engl J Med*. 2021;384(19):1866-8.
177. Davies NG, Abbott S, Barnard RC, Jarvis CI, Kucharski AJ, Munday JD, et al. Estimated transmissibility and impact of SARS-CoV-2 lineage B.1.1.7 in England. *MedRxiv*. 2020;12(24):20248822.
178. Davies NG, Jarvis CI, CMMID COVID-19 Working Group, et al. Increased mortality in community-tested cases of SARS-CoV-2 lineage B.1.1.7. *Nature*. 2021;593:270-4.
179. Reuschl AK, Thorne L, Zuliani Alvarez L, Bouhaddou M, Obernier K, Soucheray M, et al. Host-directed therapies against early-lineage SARS-CoV-2 retain efficacy against B.1.1.7 variant. *bioRxiv*. 2021;24:427991.
180. Wang P, Nair MS, Liu L, Iketani S, Luo Y, Guo Y, et al. Antibody Resistance of SARS-CoV-2 Variants B.1.351 and B.1.1.7. *bioRxiv*. 2021;1(25):428137.
181. Deng X, Garcia-Knight MA, Khalid MM, Servellita V, Wang C, Morris MK, et al. Transmission, infectivity, and antibody neutralization of an emerging SARS-CoV-2 variant in California carrying a L452R spike protein mutation. *MedRxiv*. 2021;3(7):21252647.
182. Jangra S, Ye C, Rathnasinghe R, Stadlbauer D, Alshammery H, Amoako AA, et al. SARS-CoV-2 spike E484K mutation reduces antibody neutralisation. *Lancet Microbe*. 2021;2(7):e283-e284.
183. Yadav PD, Sapkal GN, Abraham P, Ella R, Deshpande G, Patil DY, et al. Neutralization of variant under investigation B.1.617 with sera of BBV152 vaccinees. *BioRxiv*. 2021;4(23):441101.
184. Gupta RK. Will SARS-CoV-2 variants of concern affect the promise of vaccines? *Nat Rev Immunol*. 2021;21(8):340–341.
185. Kemp S, Harvey W, Datir R, Collier D, Ferreira I, Carabeli A, et al. Recurrent emergence and transmission of a SARS-CoV-2 Spike deletion Δ H69/V70. *bioRxiv*. 2020;2020.12.14.422555.
186. Collier DA, De Marco A, Ferreira IA, Meng B, Datir RP, Walls AC, et al. Sensitivity of SARS-CoV-2 B. 1.1. 7 to mRNA vaccine-elicited antibodies. *Nature*. 2021:1-10.
187. Wang P, Casner RG, Nair MS, Wang M, Yu J, Cerutti G, et al. Increased Resistance of SARS-CoV-2 Variant P. 1 to Antibody Neutralization. *Cell Host Microbe*. 2021:2021.03.01.433466.
188. Hon C-C, Lam T-Y, Shi Z-L, Drummond AJ, Yip C-W, Zeng F, et al. Evidence of the recombinant origin of a bat severe acute respiratory syndrome (SARS)-like coronavirus and its implications on the direct ancestor of SARS coronavirus. *J Virol*. 2008;82(4):1819-26.
189. Cerutti G, Guo Y, Zhou T, Gorman J, Lee M, Rapp M, et al. Potent SARS-CoV-2 neutralizing antibodies directed against spike N-terminal domain target a single supersite. *Cell Host Microbe*. 2021;29(5):819-33.
190. Fratev F. The N501Y and K417N mutations in the spike protein of SARS-CoV-2 alter the interactions with both hACE2 and human derived antibody: A Free energy of perturbation study. *bioRxiv*. 2020;2020.12.23.424283.
191. Greaney AJ, Starr TN, Gilchuk P, Zost SJ, Binshtein E, Loes AN, et al. Complete mapping of mutations to the SARS-CoV-2 spike receptor-binding domain that escape antibody recognition. *Cell host & microbe*. 2021;29(1):44-57.
192. Bian L, Gao F, Zhang J, He Q, Mao Q, Xu M, et al. Effects of SARS-CoV-2 variants on vaccine efficacy and response strategies. *Expert review of vaccines*. 2021:1-9.
193. Tchesnokova V, Kulakesara H, Larson L, Bowers V, Rechkina E, Kisiela D, et al. Acquisition of the L452R mutation in the ACE2-binding interface of Spike protein triggers recent massive expansion of SARS-Cov-2 variants. *bioRxiv*. 2021:2021.02.22.432189.
194. Mohammadi E, Shafiee F, Shahzamani K, Ranjbar MM, Alibakhshi A, Ahangarzadeh S, et al. Novel and emerging mutations of SARS-CoV-2: Biomedical implications. *Biomed Pharmacother*. 2021;139:111599.
195. Shinde V, Bhikha S, Hossain Z, Archary M, Borhat Q, Fairlie L, et al. Preliminary efficacy of the NVX-CoV2373 Covid-19 vaccine against the B. 1.351 variant. *med Rxiv*. 2021:2021.02.25.21252477.
196. Madhi SA, Baillie V, Cutland CL, Voysey M, Koen AL, Fairlie L, et al. Efficacy of the ChAdOx1 nCoV-19 Covid-19 vaccine against the B. 1.351 variant. *N Engl J Med*. 2021;384(20):1885-98.
197. Cohen J. South Africa suspends use of AstraZeneca's COVID-19 vaccine after it fails to clearly stop virus variant. *Science*. 2021:2-7.

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198. Wadman M, Cohen J. Novavax vaccine delivers 89% efficacy against COVID-19 in UK—but is less potent in South Africa. *Science*. 2021;12(2774).
199. Hitchings M, Ranzani OT, Torres MS, de Oliveira SB, Almiron M, Said R, et al. Effectiveness of CoronaVac in the setting of high SARS-CoV-2 P. 1 variant transmission in Brazil: A test-negative case-control study. *medRxiv*. 2021:2021.04.07.21255081.
200. Randolph HE, Barreiro LB. Herd Immunity: Understanding COVID-19. *Immunity*. 2020;52(5):737-41.